

MAHATMA GANDHI UNIVERSITY Priyadarshini Hills, Kottayam 686 560

CURRICULUM OF CHOICE BASED CREDIT SYSTEM FOR UNDERGRADUATE

ZOOLOGY PROGRAMME

2017 ADMISSION ONWARDS

PREPARED BY BOARD OF STUDIES AND FACULTY OF SCIENCE

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The unstinted support provided by some profound personalities and their guidance has immensely contributed to the successful completion of this syllabus restructuring venture.

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Any venture of this magnitude requires the whole hearted support of several individuals in their official capacity or otherwise, directly or indirectly associated with the process. The magnanimity of each and every such person is being etched here in golden words.

Dean – Faculty of Science M G University, Kottayam.

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1. INTRODUCTION

Curriculum restructuring is the responsibility of any University that offers different courses. As per U.G.C guidelines it should be revised every three years. Accordingly the authorities of M.G University have taken the initiative to restructure the syllabus of each course for the benefit of the students.

According to Altman and Cashin "the primary purpose of a syllabus is to communicate to one's students what the course is about, why the course is taught, where it is going, and what will be required of the students, for them to complete the course with a passing grade. Claire Johnson says "the syllabus functions as a major communication device that provides details of how student learning will be assessed and about the roles of both Students and Instructors in the learning and assessment process. The syllabus is a "description" and "plan" for a course and if well written may be a tool that improves student learning, facilitates faculty teaching, improves communication between faculty members about their courses and assists with monitoring program quality says Claire Johnson (2006). Additionally Parke,Fix and Harris suggest that " the syllabus serves as a contract between the Instructor and the learner".

The M.G. University is well recognised among Universities for their quickness in updating technologies. It is the recipient of the award for computerization among the Universities. Ever since its inception, the M.G.University has had the history of updating its syllabi for the various courses offered. Presently, this exercise has resulted in the syllabus of Zoology, for the undergraduate students undergoing model I, II and III courses, reaching successful updation.

India, a mega biodiversity nation stands out as a fascinating destination, exhibiting an incredible diversity in its fauna and flora. Innumerable species of organisms live here contributing to India's most beautiful natural topography.. All round development becomes possible only by exploring the vast potential of the still unknown resources of nature. Zoology undergraduate students are therefore made to go through the realities like Biota, anatomy and physiology of living organisms, DNA biotechnology, diseases, pathogens and parasites, environment, resources, protection of endangered organisms and their conservation, the "right to live" of all organisms in this world, and finally scientific knowledge about cottage industries, prompting students to involve in self employment programs.

The constant and unstinted efforts of the entire faculty of Zoology within M.G.University has been instrumental in framing this syllabus for the subject Zoology in this form. The faculty of Zoology proudly and sincerely submit herewith this updated syllabus in Zoology for its implementation in the coming three years.

2. AIMS & OBJECTIVES

The BSc. Zoology programme is designed to help the students to:

- 1. Acquire basic knowledge of various disciplines of Zoology and General Biology meant both for a graduate terminal course and for higher studies.
- 2. Inculcate interest in nature and love of nature.
- 3. Understand the rich diversity of organisms and their ecological and evolutionary significance
- 4. Imbibe basic skills in the observation and study of nature, biological techniques, experimental skills and scientific investigation
- 5. Create awareness on the internal harmony of different body systems and the need for maintaining good health through appropriate lifestyle.
- 6. Acquire basic knowledge and skills in certain applied branches for self employment
- 7. Impart awareness of the conservation of the biosphere.

3. DURATION OF THE COURSE

The duration of U.G. programmes have 6 semesters. There shall be two Semesters in an academic year, the odd semester commences in June and on completion, the even Semester commences after a semester-break of one or two days with two months vacation during April and May. (The commencement of first semester may be delayed owing to the finalization of the admission processes.)

4. PROGRAMME STRUCTURE - B.Sc Model I

a	Programme Duration	6 Semesters
b	Total Credits required for successful completion of the Programme	120
c	Credits required from Common Course I	22
d	Credits required from Common Course II	16
e	Credits required from Core course and Complementary courses including Project	79
f	Open course	3
g	Minimum attendance required	75%

B.Sc Model II

А	Programme Duration	6 Semesters
В	Total Credits required for successful completion of the Programme	120
С	Credits required from Common Course I	16
D	Credits required from Common Course II	8
E	Credits required from Core + Complementary + Vocational Courses including Project	93
F	Open course	3
G	Minimum attendance required	75%

B.Sc Model III

А	Programme Duration	6 Semesters
В	Total Credits required for successful completion of the Programme	120
С	Credits required from Common Course I	8
D	Credits required from Core + Complementary + Vocational Courses including Project	109
Е	Open course	3
F	Minimum attendance required	75%

5. EVALUATIONS:.

The evaluation of each paper shall contain two parts:

- (i) Internal Assessment
- (ii) External Assessment

The internal to external assessment ratio shall be 1:4.

EXTERNAL EXAMINATION AND INTERNAL EVALUATION

The external theory examination of all semesters shall be conducted by the University at the end of each semester. Internal evaluation is to be done by continuous assessment.

ASSIGNMENTS

At least one assignment should be done in each semester for all papers.

INTERNAL ASSESSMENT TEST PAPERS

The test papers & evaluations of all components are to be published and are to be acknowledged by the candidates. All documents of internal assessments are to be kept in the college for two years and shall be made available for verification by the University. The responsibility of evaluating the internal assessment is vested on the teacher(s), who teach the paper.

EXTERNAL EXAMINATION

The external theory examination of all semesters shall be conducted by the University at the end of each semester. Students having a minimum of 75% average attendance for all the courses only can register for the examination. Condonation of shortage of attendance to a maximum of 10 days in a semester subject to a maximum of 2 times during the whole period of the programme may be granted by the University on valid grounds. This condonation shall not be counted for internal assessment. Benefit of attendance may be granted to students attending University/College union/Co-curricular activities by treating them as present for the days of absence, on production of participation/attendance certificates, within one week, from competent authorities and endorsed by the Head of the institution. This is limited to a maximum of 10 days per semester and this benefit shall be considered for internal assessment also. Those students who are not eligible even with condonation of shortage of attendance shall repeat the **semester** along with the next batch after obtaining readmission.

All students are to do a **project in the area of core course.** This project can be done individually or in groups (not more than three students) which may be carried out in or outside the campus. Special sanction shall be obtained from the Vice Chancellor to those **new generation programmes** and programmes related to projects which involve larger groups. The projects are to be identified during the fifth semester of the programme with the help of the supervising teacher. The report of the project in duplicate is to be submitted to the department at the sixth semester and are to be produced before the examiners appointed by the University. External Project evaluation and Viva / Presentation is compulsory for all subjects and will be conducted at the end of the programme. There will be no supplementary exams. For reappearance/ improvement, the students can appear along with the next batch.

6. CONSOLIDATED SCHEME FOR BSC ZOOLOGY PROGRAMME MODEL-1 (1-VI SEMESTERS) (TOTAL CREDITS 120)

(TOTAL HOURS 150 Hrs.)

	Course	Course	Course	Hrs per	Credits	Marks ratio	
No	Code	Course Title	Category	week		Intl	Extl
1		English - 1	Common Course I	5	4	1	4
2		English - 2	Common Course II	4	3	1	4
3		Second Language – 1	Common Course III	4	4	1	4
4	ZY1CRT01	General perspectives in Science & Protistan Diversity	Core Course I : Theory	2	2	1	4
5		Practical : General perspectives in Science & Protistan Diversity	Core Course I : Practical	2	0	0	0
6		Chemistry 1/ Biochemistry 1	Complemen tary Course I : Theory	2	2	1	4
7		Chemistry 1	Complemen tary Course I : Practical	2	0	0	0
8		Botany 1	Complemen tary Course II : Theory	2	2	1	4
9		Botany 1	Complemen tary Course II : Practical	2	0	0	0
			25 hrs	17			

	Course				Credi	Marks ratio		
No	Code	Course Title	Course Category	week	ts	Intl	Extl	
1		English 3	Common Course IV	5	4	1	4	
2		English 4	Common Course V	4	3	1	4	

3		Second Language -2	Common Course VI	4	4	1	4
4	ZY2CRT02	Animal Diversity- Non Chordata	Core Course II : Theory	2	2	1	4
5		Animal Diversity – Non Chordata	Core Course II : Practical	2	2	1	4
6		Chemistry – II / Biochemistry - II	Complementary Course I : Theory	2	2	1	4
7		Chemistry – II / Biochemistry - II	Complementary Course I : Practical	2	2	1	4
8		Botany – II	Complementary Course II : Theory	2	2	1	4
9		Botany – II	Complementary Course II : Practical	2	2	1	4
	Total				23		

	Course		Course Hrs per			Mar	rks ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1		English 5	Common Course VII	5	4	1	4
2		Second Language 3	Common Course VIII	5	4	1	4
3	ZY3CRT03	Animal Diversity – Chordata	Core Course III : Theory	3	3	1	4
4		Animal Diversity- Chordata	Core Course III : Practical	2	0	0	0
5		Chemistry – III / Biochemistry - III	Complementary Course I : Theory	3	3	1	4
6		Chemistry – III / Biochemistry - III	Complementary Course I : Practical	2	0	0	0
7		Botany III	Complementary Course II : Theory	3	3	1	4
8		Botany III	Complementary Course II :	2	0	0	0

			Practical			
Total			25 hrs	17		

	Course			Hrs per		Marks	ratio
No	Code	Course Title	Course Category	week	Credits	Intl	Extl
1		English - 6	Common Course IX	5	4	1	4
2		Second language - 4	Common Course X	5	4	1	4
3	ZY4CRT04	Research methodology, Biophysics & Biostatistics	Core Course IV : Theory	3	3	1	4
4		Research methodology, Biophysics & Biostatistics	Core Course IV : Practical	2	2	1	4
5		Chemistry – IV / Biochemistry - IV	Complementary Course I : Theory	3	3	1	4
6		Chemistry – IV / Biochemistry - IV	Complementary Course I : Practical	2	2	1	4
7		Botany - IV	Complementary Course II : Theory	3	3	1	4
8		Botany - IV	Complementary Course II : Practical	2	2	1	4
			25 hrs	23			

	Course		Course	Hrs		Marl	ks ratio
No	Code	Course Title	Category	per week	Credits	Intl	Extl
1	ZY5CRT05	Environmental Biology & Human rights	Core Course V : Theory	3	3	1	4
2	ZY5CRPO5	Environmental Biology & Human rights	Core Course V : Practical	2	0	0	0
3	ZY5CRT06	Cell Biology & Genetics	Core Course VI : Theory	3	3	1	4
4	ZY5CRPO6	Cell Biology & Genetics	Core Course VI : Practical	2	0	0	0
5	ZY5CRT07	Evolution, Ethology & Core Course Zoogeography VII : Theory		3	3	1	4
6	ZY5CRPO7	Evolution, Ethology & Zoogeography	Core Course VII : Practical	2	0	0	0
7	ZY5CRT08	Human Physiology, Biochemistry & Endocrinology	Core Course VIII : Theory	3	3	1	4
8	ZY5CRPO8	Human Physiology, Biochemistry & Endocrinology	Core Course VIII : Practical	2	0	0	0
9	ZY5OPT01	1 – Vocational Zoology (Apiculture, Vermiculture, Ornamental fish culture)	Open Courses for other streams (<i>Select</i>	4	3	1	4
	ZY5OPT02	2 – Public health and Nutrition	any one out of three)				
	ZY5OPT03	3 – Man, nature & Sustainable Development					
10	ZY6CRPRP	Project work (Credit 2 will be given in 6 th semester with investigatory project).		1	0		
		Total		25 hrs	15		

	Course	Course	Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1	ZY6CRT09	Developmental Biology	Core Course IX : Theory	3	3	1	4
2	ZY6CRP09	Developmental Biology	Core Course IX : Practical	2	2	1	4
3	ZY6CRT10	Microbiology & Immunology	Core Course X : Theory	3	3	1	4
4	ZY6CRP10	Microbiology & Immunology	Core Course X : Practical	2	2	1	4
5	ZY6CRT11	Biotechnology, Bioinformatics and Molecular Biology	Core Course XI : Theory	3	3	1	4
6	ZY6CRP11	Biotechnology, Bioinformatics and Molecular Biology	Core Course XI : Practical	2	2	1	4
7	ZY6CRT12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course XII : Theory	3	3	1	4
8	ZY6CRP12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course XII : Practical	2	2	1	4
	ZY6CBT01	Elective 1: Ecotourism & Sustainable Development					
	ZY6CBT02	Elective 2: Agricultural pest management	Choice Based Core Elective				
9	ZY6CBT03	Elective 3: Vector & Vector borne Diseases	Courses (Select any one out of	4	3	1	4
	ZY6CBT04	Elective 4: Nutrition, Health & life style management	four)				
10	ZY6PRP01	Project Work	Project	1	2		
		Total		25 hrs	25		

7. SCHEME OF DISTRIBUTION OF INSTRUCTIONAL HOURS

Name of semester	Theory	Practical
First semester	2	2
Second semester	2	2
Third semester	3	2
Fourth semester	3	2
Fifth semester	16	8
Field study & group activity	1	
Sixth semester	16	8
Project work (in sixth sem.), Visit to research institute	1	
Total	44	24

B.Sc. ZOOLOGY PROGRAMME : CORE COURSES -MODEL I

RECORDS

- 1. General perspectives in Science & Protistan Diversity
- 2. Animal Diversity –Non Chordata
- 3. Animal Diversity Chordata
- 4. Research Methodology, Biophysics and Biostatistics
- 5. Environmental Biology & Human rights
- 6. Cell Biology & Genetics
- 7. Evolution and Ethology & Zoogeography
- 8. Human Physiology, Biochemistry & Endocrinology
- 9. Developmental Biology
- 10. Microbiology and Immunology
- 11. Biotechnology, Bioinformatics & Molecular Biology
- 12. Occupational Zoology

Each Record will be having external and internal evaluation. A total of 1 credit will be allotted for each record and respective practical.

FIELD STUDY/ (STUDY TOUR)

Study tour/ field study, visit to research institute and various places of zoological Importance

A study tour is compulsory. Field study/study tours should be conducted for not less than 6 days (completed during the entire programme), preferably spreading the study in the first to sixth semesters. Students are expected to visit research institutes and various places of zoological importance.

GROUP ACTIVITY

Students are expected to do one group activity in the fifth semester and submit the report in the sixth semester for external practical examination, along with study tour report.

A maximum of ten students can choose any one group activity like aquarium management, vermicomposting, bee keeping, and conduct of zoological exhibitions, designing of posters of zoological importance, surveys related to disease outbreaks, community health programmes or any matter of zoological interest.

A brief report on any two activities mentioned in each elective paper should be submitted as a component of assignment for internal evaluation.

8. SCHEME OF DISTRIBUTION OF HOURS AND CREDIT

B.Sc. ZOOLOGY PROGRAMME : MODEL -1 CORE COURSE

Semester	Title of the Course		Inst Hrs/week	Credit
1	General perspectives in Science & Protistan Diversity	36	2	2
1	General perspectives in Science & Protistan diversity (Practical)	36	2	0
2	Animal Diversity – Non Chordata	36	2	2
2	Animal Diversity – Non Chordata (Practical)	36	2	2
3	Animal Diversity - Chordata	54	3	3
3	Animal Diversity - Chordata (Practical)	36	2	0
4	Research Methodology, Biophysics and Biostatistics	54	3	3
4	Research Methodology, Biophysics and Biostatistics (Practical)	36	2	2
5	Environmental Biology & Human rights	54	3	3
5	Environmental Biology & Human rights (Practical)	36	4	0
5	Cell Biology & Genetics	54	3	3
5	Cell Biology & Genetics (Practical)	36	2	0
5	Evolution, Ethology & Zoogeography	54	3	3
5	Evolution, Ethology & Zoogeography (Practical)	36	2	0
5	Human Physiology, Biochemistry & Endocrinology	54	3	3

5	Human Physiology, Biochemistry & Endocrinology (Practical)		2	0
6	Developmental Biology	54	3	3
6	Developmental Biology (Practical)	36	2	2
6	Microbiology & Immunology	54	3	3
6	Microbiology & Immunology (Practical)	36	2	2
6	Biotechnology, Bioinformatics & Molecular biology	54	3	3
6	Biotechnology, Bioinformatics &Molecular Biology (Practical)	36	4	2
6	Occupational Zoology (Aquaculture, Apiculture,Vermiculture & Quail farming)		3	3
6	Occupational Zoology(Aquaculture, Apiculture,Vermiculture & Quail farming) (Practical)	36	2	2

9. SCHEME OF DISTRIBUTION OF HOURS AND CREDITS :

OPEN COURSES (FOR OTHER STREAMS) & ELECTIVES

Semester	Course No	Title of the Course (Select any one from each group)	Hrs	Inst Hrs/ week	Cre dit
5	Open course – 1	Vocational Zoology (Apiculture,Vermiculture & Ornamental Fish Culture)			
5	Open course-2	Public Health & Nutrition			
5	Open course-3	Man, Nature & Sustainable Development	72	4	3
6	Choice Based Core Elective - 1	Ecotourism & Sustainable Development			
6	Choice Based Core Elective - 2	Agricultural Pest Management			
6	Choice Based Core Elective - 3	Vector & Vector Borne Diseases	72	4	3
6	Choice Based Core Elective - 4	Nutrition, Health & Life Style Management			

10. SCHEME OF EXAMINATIONS

a. SCHEME OF EXAMINATION - THEORY (CORE COURSE)

Theory Examinations will be conducted by the University at the end of the respective semester in which the course is conducted. Duration 3 Hrs (Internal External ratio =1:4)

			0	Duration	Mark	s ratio	
Sem	Course Code	Course Title	Course Category	Hrs	Internal (I)	External (E)	Credits
1	ZY1CRT01	General perspectives in Science & Protistan Diversity	1	2	1	4	2
2	ZY2CRT02	Animal Diversity – Non Chordata	2	2	1	4	2
3	ZY3CRT03	Animal Diversity- Chordata	3	3	1	4	3
4	ZY4CRT04	Research Methodology, Biophysics and Biostatistics	4	3	1	4	3
5	ZY5CRT05	Environmental Biology & Human rights	5	3	1	4	3
5	ZY5CRT06	Cell Biology & Genetics	6	3	1	4	3
5	ZY5CRT07	Evolution, Ethology & Zoogeography	7	3	1	4	3
5	ZY5CRT08	Human Physiology, Biochemistry & Endocrinology	8	3	1	4	3
6	ZY6CRT09	Developmental Biology	9	3	1	4	3
6	ZY6CRT10	Microbiology and Immunology	10	3	1	4	3
6	ZY6CRT11	Biotechnology, Bioinformatics & Molecular Biology	11	3	1	4	3
6	ZY6CRT12	Occupational Zoology (Aquaculture, Apiculture,Vermiculture	12	3	1	4	3

		& Quail farming)					
Zool ogy Open Cour	ZY5OPT01	Vocational Zoology (Apiculture, Vermiculture, & Ornamental Fish Culture)	Open course (for other streams)-1				
se Sem	ZY5OPT02	Public Health & Nutrition	Open course -2	4	1	4	3
5 (Sele ct any 1 from 3)	ZY5OPT03	Man, Nature & Sustainable Development	Open course -3				
	ZY6CBT01	Ecotourism & Sustainable Develpoment	Choice Based Course-1				
Sem 6 (Sele ct	ZY6CBT02	Agricultural Pest Management	Choice Based Course-2				
any 1 from the	ZY6CBT03	Vector & Vector Bourne Diseases	Choice Based Course-3	4	1	4	3
four)	ZY6CBT03	Nutrition,Health & Life Style Management	Choice Based Course -4				

b. SCHEME OF PRACTICAL EXAMINATIONS

University Practical Examinations will be conducted at the end of even semester 2, 4 and 6.

Sem	Cult	Practic	C	Duration	Marks	s ratio	Credit
e ster	Code	al No.	l No. Course Title Hrs	Internal (I)	Externa l (E)	S	
2	ZY2CRP01	Ι	General Perspectives in Science, Protistan Diversity & Animal Diversity – Non Chordata	3 Hrs	1	4	2
4	ZY4CRP02	Π	Animal Diversity – Chordata, Research methodology, Biophysics &	3 Hrs	1	4	2

			Biostatistics				
6	ZY6CRP03	III	Environmental Biology, Toxicology, Cell Biology & Genetics	3 Hrs	1	4	2
6	ZY6CRP04	IV	Evolution, Ethology, Zoogeography, Human Physiology, Biochemistry & Endocrinology	3 Hrs	1	4	2
6	ZY6CRP05	V	Developmental Biology, Microbiology & Immunology	3 Hrs	1	4	2
6	ZY6CRP06	VI	Biotechnology, Bioinformatics, Molecular Biology & Occupational Zoology	3 Hrs	1	4	2

c. Total Credits for Core Course

Theory	
Core + Choice Based Core (Elective)	37
Open course	3
Practical	12
Project	2
Total	54

d. SCHEME OF FIELD STUDY, RESEARCH INSTITUTE VISIT, GROUP ACTIVITY, PROJECT AND VIVA-CORE COURSE (Credit 2) Marks Maximum 100

	Internal Assessments (I)	External Assessments (E)
Project:- Log book showing the progress of project work duly signed by the supervising teacher & HOD	20	Project report – 50 marks Title- 2 Marks Abstract- 3 Marks Introduction & Review of literature- 10 Marks Methodology- 10 Marks Results- 10 Marks Discussion & Conclusion - 10 Marks Neat presentation and Novelty- 5 Marks Presentation & Viva- 30Marks (Student can present the project using OHP or LCD, in 7 Minutes) Viva Voce.
Total	20	80

e. ON THE JOB TRAINING PROGRAMME (OJ)

Each student is expected to do an 'on the job training programme' (OJ) outside the campus during the VI semester for Model II & Model III Programmes to acquire experimental skills, practical experience, scientific knowledge and also to equip themselves for self employment. The evaluation includes internal assessment (20 marks) and external assessment (80 marks). The external assessment includes the following components,

> Attendence – 10 marks Report submission – 40 marks Presentation & viva – 30 marks

11. COMPLEMENTARY ZOOLOGY COURSES OFFERED BY ZOOLOGY DEPARTMENT FOR - MODEL I – BSc BOTANY / BSc HOME SCIENCE MODEL II – BSc BOTANY / VOCATIONAL SUBJECTS MODEL III – BSc (BIOLOGICAL TECHNIQUES AND SPECIMEN PREPARATION)

Semester	Code	Title of the Course	Hrs	Inst Hrs/week	Credit
1	ZY1CMT01	Non Chordate Diversity	36	2	2
1		Non Chordate Diversity (Practicals)	36	2	0
2	ZY2CMT02	Chordate Diversity	36	2	2
2		Chordate Diversity (Practicals)	36	2	0
2	ZY2CMP01	Practical 1 Non Chordate Diversity + Chordate Diversity (Practicals)			2
3	ZY3CMT03	Physiology and Immunology	54	3	3
3		Physiology and Immunology (Practicals)	36	2	0
4	ZY4CMT04	Applied Zoology	54	3	3
		Applied Zoology (Practicals)	36	2	
4	ZY4CMP02	Practical 2 Physiology and Immunology + Applied Zoology (Practical)			2

12. SYLLABUS:

B.Sc ZOOLOGY PROGRAMME

MODEL – I

THEORY & PRACTICALS

SEMESTER 1. ZY1CRT0I. CORE COURSE 1.

GENERAL PERSPECTIVES IN SCIENCE & PROTISTAN DIVERSITY

Objectives:

- To create an awareness on the basic philosophy of science, concepts and scope
- To understand different levels of biological diversity through the systematic classification
- To familiarize taxa level identification of animals
- To make interest in Protistan diversity
- To impart knowledge on parasitic forms of lower invertebrates.

PART I PERSPECTIVES IN SCIENCE

Module I Introduction to Scientific Studies

Types of knowledge: practical, theoretical, and scientific knowledge. What is science, features of science, Deductive and inductive models, scientific temper, empiricism vocabulary of science.

Module II What is Biology?

Life and its manifestations, History of Biology: Biology in ancient times Landmarks in the progress of Biology. Branches of Zoology, Scope of Zoology

PART II SYSTEMATICS

Module III-Taxonomical Principles and tools

Systematic, Taxonomy, Phylogeny [Brief account], Approaches to taxonomy, Molecular taxonomy, .Bar coding. Zoological nomenclature, International Code of Zoological Nomenclature (ICZN), Law of Priority. Five Kingdom Classification; Linnaean classification, Basis for Animal kingdom classification [Levels of organization, Symmetry, Coelom]

Identification tools

10 Hrs

8Hrs

4 Hrs

Credits 2

36 Hrs

4Hrs

Taxonomic key. Types: Single access key- Dichotomous [linked and nested] and Polytomous key, Multi access key, Computer aided Interactive Key Advantages and Disadvantages

PART III: PROTISTAN DIVERSITY18 HrsModule IV – Kingdom Protista Type: Paramecium5 Hrs

Salient features of KingdomProtista

Classification of Protista up to phyla

1. Phylum Rhizopoda :Eg. Amoeba 2. Phylum Actinopoda : Eg. Actinophrys 3. Phylum Dinoflagellata : Eg. Noctiluca 4. Phylum Parabasalia : Eg. Trychonympha 5. Phylum Metamonada : Eg. Giardia 6. Phylum Kinetoplasta : Eg. Trypanosoma 7. Phylum Euglenophyta : Eg. Euglena 8. Phylum Cryptophyta : Eg. Cryptomonas 9. Phylum Opalinata : Eg. Opalina 10. Phylum Bacillariophyta :Eg. Diatoms 11. Phylum Chlorophyta :Eg. Volvox 12. Phylum Choanoflagellata :Eg. Proterospongia 13. Phylum Ciliophora : Eg. Balantidium coli 14. Phylum Sporozoa : Eg. Plasmodium 15. Phylum Microsporidia :Eg. Nosema

16. Phylum Rhodophyta :Eg. Red Alga

(Mention any five general characters for each phylum. Detailed accounts of examples are not necessary.)

General Topics:

 Parasitic protists (diseases mode of transmission and prophylactic measures) -Entamoeba, Trypanosoma, Plasmodium (detailed account of life cycle), Leishmania.

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3 Hrs

10 Hrs

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SEMESTER 1

CORE COURSE PAPER 1 PERSPECTIVES IN SCIENCE & PROTISTAN DIVERSITY

(PRACTICAL)

36 Hrs

2 Credits

- Taxa, identification techniques
 Bird body parts
 Butterfly/ dragonfly body parts
- Identification using keys
 Insect, Fish, Snake (Poisonous & Non Poisonous)
 (Any 3 specimens from each category)
- 3. General identification The students are expected to identify any 6 Protiatans studied by their generic names and write the general characters of their Phylum.
- 4. Identification of any 4 economically important protists/parasitic protists (Slides/figures may be used for identification)
- 5. Identification of two Protistan from pond water

SEMESTER 11. ZY2CRT02

CORE COURSE 11: ANIMAL DIVERSITY - NON CHORDATA

36 Hrs

Credits 2

Objectives:

- To create appreciation on diversity of life on earth
- To understand different levels of biological diversity through the systematic classification of invertebrate fauna
- To familiarize taxa level identification of animals
- To understand the evolutionary significance of invertebrate fauna
- To instill curiosity on invertebrates around us
- To impart knowledge on parasitic forms of lower invertebrates.

MODULE I Kingdom Animalia

Outline classification of Kingdom Animalia

Three branches - Mesozoa, parazoa and Eumetazoa

Mesozoa: Phylum Orthonectida - eg. Rhopalura (mention 5 salient features)

Parazoa:

1. Phylum Placozoa – Eg. *Trycoplax adherens*

2. Phylum Porifera – Classification upto classes; Mention gemmules

Class I- Calcarea. Eg.Sycon.,

Class II – Hexactinellida .Eg.Euplectella.

Class III - Demospongia Eg. Cliona.

General Topics

1. Canal system in sponges.

Phylum Coelenterata -Classification upto classes

Class I - Hydrozoa	Eg. Eg. Obelia - mention Metagenesis
Class II- Scyphozoa	Eg. Rhizostoma.
Class III- Anthozoa	Eg. Metridium.

General Topics:

1. Coral and coral reefs with special reference to conservation of reef fauna.

2. Polymorphism in Coelenterates

Phylum Ctenophora - Eg. Pleurobrachia.

MODULE II

Phylum Platyhelminthes Salient features; classification up to classes 3 Hrs

Class I - Turbellaria. Eg. *Planaria*.

Class II – Trematoda Eg. Fasciola

Class III- Cestoda Eg. Taenia saginata.

General Topics:

1. Life history of Fasciola hepatica.

2. Platyhelminth parasites of Man and Dog (Schistosoma, Taenia solium, Echinococcus).

2 Hrs

Phylum Nemathelminthes(Nematoda)

Salient features, classification up to classes

Class: Phasmidia Eg. *Enterobius*, Class: Aphasmidia Eg. *Trichinella*

General Topic

Pathogenic nematodes in man. (Wuchereria bancrofti, Ascaris lubricoides, Ancylostoma duodenale, Trichinella).

Phylum Annelida:

Salient features, Classification upto classes.

Class I- Archiannelida	Eg. Polygordius
Class II -Polychaeta	Eg. Chaetopterus
ClassIII- Oligochaeta	Eg. Megascolex.
Class IV- Hirudinea	Eg. Ozobranchus, Hirudinaria

MODULE III

Phylum Onychophora

Eg. Peripatus (Mention its affinities).

Phylum Arthropoda Salient features, Classification upto classes

Type: Prawn – Fenneropenaeus (Penaeus)

1. Sub Phylum - Trilobitomorpha

Class -Trilobita (mention the salient features).

Eg. Triarthrus – A trilobite (extinct)

2. Subphylum – Chelicerata

Class 1 Merostomata (Xiphosura) (Eg. Limulus)

Class 3 Pycnogonida (Eg. *Pycnogonum* – Sea spider)

3. Subphylum- Crustacea

Class 1 Branchiopoda Eg. Daphnia

Class 2 Ostracoda Eg. Cypris -seed shrimp

Class 3 Copepoda Eg. Cyclops

- Class 4 Remipedia Eg. Speleonectes (eyeless crustacean seen in caves)
- Class 5.Branchiura Eg., Argulus (common fish louse)
- Class 6 Cirripedia Eg. <u>Sacculina</u> (parasitic castrator of crabs)
- Class 7 Malacostraca Eg. Squilla (spot tail mantis shrimp)

4. Subphylum- Uniramia

Class 1 Chilopoda	Eg. Scolopendra – (Centipede)
Class 2 Symphyla	Eg. Scutigerella – (garden centipedes or pseudocentipedes)
Class 3 Diplopoda	Eg. Spirostreptus- (Millipede)
Class 4 Pauropoda	Eg. Pauropus

2 Hrs

14 Hrs

Class 5 Hexapoda (Insecta) Eg. Bombyx mori – (silk moth)

MODULE IV

Phylum Mollusca		3 Hrs
Salient features, Classification	upto classes	
Class I- Apalcophora	Eg. Neomenia	
Class II- Monoplacophora	Eg. Neopilina	
Class III Amphineura	Eg. Chiton	
Class IV Gastropoda	Eg. Aplysia	
Class V Scaphopoda	Eg. Dentalium	
Class VI Pelecypoda (Bivalvia)) Eg. Pinctada	
Class VII Cephalopoda	Eg. Sepia	
Phylum Echinodermata		3 Hrs
Classification upto classes		
Class I- Asteroidea	Eg. Astropecten	
Class II- Ophiuroidea	Eg. Ophiothrix	
Class III- Echinoidea	Eg. Echinus	
Class IV- Holothuroidea	Eg. Holothuria	
Class V – Crinoidea	Eg.Antedon	
General Topics		
1. Water vascular system in Ec	hinodermata	
Phylum Hemichordata:		2 Hrs
Eg. Balanoglossus		
Minor Phyla		
1. Chaetognatha E	Eg. Sagitta	
2. Sipunculida	Eg. Sipunculus	

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PRACTICAL

ANIMAL DIVERSITY- NON CHORDATA

36 Hrs. Credit 1

Scientific Drawing:-

Make scientific drawings of 5 locally available invertebrate specimens belonging to different phyla.

Anatomy:-

Study of sections. (Any two)

- 1. Hydra.
- 2. Ascaris(male and female)
- 3. Earthworm
- 4. Fasciola

Dissections

- 1. Prawn Nervous system
- 2. Cockroach Nervous system

Mounting:-

- 1. Prawn appendages.
- 2. Mouth parts Cockroach/ Plant bug/ House fly / Mosquito. (Any Three)

Identification:-

General identification &classification - The students are expected to identify, classify and describe the following Phylum -wise number of animals by their common names, generic names and 30% of these by their scientific names. Porifera-1, Coelenterata-3, Platyhelminthes-2, Annelida-2, Arthropoda-5, Mollusca- 4, Echinodermata-3 Identification of (a) Parasitic protest – any 2 (b) larval forms of *Fasciola-* any 2 (c) Nematode parasites of man- any 3 (Slides/figures may be used for study)

Taxonomic identification with key:-

Identification of insects up to the level of Order (any Four).

SEMESTER 111. ZY3CRT03

CORE COURSE 111: ANIMAL DIVERSITY – CHORDATA

54 Hrs

3 Credits

Objectives

- To acquire in depth knowledge on the diversity of chordates and their systematic position.
- To make them aware of the economic importance of some classes.
- To understand the evolutionary importance of selected chordate groups

MODULE I

Introduction	1 Hr
General Characters and outline classification of Chordata up to class, Or	igin of Chordates –
mention theories in brief	
Protochordates: General characters and Classification	2 Hrs
1. Sub phylum: Urochordata	
Class I Larvacea Eg. Oikopleura	
Class II Ascidiacea Eg: Ascidia (Mention Retrogressive Metamorphosi	is)
Class III Thaliacea Eg: Doliolum	
2. Sub phylum: Cephalochordata	2 Hrs
Example - Amphioxus (Structure and affinities)	
MODULE II	
3. Sub phylum: Vertebrata General characters and Classification	2 Hrs
4. Division 1– Agnatha	2 1115
Class I Ostracodermi Eg: <i>Cephalaspis</i>	
Class II Cyclostomata Eg: Petromyzon	10 11
Division 2 – Gnathostomata	10 Hrs
Super class Pisces General Characters and Classification	
Class: Chondrichthyes - General Characters	
Sub class – ElasmobranchI Eg: <i>Narcine</i>	
Sub class - Holocephali Eg: Chimaera	
Class: Osteichthyes - General Characters	
Sub class – Choanichthyes	
Order 1 Crossopterigii(Coelocanths) Eg: Latimeria(Evolu	utionary
Significance)	
Order 2 Dipnoi Eg: <i>Lepidosiren</i> - Distribution, affinities an	nd systematic
position of lung fishes.	
Sub class: - Actinopterygii	
Super order 1. ChondrosteI Eg: Acipencer	
Super order 2. Holostei Eg: Amia	
Super order 3. Teleostei Eg: Sardine	
General topics	

1. Accessory respiratory organs in fishes.

- 2. Parental care in fishes.
- 3. Scales in fishes.
- 4. Migration in fishes

MODULE III

Super class: Tetrapoda General characters, Classification up to Orders 11 Hrs

Class Amphibia - Type Frog (Euphlyctis hexadactylus)

Order I Anura Eg: Hyla

Order II Urodela Eg: *Amblystoma* (mention axolotl larva and Paedomorphosis /neotony)

Order III Apoda Eg: Ichthyophis.

Class Reptilia

Sub class I: Anapsida	
Order Chelonia	Eg: Chelone
Sub class II: Parapsida	Eg: Ichthyosaurus
Sub class III: Diapsida	
Order I Rhynchocephalia	Eg: Sphenodon
Order II Squamata	Eg: Chamaleon
Order III. Crocodilia	EgCrocodylus
Sub class IV: Synapsida	Eg: Cynognathus

General topic

Identification of poisonous and non-poisonous snakes

Class Aves

5 Hrs

4 Hrs

Sub class I: Archeornithes	Eg: Archaeopteryx (Affinities)
Sub class II: Neornithes	
Super order I: Palaeognathe	Eg: Struthio
Super order II: Neognathe	Eg: Brahminy kite

General topics

1. Migrations in birds

2. Flight adaptations in birds

MODULE IV

Class Mammalia Type: Rabbit (Oryctolagus cuniculus)

Brief mention of general characters and classification up to order with example. (Mention any five salient features of each order, detailed accounts of examples are not necessary)

17 Hrs

Sub class I: Prototheria

Sub class II: Metatheria

Sub class III: Eutheria

Eg: Echidna, Ornithorhychus Eg: Macropus Eg: Talpa

Order 1 Insectivora	Eg: Talpa
Order 2 Dermoptera	Eg: Galeopithecus
Order 3 Chiroptera	Eg: Pteropus
Order 4 Primates	Eg: Loris
Order 5 Carnivora	Eg: Panthera
Order 6 Edentata	Eg: Armadillo
Order 7 Pholidota	Eg: Manis
Order 8 Proboscidea	Eg: Elephas
Order 9 Hydracoidea	Eg: Procavia
Order 10 Sirenia	Eg: Dugong
Order 11 Perissodactyla	Eg:Rhinoceros
Order 12 Artiodactyla	Eg: Camelus-mention ruminant stomach
Order 13 Lagomorpha	Eg: Oryctolagus
Order 14 Rodentia	Eg: Hystrix (Porcupine)
Order 15 Tubulidentata	Eg: Orycteropus
Order 16 Cetacea	Eg: Delphinus

General topics

- 1. Dentition in Mammals
- 2. Aquatic Mammals and their adaptations.

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PRACTICAL

ANIMAL DIVERSITY - CHORDATA

36 Hrs

Credit 1

1. Scientific Drawing

Make scientific drawing of 5 locally available vertebrate specimens belonging to different classes

2. Dissections

Frog: Photographs/diagrams/one dissected & preserved specimen each/models may be used for study.

- 1. Frog Viscera
- 2. Frog Digestive System
- 3. Frog Arterial System
- 4. Frog 9th& 1st Spinal nerve
- 5. Frog Sciatic Plexus
- 6. Frog Brain
- 3. Mounting of placoid scales; study of cycloid and ctenoid scales

4. Osteology

Frog vertebrae - typical, atlas, 8th and 9th

Rabbit – Atlas, Axis and typical vertebra

Pectoral and pelvic girdles of Frog and Rabbit

Bird - Keel and Synsacrum

Turtle/Tortoise - plastron and carapace

5. Study of sections.

Amphioxus T. S. through pharynx/T.S. through intestine

6. Identification:-

General identification-

Identify, classify and describe the following animals by their generic names and 30 % of them by their scientific names.

Protochordata-1, Pisces-5, Amphibia-5, Reptilia- 5, Aves-2, Mammalia-2.

Taxonomic identification with key:-

- i) Identification of fishes up to the level of order.
- ii) Identification of snakes up to family.

SEMESTER IV. ZY4CRT04

CORE COURSE IV

RESEARCH METHODOLOGY, BIOPHYSICS AND BIOSTATISTICS

54 Hrs

3 Credits

Objectives

- 1. To familiarise the learner the basic concept of scientific method in research process.
- 2. To have a knowledge on various research designs.
- 3. To develop skill in research communication and scientific documentation.
- 4. To create awareness about the laws and ethical values in biology.
- 5. To equip the students with the basic techniques of animal rearing collection and preservation
- 6. To help the student to apply statistical methods in biological studies.

RESEARCH METHODOLOGY

Module I

Basic concepts of research: Meaning, Objectives, Approaches, Types of research.

Research Process: Scientific method in research (eight steps).

Importance of literature reviewing in defining a problem,

Identifying gap areas from literature review.

Research Communication and scientific documentation: Project proposal writing,

Research report writing, (Structure of a scientific paper), Thesis, dissertation, research article.

Presentation techniques: Oral presentation, Assignment, Seminar, Debate, Workshop,

Colloquium, Conference

Sources of Information: Primary and secondary sources. Library- Books, Journals,

Periodicals, Reviews, Internet.

Search engines Online libraries, e-Books, e-Encyclopedia, Institutional Websites. Plagiarism

Module II

12 Hrs

Animal Collection – Tools &techniques

Sampling techniques Quadrate Line transect Measurements Density Abundance Frequency Biodiversity indices – concepts Simpson index

Collection methods, techniques and equipments

Plankton Insects Fish Bird

Rearing techniques

Laboratory and field.

Units of measurements- units, SI system, Equivalent weight, normality, molarity

BIOPHYSICS

Module III

14 Hrs

Basic understanding on principle and uses of the following:

Microscopy

(a) Light microscopy, Bright field (Compound Microscope), Phase contrast, Dark field microscopy, Fluorescence, Polorization microscopy, Video microscopy.

(b) Electron - Scanning (SEM), Transmission (TEM) and STEM

Micrometry – Stage and Eyepiece micrometers Camera Lucida Instrumentation pH Meter Separation Techniques: Centrifuge, Chromatography, Electrophoresis Analytical techniques: Colorimeter, Spectrophotometer, X-ray crystallography

BIOETHICS

Module IV

5 Hrs

10 Hrs

Bioethics : Introduction, Animal rights and animal laws in India, Prevention of cruelty to animals Act 1960, Biodiversity Act 2003.

Concept of 3 R – conservation (Refined- to minimize suffering, Reduced – to minimize animals, Replaced – modern tools and alternate means), Animal use in research and education.

Laboratory animal use, care and welfare, Animal protection initiatives- Animal Welfare Board of India, CPCSEA, ethical commitment. Working with human: Consent,harm, risk and benefits.

BIOSTATISTICS

Module V

Sample & Sampling techniques: Collection of data, classification of data, frequency distribution tables, graphical representation: - Bar diagrams, Histogram, Pie diagram and Frequency curves - Ogives.

Measures of Central Tendency: Mean, Median, Mode (Problem - Direct method only) Measures of dispersion: Range, Quartile Deviation, Mean Deviation, Standard Deviation, Standard error. (Merits & demerits and problems on SD).

Correlation: Definition, Types of correlation.(mention in brief)

Test of Hypothesis and Test of Significance: Basic concept, Levels of significance, test of significance, Procedure for testing hypothesis, types of hypothesis- Null hypothesis and Alternate hypothesis.

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CORE COURSE IV

RESEARCH METHODOLOGY, BIOPHYSICS AND BIOSTATISTICS

(PRACTICAL) 2 credits

PART A. RESEARCH METHODOLOGY

Animal collection Tools, Techniques & Estimation

- 1. Quadrate study
- 2. Transect study
- 3. Sampling Methods
- 4. Species area curve
- 5. Simpson index

PART B - BIOPHYSICS

- 1. Study of simple and compound light microscopes
- 2. Micrometry -calibration and measurement of microscopic objects -low power
- 3. Camera Lucida (draw a few diagrams using Camera Lucida)
- 4. Paper chromatography (demonstration only)
- 5. Instrumentation demonstration (write notes on principle, equipment and its use)

pH Meter, Colorimeter/ Spectrophotometer, Centrifuge

PART C BIOSTATISTICS

- 1. MS Excel : To create mean and median, Construction of bar diagram, Pie diagram and Line graphs.
- 2. MS Access: To create grade of students
- 3. Internet: Access a web page on any biological topic.
- 4. Frequency distribution of the given samples to find out arithmetic mean, median, mode.
- 5. Range and standard deviation for a biological data
- 6. Correlation using any biological data.

7. Graphical representation of data. Construction of bar diagrams, Histograms, Pie diagram and Line graphs.

SEMESTER V. ZY5CRT05

CORE COURSE V

ENVIRONMENTAL BIOLOGY AND HUMAN RIGHTS

54 Hrs

Objectives

To instill the basic concepts of Environmental Sciences, Ecosystems, Natural Resources, Population, Environment and Society

To make the students aware of natural resources, their protection, conservation, the factors polluting the environment, their impacts and control measures.

To teach the basic concepts of toxicology, their impact on human health and remedial measures

To create a consciousness regarding Biodiversity, environmental issues & conservation strategies

To develop the real sense of Human rights – its concepts & manifestations

MODULE 1 ECOSYSTEM

12 Hrs

Basic concepts of ecosystem Components of ecosystem: Abiotic (Sunlight, temperature, soil, water, atmosphere) and Biotic components (Producers, consumers, decomposers), Ecological pyramid- number, biomass, energy, **Functions of ecosystem**: Productivity-Food chain-Food web-Energy flow-Laws of Thermodynamics.Types of Ecosystem: Terrestrial-Forest-Grassland-Desert, Aquatic-Marine-Fresh water, Wetland &Biome Concept of limiting factors: Liebig's and Shelford's laws of limiting factors.

Biogeochemical cycles: Concept, gaseous and sedimentary cycles, Carbon cycle, Nitrogen cycle. **Renewable resources** (solar,wind, hydroelectric, biomass and geothermal) **and Non renewable resources** (mineral and metal ore, fossil fuels)

MODULE 2 CONCEPTS OF POPULATION AND COMMUNITY 8 Hrs

Concept of population: Population attributes- Population growth forms, Basic concepts of growth rates, density, natality, mortality, growth curves

Animal interactions: Positive- Commensalism- Mutualism-Protocooperation, Negative-Predation-Parasitism-Competition-Antibiosis

Characteristics of a community: Species diversity- richness, eveness, stratification, dominance, ecological indicators, Ecotone and Edge effect, Keystone species, Concepts of Ecological Niche and Guild, Ecological succession, community evolution- climax.

MODULE 3 BIODIVERSITY AND ENVIRONMENTAL ISSUES 16 Hrs

Introduction to Biodiversity: Types of biodiversity- Alpha, Beta and Gamma diversity. **Concept and importance of Biodiversity**: Levels of Biodiversity-Species diversity, Genetic diversity, Microbial, Ecosystem diversity, India as a mega-diversity nation, Biodiversity hotspots

Global Environmental Issues: Ozone depletion, Greenhouse effect, Global warming, Climate change, Carbon trading, carbon credit; Carbon sequestration, Acid rain, Oil spills, Nuclear accidents, IPCC/UNFCC.

National Environmental issues: Deforestation, forest fire, pollution(air, water, soil, noise thermal, nuclear- brief account only) solid waste management, sewage, drinking water crisis and water logging,

Toxic products and disaster: Types of toxic substances – degradable, non degradable, Impact on human – case studies: Endosulphan tragedy, Bhopal disaster

Flood, drought, cyclone, earthquake and landslide (Management and mitigation)

Local Environmental issues: Landscape alteration, sand mining, quarrying, changing crop pattern, conversion of paddy lands,

Threats to water resources of Kerala: Degrading Mangrove and wetland ecosystems of Kerala, RAMSAR sites, Marine ecosystem crisis- pollution, overfishing etc. Impact of tourism on Environment.

Protected area concept – Sanctuary, National Park, Biosphere reserve, Core Zone, Buffer Zone, Corridor concept. Conservation reserves

Concept of threatened fauna – IUCN categories - extinct, extinct in the wild, critically endangered, endangered, vulnerable, near threatened, least concern and data deficient. Red and Green Data Books.

Man-animal conflict (Tiger, Elephant, Dog, Monkey) – causes and concern

Water conservation- rainwater harvestiong, watershed management

Environment education

Environmental laws (Brief account only): The Water (Prevention and Control of Pollution) Act, 1974, The Air (Prevention and Control of Pollution) Act, 1981, Indian Forests Act (Revised) 1982. The Environment (Protection) Act, 1986, Hazardous Wastes (Management and Handling) Rules, 1989, The Forest (Conservation) Act, 1980, The Wildlife Protection Act, 1972, Biodiversity Act, 2002.

MODULE 5 HUMAN RIGHTS

6 Hrs

Introduction, main concepts associated with Human Rights, Different types of human rights, Manifestations & phenomena, Role of agencies in promoting human rights, Mechanisms for checking violations of human rights, National human right commission, Constitutional provisions related to Human rights.

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- 8. Arun kumar Palai(1999) National Human Rights Commission of India, Atlantic publishers
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- Meera Asthana and Astana D.K.1990 Environmental pollution and Toxicology Alka printers.
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- Landis, Wayne and Hing-hoYu, Baca Raton, 1995. Introduction to Environmental Toxicology: Impacts of chemicals upon Ecological systems: Lewis Publishers.

PRACTICAL

ENVIRONMENTAL BIOLOGY & TOXICOLOGY

36 HRS CREDIT 1

- 1. Estimation of dissolved Oxygen
- 2. Estimation of carbon di oxide
- 3. Estimation of soil organic carbon (Demonstration only)
- 4. Identification of marine/ fresh water planktons
- 5. Counting of plankton using plankton counting chamber
- 6. Study of equipments Sechi disc, Plankton net
- 7. Study of sandy shore fauna, rocky shore fauna.
- 8. Study of animal Association
- Visit to any two important areas of bio diversity: 1. Forest, 2.Sea shore, 3. Mangrove, 3. Wet lands, 4. Bird sanctuary, 5. Wild life sanctuary, 6. Sacred groves Field study (compulsory)

SEMESTER V. ZY5CRT06

CORE COURSE VI CELL BIOLOGY AND GENETICS

54 Hrs

Credits 3

Objectives

1. To understand the structure and function of the cell as the fundamentals for understanding the functioning of all living organisms.

2. To make aware of different cell organelles, their structure and role in living organisms.

- 3. To develop critical thinking, skill and research aptitudes in basic and applied biology
- 4. To emphasize the central role of genes and their inheritance in the life of all organisms.

CELL BIOLOGY

Module I

Introduction of cell and Diversity of cells: History, Cell theory, Prokaryotes, Eukaryotes, Mycoplasmas, Virus, Virions and Viroids, Prions.

Cell membrane & Permeability: Molecular models of cell membrane (Sandwich model, Unit membrane model, Fluid mosaic model). Cell properties - permeability, Transport [Diffusion, Osmosis, Passive transport, Active transport, bulk transport], Cell coat and Cell recognition.

Module II

Cell Organelles :Structure and functions of following cell organelles: Endoplasmic reticulum - Structure and functions. Ribosomes (Prokaryotic and Eukaryotic) Golgi complex - Structure and functions. Lysosomes - Polymorphism - GERL concept, functions. Mitochondria - Structure and functions. Nucleus: Structure and functions of interphase nucleus, Nuclear membrane, pore complex, structure and functions of nucleolus

Chromosomes – Structure & organization, Heterochromatin, Euchromatin, Nucleosomes, Polytene chromosomes-Balbiani rings, Endomitosis, Lamp brush chromosomes.

10 Hrs

22 HRS

Module III

Cell Communication: Basic principles of cell communications,Cell signaling(in brief), Types of signaling, Mention signaling molecules (neurotransmitters, hormones, Growth Factors, Cytokines Vitamin A and D derivatives),

Cell Division: Cell cycle - G_1 , S, G_2 and M phases, Mitosis and Meiosis. The difference between Mitosis and Meiosis.

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- 3. Koshy Thomas & Joe Prasad Mathew (Editors) (2011) *Cell Biology and Molecular Biology*.
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.13. Gupta, P. K (2002) Cell and Molecular Biology, (2ed), , Rastogi Publications., Meerut

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cell structure and function. Saunder's College Publication

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GENETICS

32 Hrs 10 Hrs

Module I

MendelianGenetics: Mendel's experiments- Monohybrid Cross, Dihybrid Cross, Mendel's Laws, Test Cross, Back Cross and Reciprocal Cross. Chromosome Theory of Inheritance

Interaction of genes:Allelic: Incomplete Dominance (Four O Clock Plant).Co- Dominance (Skin colour in Cattle) Lethal Alleles: Dominant lethal gene[Creeper chicken] and recessive lethal gene

[cystic fibrosis].

Non Allelic: Complementary (Flower colour in Sweet Pea), Supplementary (Coat colour in mice), Epistasis - dominant (Plumage in poultry) and recessive (Coat colour in mice). Polygenes (Skin colour inheritance in man), Pleiotropism (Vestigial wing gene in Drosophila).

Multiple alleles – ABO Blood group system, Rh group and its inheritance. Erythroblastosis foetalis.

Module II

12 Hrs

Sex determination: Chromosome theory of sex determination (Autosome and Sex chromosomes), male heterogamy and female heterogamy, (xx-xy, xx-xo, ZZ-ZW, ZZ-ZO), Genic Balance theory of Bridges. Barr bodies, Lyon's hypothesis, Gynandromorphism, sex mosaics, intersex (Drosophila), Hormonal [free martin in calf] and Environmental (Bonelia) influence on Sex determination

Recombination and Linkage: Linkage and recombination of genes based on Morgan's work in Drosophila, Linked genes, Linkage groups, Chromosome theory of Linkage, Types of linkage- complete and incomplete. Recombination, cross over value, chromosome mapping. [Definition]

Sex Linked inheritance : Characteristics of Sex Linked inheritance, X Linked inheritance of man (Hemophilia), Y linked inheritance [Holandric genes], Incompletely Sex Linked genes

or pseudoautosomal genes (Bobbed bristles in *Drosophila*), Sex limited genes (Beard in man) and Sex influenced genes (inheritance of baldness in man).

Module III

Mutation: Types of mutations - Somatic, germinal, spontaneous, induced, autosomal and allosomal, chromosomal mutations, structural and numerical changes. Gene mutations. [Addition, Deletion and substitution].

HumanGenetics: Karyotyping, Normal Human chromosome Complement, Pedigree analysis, Aneuploidy and Non- disjunction. Autosomal abnormalities (Down syndrome, Cry du chat syndrome) Sex chromosomal abnormalities (Klinefelters syndrome, Turner's syndrome) Single gene disorder (Brief mention) Autosomal single gene disorder [sickle cell anaemia), Inborn errors of metabolism such as phenylketonuria, alkaptonuria, , Albinism. Multifactorial traits – polygenic disorder- cleft lip and cleft palate.

Genetic Counseling, Eugenics and Euthenics -Brief account only

References

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SEMESTER V

CORE COURSE VI CELL BIOLOGY AND GENETICS (PRACTICAL)

36 Hrs

2 Credits

PART A: CELLBIOLOGY

- 1. Squash preparation of onion root tip for mitotic stages
- 2. Mounting of polytene chromosome (Drosophila/Chironomous.) Demonstration
- 3. Tissues (permanent slides of epithelial tissues, striated muscle, smooth muscle, cartilage, bone)
- 4. Identification of cell organelles
- 5. Preparation of temporary whole mount.
- 6. Preparation of permanent whole mount (demonstration)
- 7. Preparation of human blood smear and identification of Leucocytes

PART B : GENETICS

- 1. Genetic problems on Monohybrid, Dihybrid Crosses and Blood group inheritance
- 2. Study of normal male and female human karyotype (use photographs or Xerox copies)
- 3. Abnormal human karyotypes Down, Edwards, Klinefelter and Turner syndromes (use photographs or Xerox copies)
- 4. Sexing of Drosophila.
- 5. Study of Barr body in human buccal epithelium

SEMESTER V. ZY5CRT07

CORE COURSE - V11: EVOLUTION, ETHOLOGY & ZOOGEOGRAPHY

Objectives:

- To acquire knowledge about the evolutionary history of earth living and nonliving
- To acquire basic understanding about evolutionary concepts and theories •
- To study the distribution of animals on earth, its pattern, evolution and causative factors
- To impart basic knowledge on animal behavioural patterns and their role

Prerequisite:

- Basic knowledge on principles of inheritance and variation •
- Knowledge on molecular basis of inheritance .
- Basic understanding on the mechanism and factors affecting evolution
- Knowledge on origin and evolution of man

PART I - EVOLUTION

Module I - Origin of life

Theories - Panspermia theory or Cosmozoic theory, Theory of spontaneous generation (Abiogenesis or Autogenesis), Special creation, Biogenesis, Endosymbiosis. Chemical evolution - Haldane and Oparin theory, Miller-Urey experiment;

Direct evidences of evolution - Recapitulation Theory of Haeckel, Fossilization, Kinds of fossils, fossil dating, Homologous organs and analogous organs.

Module II - Theories of organic evolution

Lamarckism and its Criticism, Weismann's Germplasm theory, Darwinism and its Criticism, Neo-Darwinism, Theory of De Vries,

Population genetics and evolution: Hardy-Weinberg Equilibrium, gene pool, gene frequency. Factors that upset Hardy-Weinberg Equilibrium, Effects of genetic drift on population: Bottleneck effect and founder effect

Module III – Nature of evolution

Species and Speciation: Species concept, subdivisions of species (sub species, sibling species, cline and deme), Speciation: Types of speciation, Phyletic speciation (autogenous

9 Hrs

13 Hrs

30 Hrs

8 Hrs

54 Hrs

Credits 3

and allogenous transformations), True speciation, Instantaneous and gradual speciation, allopatric and sympatric speciation

Isolation: Types of isolating mechanisms-Geographic isolation (mention examples) and Reproductive isolation. Role of isolating mechanisms in evolution

Microevolution, Macroevolution (Adaptive radiation -Darwin finches) Mega evolution, Punctuated equilibrium, Geological time scale, and Mass extinction (brief account only). **Evolution of Horse**

PART II- ETHOLOGY	14 Hrs
Module IV – Introduction	1 Hr
Definition, History and scope of ethology	
Module V – Learning, imprinting and behaviour	9 Hrs
Types of learning with examples; patterns of behaviors – types of rhythms, navigation,	
homing instinct, hibernation, aestivation; pheromones- types and their effect on behavior,	
hormones and their action on behavior (aggressive and parental behavior)	
Module VI – Social organization	4 Hrs

Social organization in insects (ants) and mammals (monkey), Courtship behaviour and reproductive strategies

PART III- ZOOGEOGRAPHY	10 Hrs
Module VII – General Topics	4 Hrs

Module VII – General Topics

Continental drift theory, Types and means of animal distribution, Factors affecting animal distribution; insular fauna - oceanic islands and continental islands,

Module VIII - Zoogeographical realms

Palaearctic region, Nearctic region, Neotropical region, Ethiopian region, Oriental region, Australian region (brief account with physical features and fauna, Wallace's line, Weber's line, Biogeography of India with special reference to Western Ghats

References:

EVOLUTION

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ETHOLOGY

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ZOOGEOGRAPHY

1. Briggs, J.C. (1996). Global Biogeography. Elsevier Publishers.(Module VI and VII).

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PRACTICAL

EVOLUTION, ETHOLOGY AND ZOOGEOGRAPHY

36 Hrs

Credit 1

- 1. Identification of Zoogeographical realms using map
- 2. Study on endemic species of each realm
- 3. Show the discontinuous distribution of (lung fishes, camel, elephant)
- 4. Providing a map trace the route of HMS Beagle
- Providing a map mark any two continental/oceanic islands.: Greenland, Madagascar, New Zealand, New Guinea, Maldives, Iceland, Hawaii – any two
- 6. Contributions of scientists (showing photos) Any four
- 7. Identification of different stages of horse evolution
- 8. Study on Homology and Analogy
- 9. Study on connecting links (Peripatus, Archaeopteryx, Protopterus, Echidna)
- 10. Pheromone traps
- 11. Skinner box & T Maze
- 12. Experiment to demonstrate phototaxis and chemotaxis using Drosophila/House fly

13. Identification of behaviour (Grooming/courtship dance of flamingos/stickle back fish/ Tail wagging dance/ Aggressive behaviour/ Auto/Allo grooming, Flehmen response) showing pictures (Any five)

SEMESTER V. ZY5CRT08

CORE COURSE VIII HUMAN PHYSIOLOGY, BIOCHEMISTRY, AND ENDOCRINOLOGY 54 Hrs Credits 3

Objectives:

- 1. This course will provide students with a deep knowledge in biochemistry, physiology and endocrinology.
- 2. Defining and explaining the basic principles of biochemistry useful for biological studies for illustrating different kinds of food, their structure, function and metabolism.
- 3. Explaining various aspects of physiological activities of animals with special reference to humans.
- 4. Students will acquire a broad understanding of the hormonal regulation of physiological processes in invertebrates and vertebrates.
- 5. By the end of the course, students should be familiar with hormonal regulation of physiological systems in several invertebrate and vertebrate systems.
- 6. This also will provide a basic understanding of the experimental methods and designs that can be used for further study and research.
- 7. The achievement of above objectives along with periodic class discussions of current events in science, will benefit students in their further studies in the

biological/physiological sciences and health-related fields, and will contribute to the

critical societal goal of a scientifically literate citizenry.

HUMAN PHYSIOLOGY

Module I

Nutrition: Nutritional requirements – carbohydrates, proteins, lipids, minerals (Ca, P, Fe, I), vitamins (sources and deficiency disorders). Importance of dietary fibre and antioxidants.Balanced diet, Recommended Dietary Allowance (RDA). Nutrition during pregnancy and lactation, Infant nutrition, Malnutrition(PEM).

Digestion: Anatomy and histology of digestive glands (liver, pancreas, salivary, gastric and intestinal). Digestion and absorption of carbohydrates, proteins and fats.Nervous and hormonal control of digestion.

Module II

Respiration: Phases of respiration (external respiration, gas transport and internal respiration). Respiratory pigments: Haemoglobin, Myoglobin (Structure and Function). Transport of respiratory gases - transport of oxygen, oxyhaemoglobin curve, factors affecting oxyhaemoglobin curve, transport of carbon dioxide,(chloride shift). Control of respiration.Respiratory disturbances (Hypoxia, Hypercapnia, Asphyxia).Physiological effect of smoking, carbon monoxide poisoning, Oxygen therapy and artificial respiration.

Circulation: ESR, Haemopoiesis, blood pressure, ECG. Haemostasis (blood coagulation) – clotting factors, intrinsic and extrinsic pathways, anticoagulants and its mechanism of action.Cardiovascular diseases (Jaundice, Atherosclerosis, Myocardial infarction, Thrombus, Stroke).Angiogram and angioplasty.

Module III

Excretion: Histology of Bowman's capsule and tubular part. Urine formation – glomerular filtration, tubular reabsorption, tubular secretion.Urine concentration – counter current mechanism. Acid – base balance, hormonal regulation of kidney function. Renal disorders (kidney stone, acute and chronic renal failure, and dialysis). Homeostasis: Definition, concept and importance in biological system. Thermal regulation and thermal adaptation in homeotherms.

Module IV

10 Hrs

31 Hrs

8 Hrs

8 Hrs

Nerve physiology: Ultra structure of neuron. Nerve impulse production (resting membrane potential, action potential), transmission of impulse along the nerve fiber, interneuron (synaptic) transmission, neuromuscular junction and transmission of impulses.Neurotransmitters (acetyl choline, adrenalin, dopamine).EEG. Memory, Neural disorders (brief account on Dyslexia, Parkinson's disease, Alzheimer's disease, Epilepsy). **Muscle physiology:** Ultra structure of striated muscle, muscle proteins (myosin, actin, tropomyosin, troponin), Muscle contraction and relaxation-Sliding Filament Theory, cross bridge cycle, biochemical changes and ATP production in muscle, Cori cycle. Kymograph,

Simple muscle twitch, muscle fatigue, tetanus, rigor mortis.

Module V

BIOCHEMISTRY

Carbohydrates: Basic structure, biological importance and classification of monosaccharides, oligosaccharides, polysaccharides with examples.

Proteins: Basic structure and classification of amino acids; structure, biological importance and classification of proteins with examples.

Lipids: Structure of fatty acid, saturated and unsaturated fatty acid, biological importance and classification of lipids with examples.

Vitamins and minerals: Major fat soluble and water soluble vitamins. Important minerals and trace elements required for living organisms. Biological importance of vitamins and minerals.

Enzymes: Chemical nature of enzymes, enzyme activation, enzyme inhibition, allosteric enzymes, isoenzymes, co-enzymes. Michaelis–Menten enzyme kinetics.

Module VI

10 Hrs

Carbohydrate metabolism: Glycogenesis, Glycogenolysis, Gluconeogenesis, Hexose monophosphate Shunt, Glycolysis, Citric Acid Cycle, Electron Transport Chain and ATP synthesis. Ethanol metabolism.

Protein metabolism: Deamination, Transamination, Transmethylation, Decarboxylation, Ornithine cycle.

59

15 Hrs

Lipid metabolism: Biosynthesis of fatty acids, Beta oxidation, physiologically important compounds synthesized from cholesterol.

ENDOCRINOLOGY

Endocrinology and reproduction

8 Hrs 8 Hrs

Module VII

Endocrine physiology: Hormones – classification and mechanism of hormone action. Major endocrine glands(Histology is not included) their hormones, functions and disorders (hypothalamus, pituitary gland, pineal gland, thyroid gland, parathyroid gland, islets of Langerhans, adrenal gland),. Homeostasis and feedback mechanism.

References

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Robert K. Murray and Victor W. Rodwell; 2012; Harper's Illustrated Biochemistry, Harper;. 29th edition (Lange basic science.)

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PRACTICAL

HUMAN PHYSIOLOGY, BIOCHEMISTRY, AND ENDOCRINOLOGY

36 Hrs

Credit1

HUMAN PHYSIOLOGY

- 1). Determination of haemoglobin content of blood
- 2). Total RBC count using Haemocytometer
- 3). Total WBC count using Haemocytometer
- 4). Estimation of microhaematocrit
- 5). Effect of hypertonic, hypotonic and isotonic solutions on the diameter of RBC.
- 6). Instruments: Kymograph, Sphygmomanometer and Stethoscope (principle and use)
- 7). Measurement of blood pressure using sphygmomanometer (demonstration only)

BIOCHEMISTRY

- 1. Qualitative analysis of protein, glucose, starch and lipids.
- Chromatography Determination of Rf value of amino acids and identification of amino acids (Identify the Amino Acids using different solvent front and solute front)

ENDOCRINOLOGY

- 1. Cockroach Corpora cardiaca & Corpora allata (Demonstration)
- 2. Effect of adrenalin on heart beat of Cockroach (Demonstration)

SEMESTER VI. ZY6CRT09

CORE COURSE IX DEVELOPMENTAL BIOLOGY

54 Hrs

3 Credits

Objectives:

- 1. To achieve a basic understanding of the experimental methods and designs that can be used for future studies and research.
- To provide the students with the periodicclass discussions of current events in science which will benefitthem in their future studies in the biological/physiological sciences and health-related fields
- 3. To contribute tocritical societal goal of a scientifically literate citizenry.

Module 1

10 Hrs

Introduction: Definition, Scope of developmental biology, sub-divisions (descriptive, comparative, experimental and chemical), historical perspectives, basic concepts and theories.

Reproductive Physiology: Gonads- anatomy of testis and ovary, spermatogenesis, oogenesis, gonadal hormones and their functions. Hormonal control of human reproduction - Female reproductive cycles (Estrous cycle, Menstrual cycle). Structure of mammalian sperm and egg, Pregnancy, parturition and lactation.Reproductive health and importance of sex education.

.**Egg types:** Classification of eggs based on the amount, distribution and position of yolk. Mosaic and regulative, cleidoic and noncleidoic eggs.Polarity and symmetry of egg.

Fertilization: Mechanism of fertilization-(Encounter of spermatozoa and Ova, Approach of the Spermatozoon to the Egg, Acrosome Reaction and Contact of Sperm and Ovum, Activation of Ovum, Migration of Pronuclei and Amphimixis,), Significance of fertilization, Polyspermy, Parthenogenesis- Different types and significance.

Module II

14 Hrs

Cleavage: Types, planes and patterns of cleavage, Cell lineage of Planaria. Influence of yolk on cleavage.

Blastulation: Morula, blastula formation, types of blastula with examples.

Fate maps: Concept of fate maps, construction of fate maps (artificial and natural), structure of a typical chordate fate map. Significance of fate map.

Gastrulation: Major events in gastrulation. Morphogenetic cell movements. Influence of yolk on gastrulation. Exogastrulation.Concept of germ layers and derivatives.

Cell differentiation and gene action: Potency of embryonic cells (Totipotency, Pleuripotency, Unipotency of embryonic cells). Determination and differentiation in embryonic development, Gene action during development with reference to Drosophila (maternal effect genes), Zygotic genes.

Module III

20Hrs

Embryology of Frog: Gametes, fertilization, cleavage, blastulation, fatemap, gastrulation, neurulation, notogenesis. Differentiation of Mesoderm and Endoderm, Development of eye.Metamorphosis of frog, Hormonal and environmental onrol.

Embryology of chick: Structure of egg, fertilization, cleavage, blastulation, fate map, gastrulation. Development and role of Primitive streak, Salient features of 18hour, 24 hour, 33 hour & 48 hour chick embryo.Extra embryonic membranes in chick.

Human development: Fertilisation, cleavage, blastocyst, implantation, placenta. Gestation, parturition and lactation.Human intervention in reproduction, contraception and birth control. Infertility, Invitro fertilization (test tube baby)

Module IV

5Hrs

Experimental embryology: Spemann's constriction experiments, Organizers and embryonic induction. Embryo transfer technology, cloning, stem cell research. Ethical issues.

Teratology / Dysmorphology, Developmental defects: Teratogenesis, important teratogenic agents.(Radiations, chemicals and drugs, infectious diseases) genetic teratogenesis in human beings,

Developmental defects: Prenatal death (miscarriage and still birth). Intrauterine Growth Retardation (IUGR).

Module V

General topics: Classification and functions of placenta in mammals. Prenatal diagnosis (Amniocentesis, Chorionic villi sampling, Ultra sound scanning, Foetoscopy, Maternal serum alpha-fetoprotein, Maternal serum beta-HCG).Regeneration in animals.

References

Anthony S. Fauci, Eugene Braunwald, Dennis L. Kasper, Stephen L. Hauser, Dan L. Longo,J. Larry Jameson and Joseph Loscalzo; 2008; Harriosns Principles of Internal Medicine;

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Balnisky B.I.; 1981 An Introduction to Embryology, W.B. Saunders and Co.

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Dutta 2007 Obstrestics, Church Livingston 17 Ed

Majumdar N. N -1985 Vetebrate embryology; Tata McGraw-Hill, New Delhi

- Melissa A & Gibbs, 2006; A practical Guide to Developmental Biology, Oxford university press (Int. student edition)
- Scott F. Gilbert; 2003; Developmental biology; Sinauer Associates Inc.,U.S.; 7th Revised edition.
- Vijayakumarn Nair, K. & George, P. V. 2002. A manual of developmental biology, Continental publications, Trivandrum
- Taylor D J, Green NPO & G W Stout. (2008) Biological Science third edition. Cambridge university press. Ref pp 748 biology 755

PRACTICAL

DEVELOPMENTAL BIOLOGY

36 Hrs

Credit 1

Model/Chart/ Slide may be used

64

- 1. Embryological studies- Blastula (frog, chick)
- 2. Embryo transfer, cloning, gastrula (frog, chick)
- 3. Amniocentesis
- 4. Embryotransfer technology, cloning
- 5. Study of placenta- pig and man
- 6. 18 hour, 24 hour, 33 hour and 48 hour chick embryo.
- 7. Candling method.
- 8. Vital staining- demonstration.
- 9. Male and female reproductive organs in cockroach
- 10. Calculate the fecundity of fish.
- 11. Calculate the gonado-somatic index of given fish.

SEMESTER VI. ZY6CRT10

CORE COURSE X. MICROBIOLOGY AND IMMUNOLOGY

54 Hrs 3 Credits

MICROBIOLOGY

Module I

Introduction: History and scope of microbiology. Outline classification of Microbes. (bacteria, fungus & virus)

Methods in Microbiology: Sterilization and disinfection - physical and chemical methods.

Culture media – selective media, enrichment media, differential media.Plating techniques and isolation of pure colony. Culture preservation techniques: refrigeration, deep freezing, freezing under liquid nitrogen, lyophilization.

Module II

Morphology and fine structure of bacteria: Size, shape, cilia, pili, flagella, capsule, cell wall and its composition. Cytoplasmic membrane, protoplast, spheroplast, intracellular membrane systems, cytoplasm, vacuoles, genetic material, cell inclusions, bacterial spores.

15 Hrs

Bacterial growth Curve, Staining techniques – gram staining.

Bacterial Reproduction Sexual – (conjugation, transduction)and Asexual (budding,, fragmentation). Virology: Structure of virus; Human, animal, and bacterial virus. Viral replication, cultivation of animal viruses.

Module III

8 Hrs

9Hrs

Infections & Diseases: Types of infections - primary, secondary and nosocomial infections.

(Brief Account only)Contagious diseases - epidemic, endemic and pandemic, mode of

Transmission - food, water, air, vectors and carriers.

Diseases: Epidemiology, symptomology, diagnosis and treatment. Bacterial - Clostridium

tetany (tetanus), Viral – HIV virus (AIDS), fungal – Candida albicans (candidiasis).

IMMUNOLOGY

Module IV

Introduction to Immunology: Innate and acquired immunity, passive (natural and artificial) and active immunity (Natural and Artificial).Mechanisms of innate immunity - barriers, inflammation, phagocytosis.

Lymphoid organs: Primary (Thymus, Bone marrow) and secondary lymphoid organs (lymph nodes, spleen).

Lymphocytes: T and B cells, Natural killer cells, memory cells, macrophages.

Module V

9 Hrs

Antigens, Types of antigens, haptens, adjuvants, immunoglobulin structure, classes and functions of immunoglobulins.

Types of Immunity- , humoral & cell mediated immunity Monoclonal & polyclonal antibodies

Antigen – antibody reactions, Precipitation test, Agglutination test, VDRL WIDAL, ELISA.

Auto immune diseases: Pernicious Anemia, Rheumatoid Arthritis. Immunodeficiency -

AIDS. Hyper sensitivity- Type I, (E.g. Anaphylaxix) II(Transfusion reaction), III (Arthus reaction) and IV (Mantaux Test) (in brief).

Vaccines

3 Hrs

Introduction Types of vaccines, Current Vaccines, Recent trends in vaccine preparation

References

- 1. Ananthanarayan R & Jayaram Paniker C K. (2009) Text Book of Microbiology Orient Longman Private Ltd.
- Gladys Francis & Mini K.D., (Editors) (2012), Microbiology, Zoological Society of Kerala, Kottayam.
- 3. Kuby J, Kindt T., Goldsby R. and Osborne B. (2007). Kuby immunology
- 4. Sharma K. (2005) Manual of Microbiology: Tools and Techniques, Ane books
- Susan Panicker & George Abraham (Editors) (2008), Micro Biology and Immunology, Zoological Society of Kerala, Kottayam.
- 6. Colemen: (2002). Fundamentals of Immunology
- Darla J. Wise & Gordon R. Carter: (2004): Immunology A Comprehensive Review Iowa state University Press. A Blackwell science company,
- 8. Hans G. Sch, Legal General Microbiology, Seventh Ed. Cambridge Low Price Ed.
- Helen Hapel, Maused Harney Siraj Misbah and Next Snowden: (2006) Essentials of Clinical Immunology Fifth Ed. Blackwell Publishing Company,
- Heritage, J, E.G.V. Evaus and R.A.Killungten (2007): Introductory Microbiology Cambridge University Press 6. Ivan Roitt I (2002) Essentials of Immunology ELBS.

PRACTICAL

MICROBIOLOGY AND IMMUNOLOGY

72 Hrs

2 Credits

- 1. Instruments -Autoclave, Hot air oven, Bacteriological incubator Laminar air flow
- 2. Preparation of solid and liquid media for microbial cultures.
 - (Ingradients, pH and method of preparation) (Demonstration)
 - (a) Solid media (1) Nutrient agar (2) Mac Conkey's agar
 - (b) Liquid Media(1) Nutrient broth (2) Peptone water.
- 3. Culture methods (Demonstration)
 - (a) Streak plate technique and isolation of pure colonies.
 - (b) Lawn culture (c) Pour plate culture (d) Liquid culture
- 4. Examination of microbes in living condition

Hanging drop method for demonstrating motility of bacteria.

- 5. Gram staining preparation, procedure, identification of Gram + ve and Gram –ve bacteria.
- 6. Antibiotic sensitivity test (demonstration only)
- 7. Streak plating (individual performance)
- 8. Preparation of a fungal smear Lactophenol cotton blue staining and mounting
- 9. Determination of ABO blood groups and Rh factor (Antigen antibody Reaction)
- 10. Study through photographs/ illustration, the primary immune (Bone marrow and thymus) and secondary immune (spleen and lymph nodes) organs in Rat/Man

SEMESTER VI. ZY6CRT11 CORE COURSE XI. BIOTECHNOLOGY, BIOINFORMATICS AND MOLECULAR BIOLOGY

BIOTECHNOLOGY

Module I

11Hrs

Introduction: Scope, Brief History, Scope and Importance

Tools and Techniques in Biotechnology: Enzymes (restriction endonucleases, ligases, linkers & adapters), Vectors-[Plasmids, Phage vectors, Cosmids, Artificial Chromosomes] Host cells. Basic steps & techniques in rDNA technology

Gene Libraries, Construction of genomic library and cDNA Library. PCR technique and DNA amplification, Brief description of screening methods – Probes, Nucleic Acid hydridization, In situ Hybridization, Fluorescence in situ Hybridization (FISH), Colony hybridization. Methods of transfer of desired gene into target cell.Blotting Techniques-Southern, Northern, Western blotting.DNA Finger printing (DNA Profiling) and its application. Molecular markers - RFLP

Module II

9 Hrs

Animal Cell Culture: Brief account on methods, substrates, media and procedure of animal cell culture, Stem Cells, types and potential use, Organismal Cloning- reproductive & therapeutic- brief account only.

Applications of Biotechnology: Applications in Medicine(insulin, growth hormone, gene therapy), Agriculture(GM plants and biopesticides),Environment(bioremediation), Industry (Single Cell Protein) and applications of Fermentation Technology- lactic acid, vitamins, food and beverages.

Potential Hazards of Biotechnological Inventions: Risks related to genetically modified organisms (GMO) and biologically active products, Biological warfare & Biopiracy. Protection of biotechnological inventions.Intellectual Property Rights, Patenting and patent protection.

References

1. Singh B.D Biotechnology 2002. Kalyan Publishers New Delhi.

2. Brown C.H., Campbell I & Priest F, G. 1987. Introduction of Biotechnology (Blackwell scientific publishers Oxford).

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3. Colin Ratledge Bijorn Kristiansesn, 2008. Basic Biotechnology 3 rd ed. Cambridge University.

4. Janarathanan S & Vincent S. 2007. Practical Biotechnology, Method of Protocols. University Press.

5. John E. Smith. Biotechnology Cambridge Low priced ed. (Third Ed) 2005 Madingan, Martinko and Parker 2002, Biology of Microorganisms, Brock Eighth Ed. Prentice Hall.

6. Singh B.D. Biotechnolgy 2002, Kalyan Publishers New Delhi.

7. Sudha Gangal 2007. Biotechnology Principles and & practice of Animal Tissue culture, Universities Press.

BIOINFORMATICS

Module III

Introduction: Definition, importance and role of bioinformatics in life sciences. Computational Biology.

Biological databases: Nucleotide sequence databases (NCBI- GENBANK, DDBJ and EMBL). Protein databases - structure and sequence databases (PDB, SWISSPROT and UNIPROT). Introduction to Sequences alignments: Local alignment and Global alignment, Pair wise alignment (BLAST and FASTA] and multiple sequence alignment. Phylogenetic Tree construction and Analysis

Module IV

Molecular visualization software - RASMOL. Basic concepts of Drug discovery pipe line, computer aided drug discovery and its applications. Human Genome Project.

MOLECULAR BIOLOGY

Module V

Nature of Genetic Materials: Discovery of DNA as genetic material – Griffith's transformation experiments. Avery Macarty and Macleod, Hershey Chase Experiment of Bacteriophage infection, Prokaryotic genome; Eukaryotic genome.Structure and.types of

8 Hrs

14 Hrs

6 Hrs

20 Hrs

DNA & RNA.DNA replication. Modern concept of gene (Cistron, muton, recon, viral genes)., Brief account of the following-- Split genes (introns and exons), Junk genes, Pseudogenes, Overlapping genes, Transposons.

Module VI

12 Hrs

Gene Expressions: Central Dogma of molecular biology and central dogma reverse, one gene-one enzyme hypothesis, One gene-one polypeptide hypothesis Characteristics of genetic code, Contributions of Hargobind Khorana.

Protein synthesis [prokaryotic]: Transcription of mRNA, Reverse transcription, post transcriptional modifications, Translation, Post translational modifications.

Gene regulations: Prokaryotic(inducible & repressible systems) Operon concept -Lac operon and Tryptophan operon, Brief account of Eukaryotic gene regulation.

References

1. Bruce Albert, Bray Dennis, Levis Julian, Raff Martin, Roberts Keith and Watson James

(2008). Molecular Biology of the Cell, V Edition, Garland publishing Inc., New York

and London.

- De Robertis, E.D.P. and De Robertis, E.M.F. (2006).Cell and Molecular Biology.VIII Edition. Lippincott Williams and Wilkins, Philadelphia.
- 3. Gupta, P. K (2002) Cell and Molecular Biology, (2ed), , Rastogi Publications., Meerut
- 4. James Darnell. (1998) Molecular Biology. Scientific American Books Inc
- Thomas AP(Editor). 2011 Cell & Molecular Biology The Fundamentals. Green leaf publications .TIES Kottayam
- 6. Zoological Society of Kerala Study material. (2011) Cell and Molecular Biology

PRACTICAL.

BIOTECHNOLOGY, BIOINFORMATICS & MOLECULAR BIOLOGY BIOTECHNOLOGY

- Identify and comment on the item provided: (Western blotting / Southern blotting / Northern blotting / PCR)
- 2. Write down the procedure involved in DNA isolation

BIOINFORMATICS

1. Download/use print out/pictures of genome sequences of any 2 organisms. Identify

and mention the characteristic features of both.

- 2. Download/ use print out/pictures of a protein sequence , identify it & comment on its amino acid composition
- 3. Download / use print out/pictures of a macromolecule. Write a brief note on the bioinformatics tool used to visualize its structure.

MOLECULAR BIOLOGY

1. Identify and comment on its molecular composition / structural orientation / functional significance (Any tissue / Cell organelles/ DNA, DNA replication, RNA different types using models or diagrams)

V1 SEMESTER. ZY6CRT12 CORE COURSE XII OCCUPATIONAL ZOOLOGY . (APICULTURE, VERMICULTURE, QUAIL FARMING & AQUACULTURE)

54 Hrs Credits 3

Objectives:

- 1. To equip the students with self employment capabilities.
- 2. To provide scientific knowledge of profitablefarming.
- 3. To make the students aware of cottage industries.

Module 1. APICULTURE

Definition, Different species of honey bees, Organization of honey bee colony, Social life and adaptation of honey bees. Communication among honey bees. Bee keeping methods and equipments, Management and maintenance of an apiary, Growth period, honey flow period and dearth period Division of the colony, uniting two colonies, , replacing old queen with new queen, swarming management, monsoon management. Enemies of bees. Diseases of bees, Bee pasturage. Uses of honey bees, By-products of honey bees, Honey and wax composition. Testing the quality of honey.Extraction of wax, Uses of honey and wax.Royal jelly, Propolis. Apitherapy, Agencies supporting apiculture.

Activity :Visitto an apiculture unit.

Field visit and report submission - 10 Hrs

Field visit and report submission on any two items are taken for internal evaluation.

MODULE: 2. VERMICULTURE

Introduction, Ecological classification of earth worms. Species of earth worms used for vermicultre, Reproduction & life cycle, Role of earth worm in solid waste management, in agriculture, in medicine etc. Preparation of vermibed, Maintenance & monitoring, Preparation of vermicompost, Preparation of vermiwash.

8 Hrs

Activity : Submission of a report after preparing a vermiculture unit or visiting a vermicomposting unit.

MODULE: 3.QUAIL FARMING (Coturnix coturnix) 4 Hrs

Introduction, care of quail chicks, care of adult quails, care of breeding quails, ration for quail, care of hatching eggs, health care, use of quail egg and meat.Sources of quality chicks.

MODULE: 4. AQUACULTURE. 24 Hrs

Advantages and salient features of aquaculture, Types of Aquaculture, Biotic and abiotic features of water, Importance of algae in aquaculture, Common cultivable fishes of Kerala, Fish diseases, Composite fish culture, Integrated fish culture, Carp culture, Prawn culture Mussel culture Pearl culture. Processing & Preservation.

Aquarium management - Setting up of an aquarium, Biological filter & Aeration, Breeding of gold fish, gourami (Osphronemus), fighter and Guppy (live bearer). Nutrition and types of feed for aquarium fishes, Establishment of commercial ornamental fish culture unit. Fish Transportation - Live fish packing and transport Common diseases of aquarium fishes and their management. Aquaponics (a brief introduction only).

Activity – Setting up of an Aquarium

Field visit – Visiting an Aquaculture farm

References

NPCS Board, The complete book on Bee keeping and honey processing, NIIR Project consultancy services, 106E, Kamala nagar, Delhi- 110007.

Shukla G.S, & Updhyay V.B, Economic zoology ,Rastogi Publ. Meerut.

Pradip.V.Jabde, Text book of applied zoology, 2005

Applied Zoology, Study Material Zoological Society of Kerala , CMS college Campus

Clive. A Edwards, Norman. Q. & Rhonda. 2011. Vermitechnology: earthworms, organic waste & environmental management.

Chauhan, H.V.S. Poultry, Disease, diagnosis and treatment, Wiley eastern Ltd Delhi.

Otieno.F.O 2014. Quail farming: markets & market strategies

Pillai T.V.R., Aquaculture, principles and practices.

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Harisankar J. Alappat& A. Bijukumar, Aquarium Fishes. B. R. Publ. Corporation, Delhi.

MPEDA, A hand Book on AquafarmingOrnamentalfishes, MPEDA, Kochi.

Amber Richards. 2014. Aquaponics at home.

Pradip.V.Jabde. 1993. Text book of applied zoology

Venkitaraman, P.R,1983, Text book of Economic zoology(SudharsanaPuubl. Kochi)

Addison Webb, Bee Keepingfor profit and pleasure, Agrobios Ltd.

Edwards.C.A.&Lafty, J.R.1972 Biology of earthworms(Chapman & Hall Led.London)

Applied Zoology, Study Material Zoological Society of Kerala, CMS college Campus

George cust& Peter Bird, Tropical Fresh water Aquaria, Hamlyn London.

Verreth J. Fish larval nutrition, Chapman & Hall Publ.

Bone Packer. 2014. Aquaponic system

PRACTICAL OCCUPATIONAL ZOOLOGY

36 Hrs

Credit 1

1. General Identification, Economic importance, Morphology, scientific names and common names of the following

- a) Economic important and morphology of culturable fishes (Catla, Rohu, Grass carp, Common carp, Silver carp, *Etroplus suratensis*, *Oreochromis /Tilapia*, *Mugil cephalus* and *Anabas Testudineus*)
- b) Identification and morphology of ornamental fishes (gold fish, fighter, Gourami, Angel fish, Guppy
- c) Two species of earthworms used in Vermiculture
- d) Four species of honey bees
- e) Economic importance and morphology of shell fishes (Any three species of prawn, two marine mussels, two oysters one rock oyster *Crasostria* and pearl oyster *Pinctada fucata* and freshwater mussel *Lamellidens marginalis*).

2. Castes of bees

3. Principle & uses of - Aquarium filters, Aquarium aerator, Aquarium plants, Oven, Pelletiser, Screw Press, die plate

4. Identification and study of fish parasites and diseases (five numbers each) using slides/pictures

- 5. Bee keeping equipments, Beehive, Smoker, honey extractor, Queen Cage,
- 6. Bees wax, Honey, Vermicompost (Identification-Uses)
- 7. Formulation of artificial feed for aquarium fishes demonstration
- 8. Tests for determining the adulteration in honey.
- 9. Mounting of pollen basket
- 10. Mounting of mouth parts of honey bee
- 11. Separation of cocoon from worm castings.

SEMESTER V. OPEN COURSES (FOR OTHER STREAMS)

ZY50PT01

1. VOCATIONAL ZOOLOGY

72 Hrs

4 Hrs/Week, Credits 3

Objectives of the Course

- To develop critical thinking skill and research aptitude among students, by introducing the frontier areas of the biological science.
- To emphasize the central role that biological sciences plays in the life of all organisms.
- To introduce the student to some of the present and future applications of bio-sciences
- To acquire basic knowledge and skills in aquarium management,
 Quail farming, vermicomposting and apiculture for self-employment
- To learn the different resources available and to develop an attitude towards sustainability
- Give awareness to society about need for waste management and organic farming

Module 1 Aquarium management

General introduction to Aquarium, Aims and types of aquarium (material, size and shape), Requirements of an aquarium - filtration of waste, physical, chemical and biological; Setting

an aquarium (self-sustainable with biological filters), Major indigenous aquarium fishes of Kerala.

Activity: Setting up of a freshwater aquarium and rearing of aquarium fishes

Module 2 Ornamental Fish Culture

Introduction to ornamental fishes: Present status of ornamental fish culture in India with special reference to Kerala, Breeding of Gold fish, Fighter, Gourami (*Osphroneus*), and Guppy (live bearer). Nutrition and types feed for aquarium fishes,Use of live fish feed organisms in Ornamental fish culture. Methods and techniques involved in the formulation of fish feed. Fish Transportation: Live fish packing and transport, Common diseases of

20 Hrs

aquarium fishes and their management. Establishment of commercial ornamental fish culture unit,

Activity: field visit to an ornamental fish breeding Centre to understand breeding practices of various aquarium fishes.

Module 3 Quail farming (Coturnix coturnix)

Introduction, care of quail chicks, care of adult quails, care of breeding quails ,ration for quail, care of hatching eggs, health care, use of quail egg and meat, Sources of quality chicks. **Activity**: Visit to a quail farm or viewing a quail documentary to familiarize the quail farming practices

Module 5 Vermiculture and composting

Introduction, ecological classification of earth worms, Life history, Species of earth worms used for vermicultre, Preparation of vermibed; Preparation of vermicompost, Preparation of vermiwash, Maintenance and management of vermicomposting unit, Role of vermiculture in solid waste management.

Activity: - Preparation of a vermiculture unit or visit to a vermicomposting unit.

Module 6 Apiculture

Definition, Uses of bees, species of bees cultured, organization of honey bee colony, bee keeping methods (modern method only) and equipments, management and maintenance of an apiary-growth period, dividing the colony, uniting two colonies, replacing old queen with new queen, honey flow period, Bee pasturage, Death period, Enemies of bees, Bee diseases, uses of honey and wax, Apitherapy, Propolis, Royal jelly, Agencies supporting apiculture. **Activity:** Identify different types of honey bees and rearing equipments

Field visit and report Submission

Field visit and report writing on any two items are taken for internal evaluation, instead of assignment and seminar. Conduct a workshop on various cultural practices and the preparation of byproducts.

References

- Applied Zoology, Study Material Zoological Society of Kerala, CMS College Campus, Kottayam.
- Addison Webb (1947), Bee Keeping- for profit and pleasure, Museum Press, agro bios India Ltd.
- Alka Prakash (2011), Laboratory Manual of Entomology, New age International, New Delhi.

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18 Hrs

10 Hrs

- 4. Arumugan N. (2008) Aquaculture, Saras publication.
- Biju Kumar A and Harishanker J Alappat (1995) A Complete Guide To Aquarium Keeping. Published by Books For All, New Delhi.
- Chauhan, H.V.S. and S. Roy, (2008). Fungal Diseases. In: Poultry Diseases, Diagnosis and Treatment, Chauhan, H.V.S. and S. Roy (Eds.). 3rd Ed., New Age International (P) Ltd., New Delhi
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- George Cust and & Peter Bird. (1978). Tropical Fresh water Aquaria, Published by Hamlyn London. illustrated by George Thompson.
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- Joy P.J., George Abraham K., Aloysius M. Sebastian and Susan Panicker (Eds) (1998) Animal Diversity, Zoological Society of Kerala, Kottayam
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- 15. Nalina Sundari, M.S and Santhi, R (2006) Entomology. MJP Publishers
- 16. NPCS Board of Consultants & Engineers, Chennai.(2015) The complete book on Bee keeping and honey processing, 2nd Edition, NIIR Project consultancy services, 106- E kamala Nagar Delhi – 110007.
- 17. Ronald j. Roberts (1978) Fish pathology, Cassel Ltd London.
- Vijayakumaran Nair, K, Manju, K.G. and Minimol, K. C.(2015) Applied Zoology, Academia press, Thiruvananthapuram

OPEN COURSE (FOR OTHER STREAMS)

ZY50PT02

2. PUBLIC HEALTH AND NUTRITION

72 Hrs

4hrs/Week

Credits 3

Objectives:

- To inculcate a general awareness among the students regarding the real sense of health.
- To understand the role of balanced diet in maintaining health.
- To motivate them to practice yoga and meditation in day-to-day life.

PART I HEALTH, EXERCISE & NUTRITION

Module 1 Definition and Meaning of Health

10 Hrs

Dimensions and Determination of Health

Physical Activity and Health benefits

Effect of exercise on body systems – Circulatory, Respiratory, Endocrine, Skeletal and Muscular

Programmes on Community health promotion (Individual, Family and Society) Dangers of alcoholic and drug abuse, medico-legal implications

Module 2	Nutrition and Health	10 Hrs	
	Concept of Food and Nutrition, Balanced diet		
	Vitamins, Malnutrition, Deficiency Disease		
	Determining Caloric intake and expenditure		
	Obesity, causes and preventing measures		
	Role of Diet and Exercise, BMI		
Module 3	Safety Education in Health promotion	8 Hrs	
Principles of Accident prevention			
Health and Safety in daily life.			
Health and Safety at work.			
First aid and emergency care.			

Common injuries and their management.

Modern life style and hypokinetic diseases.

Diabetese, Cardiovasculard disorders-Prevention and

Management.

Module 4 Life Skill Education

Life skills, emotional adjustment and well being,. Yoga, Meditation and Relaxation, Psychoneuroimmunology

8 Hrs

10 Hrs

PART II PUBLIC HEALTH AND SANITATION

Module 5 Public health and water quality. 11 Hrs Potable water, Health and Water quality Faecal bacteriae and pathogenic microorganisms transmitted by water. Determination of sanitary quality of drinking water, water purification techniques 15 Hrs Module 6 Public health and diseases Water borne dseases-Cholera and Typhoid.Prevention of Water borne diseases. **Food borne diseases and Prevention** Botulinum, Salmenellosis, Hepatitis A Vector borne diseases & Control measures Chikungunya, Filariasis and Dengu fever Zoonotic disease-Leptospirosis & its control Emerging diseases - Swine flue (H1N1), bird flue (H5N1), SARS, Anthrax Re-emerging diseases –TB, Malaria

Health Centre visit & Report Presentation

References:

- Gladys Francis & Mini K.D., (Editors) (2012), Microbiology, Zoological Society of Kerala, Kottayam.
- 2. Greenberg, Jerol S and Dintiman George B (1997) Wellness Creating a life of Health and Fitness , London Allyn and Bacon Inc.
- K Park, (2008) Park's Text Book of Preventive and Social Mediine 18th Edition. Banarasidass Bhenot Publication
- Norman Bezzaant HELP First Aid for everyday emergencies. Jaico Publishing House, Bombay, Delhi

- Tom Sanders and Peter Emery. (2004) Molecular basis of human nutrition: Taylor & Francis Publishers Ane Book
- Pelczar M.J. Jr. E.C.S. Chane & N.R. Krieg, Microbiology (Concept & Applications). 5th edition. Tata McGraw Publishing Company Ltd.

SEMESTER V.

OPEN COURSE (FOR OTHER STREAMS) ZY5OPT03

3. MAN, NATURE AND SUSTAINABLE DEVELOPMENT

72 Hrs 4Hrs/Week Credits 3

Objectives:

- 1. To understand how Man originated and attained present status
- 2. To learn the basic concepts of Ecosystems and its functioning
- 3. To study the use and abuse of nature by Man
- 4. To learn the different resources available on earth
- 5. To study global environmental problems and its impact on human well being
- 6. To appreciate the perspectives of Man on nature and learn the strategies for conservation
- 7. To familiarize with sustainable development and develop an attitude for sustainability

Module I. Man in Nature

Introduction

Evolution of Man

Out of Africa and Candelabra Model

The Fossils and the Molecular Evidences

Hunter-Gatherer and the Agriculturist

Speech and Languages

Cultural Evolution

Altruism and Morality

Module II. The Biosphere

Earth-Continents and Continental drift

10 Hrs

Concept of Landscapes and Habitats Lithosphere- Forest (Tropical and Temperate) Grasslands. Deserts and Montane The Biomes of the World Hydrosphere- Oceans, Estuaries Freshwater Water the Elixir of Life Atmosphere- Structure and stratification Module III.Dominance of Man on Earth 7 Hrs **Industrial Revolution** Human Population Growth **Resource Utilization Environmental Consequences** Modern Agriculture and Green Revolution **Environmental Impacts** Imperialism and its Ecological Root 7 Hrs

Module IV.Natural Resources

Renewable and Non- renewable Biodiversity Importance of Biodiversity -the Six E^S Hotspots of Biodiversity

Biotic Richness of India

Monoculture and loss of Genetic Diversity

Extinction Crisis, IUCN and Red Data Book

Module V.Global Environmental Issues Threatening Natural **Resources and Human Life**

10 Hrs

Deforestation, Landscape alterations, Soil erosion, Flood and Drought, Desertification, Overexploitation, Pollution (Air, Water and Soil- Pollutants and Consequences only), Acid rain, Ozone depletion, Greenhouse effect and Global Warming (use case studies to illustrate the points) Waste disposal (Biodegradable and Non-degradable eg. Plastic and E- waste), Oil spill Energy - Production, Consumption and its Impact on Environment Quality of the Environment and Human Health

Aldo Leopold, Thoreau, Rachel Carson Edward Abbey, Arne Ness, Carolyn Merchant, Vandana Shiva

Community initiatives

Module VII. Global Strategies for Conservation UN conference on Man and Environment-1972

on conference on Man and Environment-

Eco Spirituality, Eco-theology and Eco-feminism

Indigenous People's Perspective (tribal and traditional communities)

Native American, Amazonian, Australian Aborigines, Bishnoi Contributions of -John Muir,

UNEP and its Contributions

The World Conservation Strategy-1980

Module VI.Man's Perspective on Nature

World Commission on Environment and Development

The Earth Summit -1992

The UNFCC and IPCC

Conservation Strategies in India-MoEF

Legal System- Mention Major Conservation Acts

People's Participation in Conservation:

Chipko Movement and Narmada Bachao Andolan,

Silent Valley

Module VIII Sustainable Development

Definition and Concept

Principles and Goals

Environment versus Development Debate

Johannesburg Conference -2002

Strategies for Sustainable development

Sustainable Development in the era of Globalization

Gandhian Environmentalism

Education for Sustainable Development (UNESCO-ESD)

Building a Sustainable society

Sustainable life styles

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10 Hrs

10 Hrs

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SEMESTER VI.

ZOOLOGY CORE CHOICE BASED COURSES FOR B.Sc. ZOOLOGY PROGRAMME ZY6CBT01. ELECTIVE COURSE. 1. ECOTOURISM & SUSTAINABLE DEVELOPMENT

72 Hrs 4hrs/week

Credits 3

Objectives:

- 1. To introduce the concepts, principles and applications of tourism and its sustainability
- 2. To critically analyse the cost and benefits of ecotourism, including related laws and policies, community involvement and future trends
- 3. To develop an appreciation among students with respect to tourism development from the

sustainability perspective

4. To equip the students with basic knowledge for the emerging ecotourism industry

Module I. Fundamentals of Tourism	12 Hrs
Introduction- Tourism, concepts and definitions	
History, types, Characteristics	
The facilitating sectors	
Attractions	
Geography, heritage	
Wildlife, nature	
Quality Control	
Module II. Major areas of eco-tourism	10 Hrs
Concepts, practices and case studies for each:	
Marine tourism	
Wildlife tourism	
Adventure tourism	
Module III. Emerging trends in eco-tourism	10Hrs
Cultural tourism	
Pilgrimage tourism	
Farm tourism	
Backwater tourism	
Health tourism	
Module IV. Problems and prospects of eco-tourism	10 Hrs
Economics and benefits of ecotourism	
Cultural issues and negative aspects of ecotourism	
Environmental Impacts of Tourism	
Module V. Sustainable tourism	12 Hrs
Quality, Standards	
Systems of sustainable tourism: environmental, sociocultural, Economical	
Environment and conservation: basic principles	
Current practices of eco-conservation in tourism industry	
Sustainable tourism and society	
Community based ecotourism	
Eco-development committee (EDC) of Periyar Tiger Rerserve	
People initiatives	

Module VI. Eco-tourism guides

Ecotourism guiding and case studies

Activity

Field visit to Ecologically relevant places & Report writing10 Hrs

References

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ELECTIVE COURSE . ZY6CBT02

2. AGRICULTURAL PEST MANAGEMENT

72 Hrs

4 Hrs/week - 3 Credits

Objectives

- 1. To acquire basic skills in the observation and study of nature.
- 2. To impart basic awareness regarding pest problem and crop loss due to their dominance.
- 3. To inculcate interest in adopting biological control strategies for pest control.
- 4. To understand various pests affecting our local crops and select the best method for their control

5. To acquire basic knowledge and skills in agriculture management to enable the learner for self-employment.

Module I

Pest and crop loss: Introduction, historical perspective-origin of pest, Evolution of pest. Causes of pest outbreak- biotic, abiotic and genetic factors.Modern agricultural practices and pest problem - high yielding varieties, monoculture, fertilizers, pesticides, irrigation, and cultural practices.

Module II

Pest categories: Types of pests- insect pest and non-insect pest.

Insect pest: insect structure and function-external features (body parts), mouth parts of phytophagous insects, internal anatomy, growth, development, reproduction, life cycle and metamorphosis (one example each from ametabolous, hemimetabolous and holometabolous insect), diapause. types of insect pests-key pests, occasional pests, potential pests.

Non insect pests: General features, different types-Rodents(mention the nature of crop loss by them),Mites-Main types of mites; plant injury caused by mite, millipedes and centipedes, slugs and snails (mention the damage of invasive Giant African Snail).

Activity: Identify a minimum of 5 invasive species (plant / animal) in your locality and make a report on their ecological impact.

Module III

Pest and plants: Plant feeding insects-plant host range, types of injury, relationship of pest injury and yield.

Host plant resistance: Characterization of resistance, mechanism of resistance (antixenosis, antibiosis, tolerance), biophysical, biochemical and genetic bases of resistance.

Module IV

Pest control-principles and practices: Types of control-cultural control, biological control, chemical control, integrated pest management, miscellaneous control.

89

7 Hrs

20 Hrs

5 Hrs

Cultural control: Water management, tillage, sanitation, plant diversity, crop rotation, planting time, harvesting practices etc

Biological control: Parasitoids and predators, control by insect pathogens. Techniques in biological control-conservation, introduction and augmentation. Biopesticides

Chemical control: Origin of chemical control, chemistry, mode of action and nomenclature (organochlorines, organophosphates, carbamates, synthetic pyrethroids, miscellaneous group) of pesticides, pesticide formulations and pesticide appliances (sprayers and dusters). Brief mention of attractants, repellents, chemosterilants and pheromones

Activity 1: Conduct a workshop on preparation of biopesticides of various types suitable for kitchen garden and agricultural fields.

Integrated Pest Management (IPM)

Miscellaneous control: Mechanical (hand picking, exclusion by screens and barriers, trapping, clipping, pruning etc), physical (hot and cold treatment, moisture, light traps etc), sterility principle

Module V

Bionomics and control of major pests of crops and stored grains: Biology, life cycle and nature of damage by different pests of following crops and their control

Pests of paddy: *Leptocorisa acuta, Scirpophaga incertulas,* Spodoptera mauritia, *Orseolia oryzae, Nilaparvata lugens*

Pests of coconut: Oryctes rhinoceros, Rhyncophorus ferrugineus, Opisina arenosella, Aceria guerreronis

Pests of Banana: Cosmopolites sordidus, Pentalonianigronervosa

Pests of vegetables - Brinjal: Leucinodesorbonalis, Euzopheraperticella, Henosepilachnavigintioctopunctata, Urentiushystricellus

Gourds -Bactoceracucurbitae, Anadevidiapeponis, Epilachna spp. Raphidopalpafoveicollis, Baristrichosanthis

Pest of stored grains: Sitophilusoryzae, Corcyra cephalonica Triboliumcastraneum, Trogodermagranarium, Callasobruchuschinensis

Activity 2: Conduct a poster exhibition on various types of pests of paddy, coconut, banana and vegetable varieties of Kerala.

Activity 3: Collect different types of pest of stored grains from the local provision shops or houses and make a taxonomic study and prepare a powerpoint presentation on them.

Activity 4: Visit a minimum of 5 kitchen gardens in the neighborhood and enlist the common traditional pest control measures used in them.

Activity 5: Organise awareness classes on the ill effects of chemical pesticides and manure on human health with the support of local examples.

References

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ELECTIVE COURSE. ZY6CBT03.

3. VECTOR AND VECTOR BORNE DISEASES

72 Hrs

10Hrs

6 Hrs

25 Hrs

3 Credits

Objectives:

Module I

Introduction:Vector : mechanical and biological vector, Reservoirs, Host-vector relationship, Vectorial capacity, Host Specificity.

Insect vectors: Mosquitoes, flies, fleas, lice, ticks and bugs- General account of ecology morphology and mouth parts

Module I1

Salient features and distribution of mosquito species:*Anopheles, Aedes, Culex, and Mansonia.*

Module II1

Study of Vector Borne disease[Life cycle and pathology]: Mosquito-borne diseases – Malaria, Dengue, Chikungunya, Filariasis. Sand fly-borne diseases – Leishmaniasis, Phlebotomus fever. Tse- tse fly – sleeping sickness. House fly borne diseases :typhoid fever, cholera, dysentery, anthrax, Myiasis, . Flea-borne diseases – Plague, Typhus fever. Louse-borne diseases – Relapsing fever, Trench fever, Vagabond's disease, Phthiriasis.

Module IV

Introduction to Vector control: Aims, objectives and advantages.History and background, recent trends, alternatives to the use of insecticides (chemical & microbial), types of vector control - selective, integrated and comprehensive vector control.

Control measures of mosquitoes, sand fly, tsetse fly and domestic flies

ModuleV

8Hrs

Introduction to epidemiology: History, Definition, scope and uses of epidemiology. Epidemiology and public health. Achievements in epidemiology: Smallpox Methyl mercury

93

poisoning Rheumatic fever and rheumatic heart disease Iodine deficiency diseases Tobacco use, asbestos and lung cancer, Hip fractures. HIV/AIDS, SARS.

Field report on two case studies of epidemiology in India. 10 Hrs

References

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ELECTIVE COURSE. ZY6CBT04 4. NUTRITION, HEALTH AND LIFESTYLE MANAGEMENT

72 Hrs.

3 Credits

Objectives:

1. To provide students with a general concept of health and the parameters that define health and wellness.

2. To understand principles of nutrition and its role in health.

- 3. To familiarize the students regarding food safety, food laws & regulations.
- 4. To provide knowledge and understanding regarding life style diseases.
- 5. To promote an understanding of the value of good life style practices, physical fitness and

healthy food habits for life style disease management.

Module I

Nutrition and health: Nutritional requirements of man, classification of major nutrients

including protein, vitamins and minerals, water, role of fibre, biological value of food components, food groups and sources, balanced diet, RDA, BMI, BMR, Calorie intake and expenditure, Healthy eating pyramid, Nutrition in infancy, preschool, school, adolescent, pregnancy, lactation and old age. Nutrition in diseases and special conditions. Food safety: Nutrition education, food sanitation and hygiene, food adulteration and consumer protection.

Module II

18 Hrs

Understanding of health: Define health, basic concepts, dimensions of health, basic
parameters of health care. (Health Parameters: Individual normal standards, devices.1. Blood
pressure, 2. Brain activities and sleep, 3.Focus or attention, 4.Pulse, 5. Body temperature,
6. Daily physical activities, 7. Electrocardiogram (ECG), 8.Cardiac fitness 9. Stress,

10. Haematological parameters, 11. BMI

Module III

15 Hrs

Introduction to Life style diseases

Common life style diseases: Alzheimer's disease and other neural disorders, asthma, cancer, cardio vascular diseases - including hypertension, Atherosclerosis and stroke, chronic obstructive pulmonary disease, Diabetes Mellitus or Type 2 Diabetes, kidney disorders and chronic renal failure, constipation, depression, gastro-intestinal disturbances including diarrhoea and peptic ulcer, liver cirrhosis and other liver diseases, obesity, osteoporosis, occupational lifestyle diseases.

Modern lifestyle disorders: sleeping habits, junk food, poor eating habits, anxiety, food poisoning

Module IV

10 Hrs

Causes of lifestyle diseases: Defects of modern food habits and unbalanced diet options, food adulteration, environmental pollution, poor life style choices, drug abuse, tobacco smoking, alcohol and drug consumption, lack of adequate exercise, wrong body posture, disturbed biological clock, stressful environmental conditions

Module V

14 Hrs

Prevention and control of life style diseases:

Healthy life style habits and practices, healthy eating habits, exercise and fitness, good sleep patterns, a strict no to alcohol, drugs, and other illegal drugs. Uncontrollable factors like age, gender, heredity and race.

Healthy diet: disease prevention through appropriate diet and nutrition, avoiding foods that are high in fats, salt and refined products. Avoid junk food and replace by natural food/ organic food.

Physical exercise: Moderate exercise for fitness of body, walking, stretching, right postures of sitting & standing, relaxation and cutting down of stress, sports, aerobic exercise and yoga. Health literacy as a public health goal: Awareness programs in schools, colleges and through mass media.

References

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13. COMPLEMENTARY ZOOLOGY COURSES OFFERED BY ZOOLOGY DEPARTMENT FOR - MODEL I – BSc BOTANY / BSc HOME SCIENCE MODEL II – BSc BOTANY, BSc (AQUACULTURE) MODEL III – BSc (BIOLOGICAL TECHNIQUES AND SPECIMEN PREPARATION)

Semester	Code	Title of the Course	Hrs	Inst Hrs/week	Credit
1	ZY1CMT01	Non Chordate Diversity	36	2	2
1		Non Chordate Diversity (Practicals)	36	2	0
2	ZY2CMT02	Chordate Diversity	36	2	2
2		Chordate Diversity (Practicals)	36	2	0
2	ZY2CMP01	Practical 1 Non Chordate Diversity + Chordate Diversity (Practicals)			2
3	ZY3CMT03	Physiology and Immunology	54	3	3
3		Physiology and Immunology (Practicals)	36	2	0
4	ZY4CMT04	Applied Zoology	54	3	3
		Applied Zoology (Practicals)	36	2	
4	ZY4CMPO2	Practical 2 Physiology and Immunology + Applied Zoology (Practical)			2

COMPLEMENTARY COURSE : ZOOLOGY -MODEL I

SEMESTER I. ZY1CMT01.

COMPLEMENTARY COURSE 1 NON CHORDATE DIVERSITY

Objectives:

- 1. To study the scientific classification of invertebrate fauna.
- 2. To learn the physiological and anatomical peculiarities of some invertebrate phyla through type study.
- 3. To learn the unity of life with rich diversity of organisms & evolutionary significance of certain invertebrate fauna
- 4. To stimulate the curiosity of students' in the biota living around them.

Module I

Introduction: Five kingdom classification

Kingdom Protista: Salient features (any five important salient features) of each phylum with one example each (detailed account of example is not necessary).

Phylum Rhizopoda	(eg: Amoeba)	
Phylum Actinopoda	(eg: Actinophrys)	
Phylum Dinoflagellata	(eg: Noctiluca)	
Phylum Parabasalia	(eg: Trychonympha)	
Phylum Metamonda	(eg: Giardia)	
Phylum Kinetoplasta	(eg: Trypanosoma)	
Phylum Euglenophyta	(eg: Euglena)	
Phylum Cryptophyta	(eg: Cryptomonas)	
Phylum Opalinata	(eg: Opalina)	
Phylum Bacillariophyta	(eg: Diatoms)	
Phylum Chlorophyta	(eg: Volvox)	
Phylum Choanoflagellata	(eg: Proterospongia)	
Phylum Ciliophora	(eg: Paramecium)	
Phylum Sporozoa	(eg: Plasmodium)	
Phylum Microsporidia	(eg: Nosema)	
Phylum Rhodophyta	(eg: Red algae)	

10 Hrs

36 hrs Credits 2 General Topic: Pathogenic Protists – Plasmodium, Entamoeba

Module II 3 Hrs Phylum Porifera: Salient features (eg: Leucosolenia)

Phylum Coelenterata: Salient features and classification upto class.

Class 1: Hydrozoa (eg: Physalia)

Class 2: Schyphozoa (eg: Aurelia)

Class 3: Anthozoa (eg: Adamsia)

General Topic: Corals and Coral reefs.

Module III

6 Hrs

Phylum Platyhelminthes: Salient features and classification up to class.

Class 1: Turbelleria (eg: Planaria)

Class 2: Trematoda (eg: Fasciola)

Class 3: Cestoda (eg: Taenia solium)

Phylum Nematoda: Salient features and classification up to class.

Class 1: Phasmida (eg: Wuchereria)

Class 2: Aphasmida (eg: Trichinella)

Phylum Annelida: Salient features and classification up to class.

Class 1: Polychaeta (eg: Nereis) Class 2: Oligochaeta (eg: Pheretima) Class 3: Hirudinomorpha (eg: Hirudinaria)

Module IV

11 Hrs

Phylum Arthropoda: Salient features. Type study - Fenneropenaeus (Penaeus) - habitat, morphology, appendages, sexual dimorphism, digestive system, respiratory system, circulatory system, excretory system, nervous system, sense organs, reproductive system, larval stages.

Classification up to class with one example each

Subphylum Trilobitomorpha

Class 1: Trilobita (Extinct) (eg: Dalmanites)

Subphylum: Chelicerata

Class 1: Merostoma (eg: Limulus)

Class 2: Arachnida (eg: Spider)

Class 3: Pycnogonida (eg: Nymphon)

Subphylum Mandibulata

Class 1: Crustacea (eg: Daphnia)

Class 2: Chilopoda (eg: Centipede)

Class 3: Symphyla (eg: Scutigerella)

Class 4: Diplopoda (eg: Millipede) Class 5: Pauropoda (eg: Pauropus) Class 6: Insecta (eg: Butterfly)

Module V

Phylum Mollusca: Salient features and classification up to class

Class 1: Aplacophora	(eg: Neomenia)			
Class 2: Monoplacophora	(eg: Neopilina)			
Class 3: Polyplacophora	(eg: Chiton)			
Class 4: Bivalvia	(eg: Perna)			
Class 5: Gastropoda	(eg: Xancus)			
Class 6: Cephalopoda	(eg: Sepia)			
Class 7: Scaphopoda	(eg: Dentalium)			
Phylum Echinodermata : Salient features and classification up to class.				
Class 1: Asteroidea	(eg: Astropecten)			
Class 2: Ophiuroidea	(eg: Ophiothrix)			

Class 2: Ophiuroidea	(eg: Ophiothrix)
Class 3: Echinoidea	(eg: Echinus)
Class 4: Holothuroidea	(eg: Holothuria)
Class 5: Crinoidea	(eg: Antedon)

Phylum Hemichordata : Salient features (eg: Balanoglossus.)

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SEMESTER I

COMPLEMENTARY COURSE 1 - PRACTCAL NON CHORDATE DIVERSITY

36 Hrs Credit 1

- 1. Scientific drawing 5 specimens
- 2. Simple identification 10 invertebrates, out of which 5 by their scientific names
- 3. T.S Earthworm, T.S Fasciola
- 4. Dissection Nervous system of Prawn
- 5. Dissection Nervous system of Cockroach
- 6. Mounting Prawn Appendages

SEMESTER II. ZY2CMTO2.

COMPLEMENTARY COURSE 2 CHORDATE DIVERSITY

- 1. To make the student observe the diversity in chordates and their systematic position.
- 2. To make the a student ware of the economic importance of some chordates.
- 3. To learn the physiological and anatomical peculiarities of some vertebrate species through type study.
- 4. To stimulate the students' curiosity in vertebrates living associated with them.

Module I

Objectives

Phylum Chordata: Fundamental characters and outline classification upto class.

Sub phylum Urochordata: General characters,

Classification:

Class 1: Larvacea (eg: Oikopleura) Class 2: Ascidiacea (eg: Ascidia), Retrogressive metamorphosis. Class 3: Thaliacea (eg: Salpa)

Sub phylum Cephalochordata: Salient features (eg: Branchiostoma)

Module II

Sub phylum Vertebrata: Salient features

Division Agnatha : salient features and classification

Class 1: Cyclostoma (eg: Petromyzon)

Class 2: Class Ostracodermi (eg: Cephalapsis)

Division Gnathostomata: Salient features

Super class Pisces

Super class Tetrapoda.

Super class Pisces: Salient features and classification

Class 1: Chondrichthyes (eg: N

arcine)

Class 2:Osteichthyes (eg: Latimeria)

General Topic: Accessory respiratory organs in fishes.

4 Hrs

36 Hrs Credits 2

6 Hrs

7 111 5

Module III

Super class Tetrapoda: Salient features

Class 1: Amphibia : Salient features. **Type study:** *Euphlyctis hexadactyla* - Habitat, morphology, sexual dimorphism, coelom and viscera, skeletal system, digestive system, respiratory system, circulatory system, excretory system, nervous system, sense organs, reproductive system, development..

Classification up to order:

Order 1: Urodela (eg: Amblystoma) Order 2: Anura (eg: Bufo) Order 3: Apoda (eg: Icthyophis)

Module IV

Class Reptilia: Salient features and classification up to subclass

Sub class 1: Anapsida (eg: Chelone)Sub class 2: Diapsida (eg: Chamaeleon)

Sub class 3: Parapsida (eg: Icthyosaurus)

General Topics: Poisonous and non poisonous snakes of Kerala.

Class Aves: Salient features and classification up to subclass

Sub class Archeornithes (eg: Archaeopteryx)

Sub class Neornithes (eg: Struthio)

General Topics: Flight adaptation of birds

Module V

Class Mammalia: Salient features and classification up to subclass

Sub class 1: Protheria (eg: Echidna)

Sub class 2: Metatheria (eg: Macropus)

Sub class 3: Eutheria (eg: Elephas)

General Topic: General adaptation of aquatic mammals with example.

References

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14 Hrs

6 Hrs

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Young J.Z. (1981). The life of Vertebrate s (Oxford University Press).

SEMESTER II

COMPLEMENTARY COURSE 2- PRACTCAL CHORDATE DIVERSITY

36 Hrs Credit I

- 1. Simple identification of 10 chordates, out of which 5 by their scientific names
- 2. Osteology Vertebrae and girdles of Frog
- 3. Snake identification 3 poisonous and 3 non poisonous snakes with key
- 4. Mounting of placoid scales of shark
- 5. Dissections: Frog: Photographs/Diagrams/ models may be used for the study.
 - 1. Frog Viscera
 - 2. Frog Digestive System
 - 3. Frog Arterial System
 - 4. Frog Brain

SEMESTER III. ZY3CMT03. COMPLEMENTARY COURSE -3 PHYSIOLOGY AND IMMUNOLOGY

54 hrs Credits 3

Objectives

- 1 To appreciate the correlation between structure and function of organisms
- 2 To make the student aware of the health related problems, their origin and treatment.
- 3 To understand how efficiently our immune system work in our body.
- 4 To acquire knowledge about preventing common diseases rather than curing.

Module I

14 Hrs

Nutrition: Types of nutrition – autotrophy, heterotrophy. Nutritional requirements – carbohydrates, proteins, lipids, minerals (Ca, Fe, I), vitamins (sources and deficiency disorders), nutritional disorders **Respiration:** Transport of respiratory gases in blood - transport of oxygen, transport of carbon dioxide, chloride shift. Respiratory disturbances – Hypoxia, Hypercapnia, Asphyxia, physiological effect of smoking, carbon monoxide poisoning.

Circulation: Composition and functions of blood. Plasma and formed elements - WBC, RBC and platelets, Mechanism of blood coagulation – clotting factors, intrinsic and extrinsic pathways, anticoagulants. ECG, Blood pressure, Arteriosclerosis, Heamophilia, cerebral and pulmonary thrombosis.

Module II

Excretion: Structure of a nephron. Urine formation – glomerular filteration, tubular reabsorption, tubular secretion. Urine concentration – counter current mechanism. Composition of urine – normal and abnormal constituents. Hormonal regulation of kidney function. Kidney stone, dialysis.

Neuro physiology: Structure of a neuron. Myelinated and non myelinated nerve fibre, nerve impulse production (resting membrane potential, action potential), Impulse propagation, All or none law, saltatory conduction, synaptic transmission. Neurotransmitters (acetyl choline, adrenalin, dopamine), brain waves, EEG. Neural disorders - Parkinson's disease, Alzheimer's disease.

Muscle physiology: Types of muscles: striated, non striated and cardiac. Ultra structure of striated muscle, Mechanism of muscle contraction, cori cycle and muscle relaxation. Muscle fatigue, oxygen debt, Rigor mortis.

14 hrs

110

Endocrinology: Introduction to Endocrine system. Mechanism of hormone action, Endocrine glands - hypothalamus, pituitary gland, pineal gland, thyroid gland, parathyroid gland, endocrine pancreas, adrenal gland, thymus gland, testis and ovary. Physiological role of hormones, Hormonal disorders.

Module IV

Module III

Immunology: Introduction to immunology, types of immunity – innate, acquired, passive, active, mechanism of innate immunity (barriers, inflammation, phagocytosis). Types of antigens. Basic structure of immunoglobulins, Classes of immunoglobulins and functions. Antigen antibody reactions, Precipitation test, agglutination test, WIDAL, VDRL, HIV test (ELISA),

Module V

Immune response system: (Brief accounts of the followings)

Primary and secondary lymphoid organs, Cells of Immune system - T&B lymphocytes, natural killer cells, macrophages, plasma cells, memory cells, Monoclonal antibodies, Hybridoma technology. **Immune disorders:** Hypersensitivity, Auto immunity (rheumatoid arthritis) & Immunodeficiency (AIDS), Vaccines - BCG, DPT, Polio vaccine.

REFERENCES

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8 hrs

12 Hrs

COMPLEMENTARY COURSE - 3 PRACTCAL PHYSIOLOGY AND IMMUNOLOGY

36Hrs

Credit 1

- 1. Preparation of Human Blood smear & identification of leucocytes
- 2. Qualitative analysis of Reducing Sugar, Protein and Lipid
- 3. Action of Salivary amylase on Starch (Demonstration Only)
- 4. Estimation of Haemoglobin (Demonstration only)
- 5. Identification of human blood groups, A, AB, B and O, Rh factor
- 6. Instruments (Principle & uses)- Sphygmomanometer, Stethoscope

SEMESTER IV. ZY4CMT04. COMPLEMENTARY COURSE - 4 APPLIED ZOOLOGY

54 hrs **Credits 3**

Objectives

- 1. To acquire basic knowledge and skills in applied branches of zoology.
- 2. To understand the technology for utilising ecofriendly organisms around them for

beneficial purpose.

3. To equip the students for self employment opportunities with scientific knowledge to perform

profitably & confidently.

Module I

Aquaculture: Advantages of aquaculture, Traditional methods of aquaculture, Biotic and abiotic factors in water, Pond culture - construction and maintenance. Types of aquaculture, composite fish culture, integrated fish culture, induced breeding of carp & prawn, Importance of algae in aquaculture. Aquarium management - Setting up of an aquarium, biological filter and aeration. Common cultivable fishes of Kerala. Fish diseases, Prawn culture, mussel culture, pearl culture, Fish processing and preservation.

Module II

Sericulture: Four species of silkworms, life history of silkworm, silk worm rearing techniques, Mounting of silkworm - Chandrika, defective cocoons, harvesting and stifling of coccons. Silkworm diseases and pest, preventive and control measures.

Module III

Vermiculture: Species of earthworms, ecological classification of earthworms, life cycle and reproduction of earthworm. Physical & chemical effects of earthworms on soil, Vermicomposting site selection, preparation of pit, maintenance, monitoring and harvesting of vermicompost.

Module IV

Apiculture: Species of honey bees, organization of honey bee colony. Bee keeping methods and equipments. Apiary management and maintenance. Bee pasturage, byproducts of honey bees and their uses. Diseases, pests of honey bees and control measures.

112

12 Hrs

24 Hrs

6 Hrs

References:

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- Sinhan, V.R.P. & Ramachandran, V., 1985, Fresh water Fish Culture (ICAR, New Delhi)

SEMESTER IV

COMPLEMENTARY COURSE - 4 PRACTCAL APPLIED ZOOLOGY

2 Hrs/week 36 Hrs Credit 1

- 1. General identification, economic importance, morphology, scientific names and common names of the following
 - a. Economic importance and morphology of culturable fishes (Catla, Rohu, Grass carp, Common carp, Silver carp, Etroplus, Tilapia)
 - b. Two species of earthworms used in Vermiculture

- c. Two species of honey bees
- d. Silkworm. Cocoon/Adult
- 2. Castes of honey bees
- 3. Bee keeping equipments Bee hive, Smoker, honey extractor
- 4. Identification and uses Bee wax, Honey, Silk, Vermicompost
- 5. Chandrika / Natrika used in sericulture

14. MODEL II VOCATIONAL COURSES

MODEL II

B.Sc. PROGRAMME (VOCATIONAL COURSES)

- 1. B.Sc. (Aquaculture)
- 2. B.Sc. (Food Microbiology)
- 3. B.Sc. (Medical Microbiology)

A. VOCATIONAL COURSES (MODEL II)

Semester	Course No.	AQUACULTURE	FOOD MICROBIOLOGY	MEDICAL MICROBIOLOGY
1	Ι	Principles and Methods in Aquaculture	General Microbiology	Fundamentals of Microbiology
1	II	Hatchery and Culture Techniques	Biological Techniques	Basics of Microbial Physiology & Genetics
1	Practical - I	Principles and Methods in Aquaculture & Hatchery and Culture Techniques	General Microbiology & Biological Techniques	Fundamentals of Microbiology and Basics of Microbial Physiology & Genetics
2	III	Capture Fishery	Microbial physiology & Genetics	Parasitology
2	IV	Biology of Fishes	Applied Microbiology	Medical Virology
2	Practical - II	Capture Fishery & Biology of Fishes	Microbial physiology Genetics &Applied Microbiology	Parasitology & Medical Virology
3	V	Fisheries Environment	Dairy Microbiology	Medical Mycology
3	Practical - III	Fisheries Environment	Dairy Microbiology	Medical Mycology
3	VI	Fish Nutrition	Industrial Microbiology	Diagnostic Microbiology
3	Practical - IV	Fish Nutrition	Industrial Microbiology	Diagnostic Microbiology
4	VII	Reproductive Physiology and Endocrinology	Basic Food Micro biology	Medical Bacteriology
4	Practical - V	Reproductive Physiology and Endocrinology	Basic Food Micro biology	Medical Bacteriology
4	VIII	Microbiology, Pathology and Post Harvest Technology	Food Microbiology and Fermented Food	Clinical Microbiology
4	Practical - VI	Microbiology, Pathology and Post Harvest Technology	Food Microbiology and Fermented Food	Clinical Microbiology

b. CONSOLIDATED DISTRIBUTION OF HOURS FOR BSc. (MODEL II VOCATIONAL COURSES)

COURSE STRUCTURE

Total Credits	: 120
Total Instructional Hours	: 150
3 Vocational Programmes	

: Aquaculture (A) / Food Microbiology (F)/ Medical Microbiology (M)

SEMESTER I

	Course		Course	Hrs per		Marks ratio	
No	Code	Course Title	Category	week	Credits	Intl	Extl
1		English - 1	Common Course	5	4	1	4
2		Second Language – 1	Common Course	5	4	1	4
3	ZY1CRT01	General perspectives in Science & Protistan Diversity	Core Course I : Theory	2	2	1	4
4		General perspectives in Science & Protistan Diversity	Core Course I : Practical	2	0	0	0
5		Principles and Methods in Aquaculture (A) / General Microbiology (F)/Fundamentals of Microbiology(M)	Vocational Course I : Theory	2	2	1	4
6		Hatcery & Culture techniques (A) / Biological technique (F)/Basics of Microbial physiology & Genetics		2	2	1	4
7	(M) Principles and Methods in Aquaculture, Hatchery and Culture techniques (A)/ General Microbiology and Biological technique (F)/Fundamentals of Microbiology,Basics of Microbial physiology		Vocational Course : Practical	2	0	0	0

	Ċ	&Genetics(M)				
8		Complementary Course – I	Complementary Course: Theory	3	2	
9	Practical		Practical	2	0	
		Total		25 hrs	16	

SEMESTER II

	Course		Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1		English 2	Common Course	5	4	1	4
2		Second Language -2	Common Course	5	4	1	4
3	ZY2CRT02	Animal Diversity- Non Chordata			1	4	
4	ZY2CRP01	Animal Diversity – Non Chordata			1	4	
5		Capture Fishery (A) /Microbial physiology & Genetics(F)/Parasitology (M)	Vocational Course III : Theory	2	2	1	4
6		Biology of Fishes (A) / Applied Microbiology (F)/Medical virology(M)	Vocational Course III : Theory	2	2	1	4
7		Capture Fishery &Biology of Fishes(A)/Microbial Voo physiology &Applied Cou Microbiology(F)/Parasito Pro logy&MedicalVirology (M)		2	2	1	4
8		Complementary Course Complement – II Course: The		3	2		
9		Practical	Practical	2	2		
	rmested II	Total		25 hrs	22		

SEMESTER III

No

Course

Course Hrs p

	Code		Category	week		Intl	Extl
1		English 3	Common Course	5	4	1	4
2	ZY3CRT03	Animal Diversity – Chordata	Core Course III : Theory	3	3	1	4
3		Animal Diversity- Chordata		2	0	0	0
4		Fisheries Environment (A) / Dairy Microbiology (F) / Medical mycology(M)	Vocational Course V : Theory	2	2	1	4
5		Fisheries Environment (A) / Dairy Microbiology (F) / Medical mycology(M)	Vocational Course : Practical	3	0	0	0
6		Fish nutrition (A) / Industrial Microbiology (F)/ Diagnostic Microbiology (M)	Vocational Course VI : Theory	2	2	1	4
7	Fish nutrition (A) /		Vocational Course : Practical	3	0	0	0
8		Complementary Course – III	Complementary Course: Theory	3	3	1	4
9		Practical	Practical	2	0	0	0
		Total		25 hrs	14		

SEMESTER IV

	Course		Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1		English - 4	Common Course	5	4	1	4
2	ZY4CRT04	Research methodology, Biophysics & Biostatistics	Core Course IV : Theory	3	3	1	4
3	ZY4CRP02	Research methodology, Biophysics & Biostatistics	Core Course IV : Practical	2	2	1	4
4	Reproductive Physiology and Endocrinology (A)/		Vocational Course VII : Theory	2	2	1	4
5		Reproductive Physiology and Endocrinology (A)/ Medical Bacteriology (M) / Basic Food Microbiology (F)	Vocational Course : Practical	3	4	1	4
6	Microbiology (1) Microbiology , Pathology and Post Harvest Technology (A)/ Clinical Microbiology (M) / Food Microbiology and Fermented Food (F)		Vocational Course VIII : Theory	2	2	1	4
7	Microbiology, Pathology and Post Harvest Technology (A)/		Vocational Course : Practical	3	4	1	4
8		Complementary Course - IV	Complementary Course: Theory	3	3	1	4
9		Practical	Practical	2	2	1	4
		Total		25 hrs	26		

SEMESTER V

	Course		Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1	ZY5CRT05	Environmental Biology & Human rights	Core Course V : Theory	3	3	1	4
2	ZY5CRPO5	Environmental Biology & Human rights	Core Course V : Practical	2	0	0	0
3	ZY5CRT06	Cell Biology & Genetics	Core Course VI : Theory	3	3	1	4
4	ZY5CRPO6	Cell Biology & Genetics	Core Course VI : Practical	2	0	0	0
5	ZY5CRT07	Evolution, Ethology & Zoogeography	Core Course VII : Theory	3	3	1	4
6	ZY5CRPO7	Evolution, Ethology & Zoogeography	Core Course VII : Practical	2	0	0	0
7	ZY5CRT08	Human Physiology, Biochemistry & Endocrinology	Core Course VIII : Theory	3	3	1	4
8	ZY5CRPO8	Human Physiology, Biochemistry & Endocrinology	Core Course VIII : Practical	2	0	0	0
	ZY5OPT01	1 – Vocational Zoology (Apiculture, Vermiculture, Ornamental fish culture)	Open Courses for other				
9	ZY5OPT02	2 – Public health and Nutrition	streams (Select any one out of three)	4	3	1	4
	ZY5OPT03	3 – Man, nature & Sustainable Development					
10	Group activity and Field Study - Report to be submitted in 6 th Semester along with Project and Practical Exam			1	0		
		Total		25 hrs	15		

Semester VI

No	Course	Course Title	Course	Hrs per	Credits	Mark	s ratio
No.	Code	Course Thie	Category	week	Credits		F (1
1	ZY6CRT09	Developmental Biology	Core Course IX : Theory	3	3	Intl 1	Extl 4
2	ZY6CRP09	Developmental Biology	Core Course IX : Practical	2	2	1	4
3	ZY6CRT10	Microbiology & Immunology	Core Course X : Theory	3	3	1	4
4	ZY6CRP10	Microbiology & Immunology	Core Course X : Practical	2	2	1	4
5	ZY6CRT11	Biotechnology, Bioinformatics and Molecular Biology	Core Course XI : Theory	3	3	1	4
6	ZY6CRP11	Biotechnology, Bioinformatics and Molecular Biology	, Core Course XI		2	1	4
7	ZY6CRT12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course XII : Theory	3	3	1	4
8	ZY6CRP12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course XII : Practical	2	2	1	4
	ZY6CBT01	Elective 1: Ecotourism & Sustainable Development					
	ZY6CBT02	Elective 2: Agricultural pest management	Choice Based				
9	ZY6CBT03	Elective 3: Vector & Vector borne Diseases	Core Elective Courses (Select any one out of four)	4	3	1	4
	ZY6CBT04	Elective 4: Nutrition, Health & life style management					
10	ZY6PRP01	Project Work	Project	1	2		
11	11ZY60JP01OJ (On Job Training for 14 days)		OJ		2		
		Total		25 hrs	27		

Model II Programmes in Zoology and Core Courses

Instructional Hours, Credit, Total Instructional Hours, University Examination, Weightage Internal and External Evaluation of Core Courses will follow the same pattern as in Model 1 Zoology Programme.

For Vocational Courses also University Examination will be conducted at the end of each Semester for Theory and at the end of even semester for Practical. Duration of examination is 3 hrs and Internal-External marks ratio is 1:4. The main objective of 'on the job training' is to acquaint the students formally to a real life work place environment. This will help to explore the relationship between knowledge and skill acquired in the college with those required in the working situations. Students are expected to do the OJ in their respective vocational subject related field.

		Weightage ratio					
Semester	Semester Course		eory]	Pract.		
		Ext.	Int.	Ext.	Int.		
1	1	4	1				
1	2	4	1	4	1		
2	3	4	1				
2	4	4	1				
3	5	4	1	4	1		
3	6	4	1				
4	7	4	1	4	1		
4	8	4	1				

C. SCHEME OF EXAMINATION - VOCATIONAL SUBJECTS

i. SCHEME OF EXAMINATION - THEORY (VOCATIONAL SUBJECT – AQUACULTURE)

Semester	Code	Course No.	Course	Hrs		External tio
					Internal	External
1	ZA1VOT01	1	Principles and Methods in Aquaculture		1	4
1	ZA1VOT02	2	Hatchery and Culture Techniques	3	1	4
2	ZA2VOT03	3	Capture Fishery	3	1	4
2	ZA2VOT04	4	Biology of Fishes	3	1	4
3	ZA3VOT05	5	Fisheries Environment	3	1	4
3	ZA3VOT06	6	Fish Nutrition	3	1	4
4	ZA4VOT07	7	Reproductive Physiology and Endocrinology	3	1	4
4	ZA4VOT08	8	Microbiology, Pathology and Post Harvest Technology.	3	1	4

SCHEME OF PRACTCAL

EXAMINATION (AQUACULTURE)

Practical Exams at the end of 2nd& 4th semesters

Semester	Code	Course	Exam duration	Internal External ratio		
				Internal	External	
2	ZA2VOP01	Practical 1 & 2 Principles and Methods in Aquaculture, Hatchery and Culture Techniques & Capture Fishery and Biology of Fishes	3 hrs	1	4	
	ZA4VOP02	Practical 3 & 4 Fisheries Environment & Fish nutrition.	3 hrs	1	4	
4	ZA4VOP03	Practical 5 & 6 Reproductive physiology, Endocrinology& Microbiology, Pathology and Post Harvest Technology 3hrs		1	4	

ii. SCHEME OF EXAMINATION - THEORY (VOCATIONAL SUBJECT – FOOD MICROBIOLOGY)

Semester	Code	Course No.	Course	Hrs		External
					Internal	External
1	ZF1VOT01	1	General Microbiology	3	1	4
1	ZF1VOT02	2	Biological Techniques	3	1	4
2	ZF2VOT03	3	Microbial Physiology & Genetics	3	1	4
2	ZF2VOT04	4	Applied Microbiology	3	1	4
3	ZF3VOT05	5	Dairy Microbiology	3	1	4
3	ZF3VOT06	6	Industrial Microbiology	3	1	4
4	ZF4VOT07	7	Basic Food Microbiology	3	1	4
4	ZF4VOT08	8	Food Microbiology and Fermented Food	3	1	4

SCHEME OF PRACTICAL EXAMINATION (FOOD MICROBIOLOGY) Practical Exams at the end of 2nd& 4th semesters

Semester	Code	Course	Exam duration		External atio
				Internal	External
2	ZF2VOP01	Practical 1 & 2 General Microbiology & Biological Techniques,Microbial Physiology, Genetics & Applied Microbiology	3 hrs	1	4
	ZF4VOP02	Practical 3 & 4 Dairy Microbiology & Industrial microbiology	3 hrs	1	4
4	ZF4VOP03	Practical 5 & 6 Basic Food Microbiology, Food Microbiology and Fermented Food	3 hrs	1	4

iii. SCHEME OF EXAMINATION - THEORY (VOCATIONAL SUBJECT – MEDICAL MICROBIOLOGY)

Sem.	Code	Course No.	Course	Hrs	Internal	External ratio
					Internal	External
1	ZM1VOT01	1	Fundamentals of Microbiology	3	1	4
1	ZM1VOT02	2	Basics of Microbial3Physiology4& Genetics4		1	4
2	ZM2VOT03	3	Parasitology	3	1	4
2	ZM2VOT04	4	Medical Virology	3	1	4
3	ZM3VOT05	5	Medical Mycology 3 1		4	
3	ZM3VOT06	6	Diagnostic Microbiology 3 1		1	4
4	ZM4VOT07	7	Medical Bacteriology	3	1	4
4	ZM4VOT08	8	Clinical Microbiology	3	1	4

SCHEME OF PRACTICAL EXAMINATION (MEDICAL MICROBIOLOGY) Practical Exams at the end of 2nd & 4th semesters

Sem.	Code	Course	Exam duration	Internal External rat	
				Internal	External
2	ZM2VOP01	Practical 1 & 2 Fundamentals of Microbiology, Basics of Microbial Physiology & Genetics, Parasitology & Medical virology	3 hrs	1	4
4	ZM4VOP02	Practical 3 & 4 Medical Mycology & Diagnostic Microbiology	3 hrs	1	4
	ZM4VOP03	Practical 5 & 6 Medical Bacteriology and Clinical Microbiology	3 hrs	1	4

d. SCHEME OF DISTRIBUTION OF HOURS AND CREDITS (VOCATIONAL SUBJECT – AQUACULTURE)

Semester	Code	Course No.	Title of vocational course	Hrs	Inst. Hrs⁄Wk	Credit
1	ZA1VOT01	1	Principles and Methods in Aquaculture	36	2	2
1	ZA1VOT02	2	Hatchery and Culture Techniques	36	2	2
1		Practical -1	Principles and Methods in Aquaculture, Hatchery and Culture Techniques.	36	2	0
2	ZA2VOT03	3	Capture Fishery	36	2	2
2	ZA2VOT04	4	Biology of Fishes	36	2	2
2	ZA2VOP01	Practical - 2	Capture Fishery and Biology of Fishes	36	2	2
3	ZA3VOT05	5	Fisheries Environment	36	2	2
3		Practical - 3	Fisheries Environment	54	3	0
3	ZA3VOT06	6	Fish Nutrition	36	2	2
3		Practical - 4	Fish Nutrition	54	3	0
4	ZA4VOT07	7	Reproductive Physiology and Endocrinology	36	2	2
4	ZA4VOP02	Practical – 5	Reproductive Physiology and Endocrinology	54	3	4
4	ZA4VOT08	8	Microbiology, Pathology and Post Harvest Technology.	36	2	2
4	ZA4VOP03	Practical - 6	Microbiology, Pathology and Post Harvest Technology	54	3	4
			TOTAL		32	26

SCHEME OF DISTRIBUTION OF HOURS AND CREDIT (VOCATIONAL SUBJECT – FOOD MICROBIOLOGY)

Semester	Code	Course No.	Title of vocational course	Hrs	Inst. Hrs⁄Wk	Credit
1	ZF1VOT01	1	General Microbiology	36	2	2
1	ZF1VOT02	2	Biological Techniques	36	2	2
1		Practical -1	General Microbiology & Biological Techniques	36	2	0
2	ZF2VOT03	3	Microbial Physiology & Genetics	36	2	2
2	ZF2VOT04	4	Applied Microbiology	36	2	2
2	ZF2VOP01	Practical - 2	Microbial physiology, Genetics & Applied Microbiology	36	2	2
3	ZF3VOT05	5	Dairy Microbiology	36	2	2
3		Practical - 3	Dairy Microbiology	54	3	0
3	ZF3VOT06	6	Industrial Microbiology	36	2	2
3		Practical - 4	Industrial Microbiology	54	3	0
4	ZF4VOT07	7	Basic Food Microbiology	36	2	2
4	ZF4VOP02	Practical – 5	Basic Food Microbiology	54	3	4
4	ZF4VOT08	8	Food Microbiology and Fermented Food	36	2	2
4	ZF4VOP03	Practical - 6	Food Microbiology and Fermented Food	54	3	4
			TOTAL		32	26

SCHEME OF DISTRIBUTION OF HOURS AND CREDIT (VOCATIONAL SUBJECT – MEDICAL MICROBIOLOGY)

Semester	Code	Course No.	Title of vocational course	Hrs	Inst. Hrs⁄Wk	Credit
1	ZM1VOT01	1	Fundamentals of Microbiology	36	2	2

1	ZM1VOT02	2	Basics of Microbial Physiology & Genetics	36	2	2
1		Practical -1	Fundamentals of Microbiology, Basics of Microbial Physiology & Genetics	36	2	0
2	ZM2VOT03	3	Parasitology	36	2	2
2	ZM2VOT04	4	Medical Virology	36	2	2
2	ZM2VOP01	Practical - 2	Parasitology &Medical Virology	36	2	2
3	ZM3VOT05	5	Medical Mycology	36	2	2
3		Practical - 3	Medical Mycology	54	3	0
3	ZM3VOT06	6	Diagnostic Microbiology	36	2	2
3		Practical - 4	Diagnostic Microbiology	54	3	0
4	ZM4VOT07	7	Medical Bacteriology	36	2	2
4	ZM4VOP02	Practical – 5	Medical Bacteriology	54	3	4
4	ZM4VOT08	8	Clinical Microbiology	36	2	2
4	ZM4VOP03	Practical - 6	Clinical Microbiology	54	3	4
			TOTAL		32	26

e. SYLLABUS (VOCATIONAL SUBJECTS) – MODEL II

THEORY & PRACTICAL

1. AQUACULTURE

SEMESTER 1

VOCATIONAL COURSE 1

PRINCIPLES AND METHODS IN AQUACULTURE

Objectives:

- 1. To make the student understand the basic concepts of the science and practice of aquaculture.
- 2. To introduce the student to the diverse practices of culturing of aquatic organisms.
- 3. The encourage the student to take up aquaculture practices as a vocation.

Module 1

Scope and importance of Aquaculture. Significance of aquaculture compared to other agricultural systems and commercial fisheries. Types of aquaculture - Freshwater, Brackish water and Mariculture. Shell fish culture, Finfish culture, Monoculture, Polyculture.

Module 2

Integrated farming – The concept of recycling of organic waste for maximum production. Rice cum fish culture. Culture practices of duck cum fish, poultry cum fish and pig cum fish culture. Sewage fed Fish Culture.

Module 3

Site selection parameters for a pond site. Soil quality parameters -physical, soil type, porosity, percolation, shear strength, rate of compassion etc. Chemical -salinity, pH, nutrients, toxic gases etc. Water quality parameters-Chemical- salinity, pH, dissolved oxygen, Carbon dioxide, Nutrients, Ammonia. Physical - Temperature, suspended solids, . Biological parameters-presence of juveniles/seedlings, predators/ competitors, plankton, nekton.

Module 4

Pond construction – Selection of site. Preparation of bunds and dykes. Calculation of earth works, sluice gates- Different types and fixing of sluice gates.Pond preparation-Drying, elimination of pests and predators. Preparation of nursery and stocking ponds. Manuring, Production of plankton. Stocking and acclimatization. Use of hapa. Stocking density. Harvesting and Harvesting methods

Module 5

7 Hrs

4 Hrs

3 Hrs

36 Hrs Credits 2

10 Hrs

Fresh water cultivable fishes and their external characters-Indian Major Carps, Catfish, Eel, Tilapia, Etroplus, Trouts, Mahseer, Channa, Clarius, Anabas, Larvivorous and Weed fishes. Cultivable species of crustaceans and mulluscs: Identification and external characters. Shrimp, freshwater prawn, crab, lobster, pearl oyster, edible oyster, mussel and clams.

Module 6

2 Hrs

Brackish water aquaculture-Introduction, the tidal range, salinity and biota. Major reservoirs of India and their fishery- Constarints in reservoir fisheries.

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VOCATIONAL COURSE 2

HATCHERY AND CULTURE TECHNIQUES

Objectives

- 1. Introduce the student to the culture practices of various indigenous edible and ornamental finfishes and edible shellfishes.
- 2. To create an understanding of the different technologies of seed production of common cultivable species
- 3. Introduce the student to various live feeds available for aquatic organisms and understand their culture methods.

Module 1

Hatchery systems - Different types- fin fish (Carp), Mollusc (Edible oyster)- Seed collection,

Spat collectors. Crustacean (Penaeid prawn)- Different Components and operation, Description

of Larval stages.

Module 2

Culture Practices of major groups of finfishes. Indian Major Carps- Nursery, Rearing and stocking ponds. Preparation of ponds. Stocking and post stocking management. Harvesting. Culture of air breathing fishes(eg. Channa).

Culture of Tilapia and Milk Fish, (Mention lablab and its preparation) production of Monosex in Tilapia.

Culture of Trouts.

Module 3

Culture of Crustaceans - Penaeid prawns- Seed resources, prawn filtration practices, (Pokkali, Bheries Ghazan lands) ,extensive, semi intensive and intensive, prawn farming. Crab culture- Crab fattening and growout.

Module 4

Culture of Bivalve mollusks.- Mussels, Pearl oysters, Edible oysters, Clams. On Bottom and Off bottom culture methods- Stake culture (Bouchot culture), Rack culture (Rack and ren, Rack and tray), Long Line culture and Raft culture.

Module 5

8 Hrs

8 Hrs

8 Hrs

6 Hrs

36 hrs Credits 2

Culture of ornamental fishes- Types of Aquaria, Aquarium accessories. Setting up and maintenance of Aquaria. Filtration of Aquarium water- Mechanical, Chemical and Biological filtration. Breeding techniques of Aquarium fishes;- gold fish (Egg scatterer) and Fighter fish (Bubble nest builder)

Culture of sea weeds and holothurians.

Culture of live feeds- micro algae, artemia, rotifer, daphnia.

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- 14. Seaweed resources of India CMFRI Bulletin, No. 41, 1987

SEMESTER II

VOCATIONAL COURSE 3

CAPTURE FISHERY

36 hrs Credits 2

Objectives:

- 1. Introduce the student to the fishery potential of Indian waters- marine and inland.
- 2. To study the major groups of finfishes and shellfishes contributing to the commercial fishery
- 3. Introduction to fishing gears and diverse fish catching methods of tropical waters.
- 4. To develop the concept of fishery as a renewable resource to be managed and study of population dynamics and different management techniques in commercial fisheries.

Module 1

8 Hrs

Commercially important orders, families, genera and species of elasmobranches and teleost of the Indian region and their identification. Identification of commercially important species of prawn, crab, lobster, bivalve, gastropod and cephalopods.

Module 2

Craft and gear-Types of fishing craft in India –Traditional, Motorised and Mechanized. Classification of fishing gear. Fishing gear material- Synthetic and natural. Properties of fishing gear, floats, sinkers and accessories. Major fishing gears and their operation. Static gear –Gill nets, Long line and Fish traps. Mobile gear-Drag nets-Trawl nets. Seine nets-Purse seine, Shore seines.

Module 3

Inland capture fishery resources of India – Riverine fisheries, Cold water fisheries resources, Lacustrine fisheries (Estuarine or brackish water fisheries and fisheries of fresh water lakes)-Important species and gears.

Module 4

Marine Fishery Resources of India- Pelagic fishery- Fisheries of oil sardine, lesser sardine, anchovies, mackerel, ribbon fishes. Demersal fisheries. Fisheries of elasmobranchs, Bombay duck, cat fishes, silver bellies, sciaenids, pomfrets, threadfins, threadfin breams and other perches, Flatfishes, Prawns, Lobsters, Crabs, Mussels, Oysters and Clams - Fishing seasons, abundance and major gears for each group.

Module 5

6 Hrs

138

8 Hrs

6 Hrs

Fishery management- Concept of population, Yield and Recruitment. Factors affecting fish stocks. Population dynamics. Problems of over fishing- Growth overfishing and Recruitment overfishing.Fishery. Management practices- Input and output controls. Maximum Sustainable Yield.

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SEMESTER II

VOCATIONAL COURSE 4

BIOLOGY OF FISHES

36 hrs Credits 2

Objectives:

- 1. To create an understanding about the morphological and anatomical organization of finfishes and shellfishes.
- 2. Introduce the student to the basic principles of Taxonomy of cultivable organisms.
- 3. To have an awareness of the fundamental biological aspects of food and feeding, age and growth and reproductive biology.

Module 1

Need for taxonomy, binomial nomenclature, Data requirements for classification of fishes, Methods for collection of taxonomic data- Morphologic and Meristic data., Study of external morphology of a typical elasmobranch and a typical teleost, Variations in form and structures used in taxonomic studies.

Module 2

Internal anatomy of fish- Alimentary canal and associated structures. Gills, Swim bladder, Accessory respiratory organs, Heart and circulatory system, skeletal system (Visceral arches, Vertebral column and skeleton of fins only) Nervous and lateral line system, sense organs (eye, ear, olfactory organs)

Module 3

12 Hrs

Excretion, osmotic and ionic regulation in marine and freshwater fishes. Swimming activity. Types of locomotion- Anguilliform, Carangiform and Ostraciform. Muscles in locomotion. Parental care, Social behaviour- Aggregation and shoaling. Migration of fishes.

Module 4

6 Hrs

Natural food of fishes. Feeding habits and types of feeding in fishes- Carnivorous, Herbivorous and Omnivorous, Predators, Grazers, Suckers, Strainers and parasites.. Feeding habits and method of feeding in prawn, bivalve and cephalopod.

Module 5

8 Hrs

Growth of fishes- Absolute and relative growth, isometric growth and allometric growth. The cube law. Methods for determination of growth checks. Length frequency analysis. Analysis of growth using hard parts like scales, otoloiths and vertebrae. Estimation of growth by direct methods. Marking and tagging of fish for growth studies. Methods of studying reproduction-Maturity Stages, Gonadosomatic Index, Ova Diameter Frequency studies. Detemination of size at first maturity and spawning season, Fecundity and its determination.

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SEMESTER III

VOCATIONAL COURSE 5

FISHERIES ENVIRONMENT

36 hrs Credits 2

Objectives

- 1. To study the environment and their effect on fish populations.
- 2. Study the use of Remote sensing techniques for the assess fish stocks
- 3. Introduction to the ancillary marine resources like seaweeds, echinoderms and corals.
- 4. To understand the different techniques and equipments for the study of environmental parameters and different fish finding devices.

Module 1

Habitat Ecology. Freshwater Habitat- Lentic (Pond, Lake), Lotic (Riffles, Pools). Marine Habitat- Zonation, Biota and adaptations. Principles of limiting factors- Shelford's law of tolerance, Liebig's law of minimum, Combined concept. Ecological succession and Ecological indicators. Photosynthetic and saprophytic food chain.

Module 2

Basic marine metereology- weather, air-sea interactions. Monsoons, seasonal changes, Circulation of water masses, Waves, Tides and sediment transportation.

Module 3

Physical and chemical parameters of Aquatic environments- temperature, salinity, oxygen, nutrition, Microelements and Macroelements. Phytoplankton and primary production- Methods of Estimation (Dark and Light bottle method, C14 Method, Chlorophyll Technique). Estimation of Zooplankton and secondary production.

Module 4

Ecology of estuaries and mangroves- Soil, Water and Biota. Biogeochemical cycles-Nitrogen, Phosphorus and Sulphur cycle.

Module 5

Ancillary marine resources-Sea weeds, corals, Echinoderms and their commercial importance. Aquatic pollution- Causes and Remedial Measures. Instruments used for Biological sampling-Plankton International Indian Ocean Expedition Net, Hardy's continuous plankton recorder), Nekton (Isaac Kidd's Midwater trawl), Benthos (Dredges, Grabs, Agassiz trawl). Fish finding devices- Echo sounder, Sonar and net sonde . Remote sensing techniques and application.. Satellite remote sensing of fish stocks- Ocean colour maps, Sea surface Temperature Contour Maps. Potential Fishing Zone (PFZ). Deep Scattering or Sonic Scattering Layer.

References

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10 Hrs

4 Hrs

10 Hrs

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SEMESTER III

VOCATIONAL COURSE 6

FISH NUTRITION

Objectives:

1. To create an understanding on the nutritional needs of aquatic organisms in culture.

- 2. To make the student have a basic concept of energy budgeting, food additives and varieties of feed ingredients used in Aquafeeds.
- 3. To have a basic understanding of the principles of feed formulation and equipments used in feed manufacture.

Module 1

Digestive system of fish, Digestive Glands and their secretions. Digestive physiology of fish-Digestion of Carbohydrates, Proteins and Fats. Proteins, carbohydrates, fats, vitamins and minerals in fish nutrition- Classification, Structure and functions.

Module 2

Nutritional Bioenergetics- Gross energy, Digestible energy (Digestibility co-efficient and measurement of digestible energy), Metabolisable energy, Heat increment (Specific Dynamic action) Net energy, Retained energy, Protein utilization. (Protein Efficiency Ratio, Protein

8 Hrs

36 hrs Credits 2

conversion ratio, Productive protein value) .Proximate Analysis- Moisture, Crude protein, Crude Lipid, Crude Fibre, Ash and , Nitrogen free extract- Methods of analysis of each component.

Module 3

Factors affecting digestibility, Nitrogen balance index, Food additives- Binders, Antioxidants, Chemo-attractants and feeding stimulants, Pigments, Antimicrobial agents and Anabolic agents. Non conventional feed stuffs. Food growth equation. Feed ingredients of plant and animal origin.

Module 4

Feed preparation techniques. Factors affecting the energy requirement of fish. Non nutrient constituents of the diet. Measurement of calorific value- Component Analysis, Wet oxidation, Bomb Calorimetry. Types of feeds (Wet, Moist and Dry feeds-advantages and disadvantages) Larval feeds- Minced diets, Microparticulate diets, Spray dried diets, Microbound diets, Microcoated diets and Microencapsulated diets.

Module 5

10 Hrs

Growth promoters. Principles of feed formulation (Pearson's Square and Linear programming). Different systems of fish feeding or Fish feeding devices. Equipments used in Feed mills-Weighing Scales, Grinders, Mincers, Mixers, Elevators, Extruders, Coolers/Dryers, Fat sprayer, Crumbler, Sifter and Bag seamer. Food Conversion Ratio and Food efficiency ratio. Economics of feed preparation.

References

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8 Hrs

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SEMESTER IV

VOCATIONAL COURSE 7

REPRODUCTIVE PHYSIOLOGY AND ENDOCRINOLOGY

36 hrs Credits 2

Objectives:

- 1. To have an understanding of the variety of reproductive techniques in finfishes and shellfishes and the factors controlling reproduction.
- 2. To study the endocrine and neurosecretory system of finfishes and shellfishes.
- 3. To understand the principles and techniques of induced breeding, and cryopreservation of fish gametes.

Module 1

6 Hrs

Reproductive systems and Sexual dimorphism in fish, crab and prawn. Types of reproduction-Viviparity, ovoviviparity, oviparity in Teleosts and Elasmobranchs. Classification of maturity stages of ovary and testes in fishes and prawns. Oogenesis and spermatogenesis in fishes. Hermaphroditism- different types. Sex reversal and sex determination in fishes.

Module 2

Organisation, structure and Functions of Neurosecretory and endocrine systems in fin fishes. Pituitary, Thyroid, Chromaffin tissue, Interrenal tissue, Pancreatic islets, Corpuscles of Stannius, Ultimobranchial Glands, Gonads, Gastro-intestinal Hormones, Pineal organ, Caudal neurosecretory system or Urophysis. Neuroendocrine control of reproduction. Role of Hypothalamus - Pituitary – Gonadal axis in control of maturationin fishes. Gonadotropin releasing hormones, gonadotropins and sex steroids.

Module 3

6 Hrs

10 Hrs

Neuroendocrine systems in crustaceans and control of reproduction. Sinus gland complex and X- organs. Pericardial and Post-commisural organs. True Endocrine organs-Y- organs, androgenic gland and Mandibular organs. Hormones produced by the neuroendocrine and true endocrine glands and their role in the control of reproduction and moulting in Crustaceans. Parasitic castration.

Module 4

Principles of induced maturation and spawning in fishes and crustaceans. Levels of control in induced breeding and maturation in fishes. Environmental control of reproduction in fishes and prawns. Use of hormones and hormone analogues in fishes- Gonadotropin releasing hormones, Gonadotropins and Sex steroids. Methods of hormonal administration. Hypophysation. Linpe Method. Ovaprim. Use of Anaesthetics. Eyestalk ablation- Its principle and application in crustacean hatcheries. Use of hormones for producing monosex population and sex reversal in fishes. Principles and methods of cryopreservation of gametes.

Module 5

8 Hrs

Types of eggs in fishes – Pelagic, Demersal and according to yolk content. Embryonic development- Cleavage, fate map of Blastula, gastrulation- Invagination, Involution, Delamination, Convergence, Epiboly. Hatching, Post Embryonic development and Larval development.

References

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(1969). Vol 3. Reproduction and Growth, Bioluminiscence, Pigments and Poisons. (1983). Vol. 9. Part A. Reproduction, Endocrine tissues and hormones. (1983). Vol.9. Part B. Reproductive Behaviour and Fertility control.

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SEMESTER IV

VOCATIONAL COURSE 8

MICROBIOLOGY, PATHOLOGY AND POST HARVEST TECHNOLOGY

36 Hrs Credits 2

Objectives

1. To have a clear understanding of the bacterial fauna associated with fish sanitation.

- 2. To have a basic idea of the factors associated with fish spoilage and the variety of fish preservation techniques.
- 3. To clearly understand the symptoms, diagnostic and remedial measures of fish diseases and have an idea of the Critical Control Points in seafood industry.

Module 1

Bacteriology- Classification of bacteria based on temperature and oxygen requirements, Bacterial growth curve, growth phasesSterilization techniques, preparation of culture media, Estimation of total plate count, Staining techniques (Gram's Staining). Important bacteria of sanitary significance-*Staphylococcus aureus, Vibrio cholerae, salmonella*. Faecal Indicator organisms- *E.coli* and Faecal streptococcus.

Module 2

Biochemical composition of fish- Moisture, Protein, Fats, NPN compounds and Minerals. Spoilage of fish - Post mortem changes and Rigor mortis. Causes of spoilage - Enzymatic, microbial, Biochemical (rancidity). Indices of spoilage - organoleptic, chemical (Total Volatile

8 Hrs

Basic Nitrogen, Hypoxanthine content, Peroxide value and microbial (direct count and Total Plate Count).

Module 3

Processing and Preservation of fish – Chilling and Freezing- Slow freezing, and quick freezing-critical temperature. Freezer burn, thawing, drip loss and glazing. Types of Freezers-Plate Freezer, Tunnel (Air Blast) Freezer, Immersion Freezer, Fluidised bed Freezer (IQF), Cryogenic Freezing, Accelerated Freeze drying, Irradiation. Canning- Principle and Procedure. Common defects in canning- Struvite formation, Panelling, Flipper, Springer, Soft swell and Hard swell.

Module 4

Curing- Types of Curing- Simple, Sun drying, dry and wet curing, Monacuring, Pit Curing, Colombo curing, Smoke curing. Special cured products- Masmine and Marinade. Common defects in curing- Dun, Rust, Pink, Maggots. Value added products. Fishery byproducts- Fish oil, Shark liver oil, Chitin, Chitosan, Isinglass, Fish meal, Shark fin rays.

Module 5

6 Hrs

6 Hrs

Diseases of fin fishes and prawns.-Protozoan, Bacterial, viral, fungal, Crustacean, Leech, Helminth diseases. Symptoms and Remedial measures. Nutritional deficiency diseases - Pin head, Rickets, Soft Shell Syndrome, Lipoid hepatic degeneration, Vitaminosis A. Hazard analysis and critical control points in seafood industry.

References

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AQUACULTURE PRACTICALS

PRINCIPLES AND METHODS IN AQUACULTURE, HATCHERY AND CULTURE TECHNIQUES

36 hrs

Credit 1

- 1. Identification and major biological characteristics of cultivable organisms
- 2. Gut content analysis.
- 3. Study of common weed and predatory fishes in aquaculture ponds
- 4. Study of aquatic insects and aquatic weeds.
- 4. Identification of different larval stages and hatchery operations of prawn
- 6 Setting up and keeping of aquariums
- 7 Visit to carp and prawn hatcheries.

SEMESTER II

PRACTICAL - II CAPTURE FISHERY & BIOLOGY OF FISHES

36 Hrs

- 1. Identification of commercially important fishes, crustaceans and molluscs.
- 2. Fish- Study of external morphology and scales..

- 3. Dissection of Alimentary canal.
- 4. Dissecting and identification of internal organs of a fish.
- 5. Prawn- Study of external morphology and nervous system
- 6. Gill structure- Herbivorous, carnivorous and omnivorous fishes.
- 7. Gill structure of a prawn Dissection
- 8. Molluscs- Study of morphology, and Dissection of Gills of bivalves
- 9. Visit to marine fish landing centre.

SEMESTER III

PRACTICAL – III FISHERIES ENVIRONMENT 54 Hrs

- 1. Determination of salinity, dissolved oxygen, pH, total alkalinity, hardness, nitrate, nitrite and ammonia and phosphate.
- 2. Determination of soil pH
- 3. Study of common marine phytoplankton, zooplankton.
- 4. Quantitative evaluation of phytoplankton and zooplankton in culture ponds
- Identification of the common Ancillary Marine Resources Corals, Sea cucumber and Sea weeds
- Equipments and Instruments used for the collection of Environmental Data Plankton samplers and Counters including haemocytometer, Digital pH meter, Salinometer, Spectrophotometer, Colorimeter etc.
- Study of Ecological sub-divisions of the sea, Principles of Remote sensing and software used (Wikimapia.org)

SEMESTER III

PRACTICAL – IV FISH NUTRITION 54 Hrs Credit 2

- 1. Comparative study of Digestive system of Herbivorous and Carnivorous fishes
- 2. Qualitative estimation of proteins, Polysaccharides and lipids
- 3. Formulation of artificial feed for aquarium fishes and prawns with locally available ingredients.
- Study of identification feed ingredients of plant origin and animal origin (oil cakes and meals eg: Groundnut oil cake, coconut oil cake, Mustard oil cake, Fish meal, Crustacean meals, Molluscan meals, Blood meal etc)
- 5. Use of Pearson's square method in balancing feed Ingredients.
- Study of equipments used in feed preparation (Oven, Pelletiser, Feed Press and Die Plate, Extruders etc.)
- 7. Study of non-conventional feed stuffs eg. Spirulina etc. and Feed Additives (Binders, Antibiotics etc).

SEMESTER IV

PRACTICAL – V REPRODUCTIVE PHYSIOLOGY AND ENCOCRINOLOGY

54 Hrs

- 1. Dissection of reproductive organs of Teleost fish.
- 2. Dissection of reproductive organs of Prawn and Crab.
- 3. Eyestalk ablation technique and electrocautery apparatus(Demonstration)
- 4. Methods of hormone injection in fish.
- 5. Observation of larval and embryonic stages in fish egg development.
- 6. Estimation of maturity stages and fecundity in fish
- 7. Equipments used in cryopreservation (Cryocan, French straws etc)

SEMESTER IV

PRACTICAL – VI MICROBIOLOGY, PATHOLOGY AND POST

HARVEST TECHNOLOGY

54 Hrs

- 1. Sterilisation techniques, preparation of culture media (TGBE and Nutrient Agar Media), nutrient agar slants, staining techniques.(Gram staining)
- 2. Determination of total plate count
- 3. Types of bacterial colonies
- 4. Instruments used in bacteriological Studies (Inoculation chamber, Autoclave, Colony counter etc.)
- 5. Examination of internal and external organs of diseased fish and shell fishes.
- 6. Identification of parasites in fishes and shell fishes.
- 7. Materials used in fish processing and packaging (Cans, Retortable pouches etc.)

ZOOLOGY - MODEL II

VOCATIONAL SUBJECT: 2. FOOD MICROBIOLOGY SYLLABUS -THEORY & PRACTICALS

SEMESTER 1 ZF1VOT01 - VOCATIONAL COURSE 1 GENERAL MICROBIOLOGY

36 Hrs

2 Credits

Objectives

- 1. To introduce students to the fascinating world of microbes
- 2. To describe the differences in prokaryotic and eukaryotic cell morphology and structure
- 3. To impart knowledge on major categories of microorganisms and analyze their classification and diversity.

Module I

The historical development of microbiology, Prokaryotes and Eukayotes, Principles of microbial taxonomy, Classification of bacteria according to Bergy's manual, classification based on molecular techniques-G+C % and RNA, DNA hybridization.

Module II

Morphology and fine structure of bacteria, size, shape and arrangements.Flagella, pili, capsule, cell wall and its composition.Cytoplasmic membrane, protoplasts, spheroplasts, intracellular membrane systems, cytoplasm, vacuoles, nuclear material, bacterial spores, cell inclusions.

Module III

Morphology and fine structure of virus, size, shape capsid and capsomeres.Capsid symmetryhelical, icosahedral and complex, Srtucture of TMV.Viral multiplication-lytic and lysogenic cycle.

Module IV5 HrsGeneral characters, Morphology and fine structure of of fungi.3 HrsModule IV3 HrsGeneral characters of algae and protozoa

10 Hrs

10 Hrs

References

- 1. Alcamo Fundamentals of microbiology, 5th Edition,.
- 2. Ananthanarayan&Paniker.Text book of Microbiology, 7th Edition, Orient Longman.
- 3. David Freifelder, Molecular biology, Narosa Publishing House
- 4. Jacquelyn G. Black, Microbiology: Principles and Explorations, 9th Edition, Willey Publication
- 5. M.J. Pelczar, E.C.S Chan & N.R. Kreig, Microbiology, 5th Edition, Tata McGraw Hill Edition.
- Michael T. Madigan, Brock Biology of Microorganisms,13th Edition, Amazon Publication
- 7. R.P. Singh, Microbiology, 1st Edition, Kalyani Publishers.
- 8. GeraldJ.Tortora,Berdell.R.Funke,ChristineL.case,Microbiology–An Introduction.Pearson Publication.

SEMESTER 1

ZF1VOT02 - VOCATIONAL COURSE 2

BIOLOGICAL TECHNIQUES

36 Hrs 2 Credits

Objectives

- 1. To describe the parts and working principle of instruments used in microbiology
- 2. To introduce other techniques used in microbiology

3. To learn the applications of the instruments and techniques in the bio-medical field.

Module I

Specimen preparation for light microscopy – wet mount preparation, hanging drop method, smear preparation. Bacterial staining -simple staining, Differential staining:- Gram's staining, acid fast staining, Special staining:- capsule staining, spore staining, negative staining – Indian ink staining. Specimen preparation for Electron microscopy.

Module II

Microscopy: - light microscopy, bright field, Dark field, phase contrast microscopy, fluorescence, Electron microscopy- SEM, TEM, STEM, Newer techniques in microscopy- confocal, scanning probe microscopy.

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8 Hrs

Module III

Introduction to immunology Techniques - Tests for Antigen Antibody reactions, Immuno diffusion, Radio Immuno Assay, ELISA, Flow cytometer

DNA/RNA Amplification techniques- Polymerase Chain Reaction(PCR) and RT-PCR; principle, procedure and application.

Module IV

Control of microorganisms- Sterilization and disinfection - physical (moist and dry heat, by filtration, by irradiation) & chemical methods (alcohols, phenol, detergents, halogens, dyes);

Module V

Chemotherapeutic agents-antibacterial-Antibiotic, antiviral-interferon, antifungal. Tests for antibiotics-Kirby bauer and tube dilution.

References

- 1. Janis Kuby, Immunology
- 2. Beacker&Deamer, The World of cell,
- Benson, Microbiological application laboratory manual in general microbiology, 8th 3. Edition. McGraw Hill Publication.
- 4. Jacquelyn G. Black, Microbiology: Principles and Explorations, 9th Edition, Willey Pub.
- Joanne M. Willey, Prescott, Harley & Klein's Microbiology, 7th Edition by McGraw & Hill 5. Pub.
- 6. M.A. Subramanian Biophysics principles and techniques, MJP Publishers
- 7. P.K. Bajpai, Biological instrumentation and methodology, S. Chand & Company
- 8. Upadhyay, Upadhyay, Nath, Biophysical Chemistry Principles & techniques, 4th Edition 2007, Himalaya Publishing House.
- 9. Gerald J.Tortora, Berdell.R. Funke, Christine L.case, Microbiology - An Introduction. Pearson Publication.

COURSE TITLE- GENERAL MICROBIOLOGY & BIOLOGICAL TECHNIQUES PRACTICAL 1

36 hrs

- 1. Cleaning and sterilization of glassware
- 2. **Bacterial Staining Method**
 - a. Simple Staining
 - b. Gram's staining

8 Hrs

6 Hrs

0 Credit

- c. Spore staining
- d. Negative staining
- 3. Examination of microbes in Living condition
 - a. Wet mount
 - b. Hanging drop method
- 4. Measurement of bacterial growth by optical density method
- 5. Instrumentation and working principle of
 - a. Laminar air flow Bench
 - b. Autoclave
 - c. Hot air oven
 - d. Colony counter
- 6. Antibiotic sensitivity test

SEMESTER 2

ZF2VOT03 - VOCATIONAL COURSE 3

MICROBIAL PHYSIOLOGY AND GENETICS

36 Hrs 2 Credits

Objectives

- 1. To understand the fundamentals of Physiology of microorganisms.
- 2. To define the science of microbiology and describe some of the general methods used in the study and culture of microorganisms
- 3. To study mutation and genetics in microorganisms.

Module I

Microbial Nutrition- Nutritional requirements and nutritional grouping of microbe. Bacterial growth, Bacterial growth curve (Batch), Fed batch, continuous culture of microbes, Influence of environmental factors on microbial growth.

Module II

Culture Media & Cultivation of Bacteria (Aerobic & Anaerobic culture methods).

Module III

Measurement of microbial size and numbers.

Module IV

8 Hrs

7 Hrs

5 Hrs

8 hrs

Mutation:- Type of mutation-Point and Frame shift mutation. Chemical and physical mutatgens.

Module V

Bacterial Genetics- Plasmids, cosmid, Bacterial recombination- conjugation, transformation and transduction.

References

- 1. Alcamo Fundamentals of microbiology, 5th Edition,.
- 2. Ananthanarayan&Paniker.Text book of Microbiology, 7th Edition, Orient Longman.
- Ronald M. Atlas and Richard Bartha, Microbial ecology, Fundamentals & applications 4th edition, Pearson Publication.
- 4. GeraldJ.Tortora, Berdell.R.Funke, ChristineL.case, Microbiology An Introduction. Pearson Publication

SEMESTER 2

ZF2VOT04 - VOCATIONAL COURSE 4

APPLIED MICROBIOLOGY 36 Hrs 2 Credits

Objectives

- 1. To describe the roles and importance of microbes in various environments
- 2. To develop an understanding of beneficial activities of microorganisms and the application of this understanding to benefit mankind.
- 3. To study the various interactions that exists among microbes and also between microbes and plants
- 4. To describe the roles and importance of microbes in various areas of agriculture

Module I

Microbiology of soil- soil profile, microbial flora, factor affecting flora, biogeochemical cyclecarbon and, nitrogen and its importance Microbial interrelationship of microorganisms, mutualism, synergism (protocooperation) commensalisms, Amensalism, Parasitism, and Predation.Interaction of microbes with plants- Microbes with plant roots- Rhizosphere concept, mycorrhizae, ecto, endo, VAM, actinomycorrhizae, phyllosphere concept.

Module II

8 Hrs

8 Hrs

Aquatic microbiology; Microbiology of water, Purification and disinfection of water.Waterborne pathogens, Sewage microorganism, BOD and COD, Sewage (waste water) treatment: primary treatment, secondary treatment (Oxidation Pond Trickling Filter, the Activated sludge, Anaerobic digesters), Tertiary treatment.

Bacteriological examination of drinking water- membrane filtration, MPN, plate count, (pour, spread), ONPG-MUG.

Module III

Role of microorganism in agriculture- Biofertilizer Bacterial- biological Nitrogen fixers-Symbiotic and free living nitrogen fixers, phosphate solubilizers, Algal biofertilizers- Azolla.

Module IV

Phytopathogens- Causative organism, mode of transmission, symptoms and control measures of Bacterial (soft rot, canker, wilt, crown gall), fungal (Blight, rot, wilt and mildew) and viral (TMV, Bunchy top disease of banana)plant diseases.

Control measures- Biopesticides (Bacterial, viral, fungal).

Module IV

Biogas, Role of microbes in biodegradation, bioremediation (natural organic compound, plastics, pesticides and petroleum pollutants).

Role of microbes in bioleaching, biomining.

References

- 1. B.P.Pandey Plant pathology, S. Chand & Company
- 2. G. Rangaswami, Agricultural Microbiology, Prentice Hall of India Pvt. Ltd.
- 3. R. P. Singh, Microbiology, KalyaniPublishers .
- Joanne M. Willey, Prescott, Harley & Klein's Microbiology, 7th Edition by McGraw & 4. Hill Publisher
- Joseph C. Daniel, Environmental aspects of Microbiology, Bright Sun publication 5.
- 6. Pelzar E. C. S. Chan and Noel R. Krieg Microbiology, Fifth edition, Michael.; Tata McGraw-Hill publishing company Ltd.
- 7. R.C.Dubey and D. K. Maheshwari, A Text book of microbiology, First Edition, S. Chand & company Ltd.

8 Hrs

4 Hrs

COURSE TITLE - MICROBIAL PHYSIOLOGY & GENETICS AND APPLIED MICROBIOLOGY PRACTICAL 2

36 hrs

2 Credit

Demo

- 1. Preparation of Solid and liquid media for microbial cultures.
 - a. Liquid media Peptone water, Nutrient broth
 - b. Solid media Nutrient agar
 - c. Semi Solid agar
- 2. Culture methods
 - a. Streak culture
 - b. Lawn culture
 - c. Stab culture

3.

nstration of selective and differential media

- a. Mac Conkey agar
- b. Blood agar
- 4. Isolation and enumeration of bacteria from rhizosphere & non- rhizosphere soil.
- 5. Standard plate count technique for the isolation and enumeration of microorganisms in water
- 6. Water quality analysis by MPN method
- 7. Isolation and identification of Rhizobium from root nodule
- 8. Basic idea about
 - a. Trickling filter
 - b. Membrane filter system
 - c. Anaerobic sludge digester

SEMESTER 3

ZF3VOT05 - VOCATIONAL COURSE 5

DAIRY MICROBIOLOGY

36 Hrs

2 Credits

Objectives

1. To provide a scientific background to dairy microbiology by reexamining the basic concepts of general food microbiology and the microbiology of raw milk.

2. The role of dairy starter cultures in manufacturing fermented dairy products, developing novel functional dairy products through the incorporation of probiotic strains

Module I

Introduction, composition and properties of milk, Nutritional importance of milk. Milk processing sequences. Source of microorganism in milk. Classification of microorganism in milk, biochemical types, temperature characteristics and pathogenicity.

Module II

Contamination and spoilage of milk and milk products.

Module III

Bacteriological examination of milk. Preservation of milk – Pasteurization (different methods and advantages), sterilization (ultra high temperature processed milk) dehydration.

Module IV

Type of milk and milk products: whole milk, low fat milk, toned milk, skim milk, vitamin D milk, low sodium milk, homogenized milk, concentrated milk, sweetened condensed milk evaporated milk, dry milk, low lactose milk. Lactic starter cultures. Probiotic role of lactic acid bacteria. Fermented milk products - curd, cream, butter, butter milk, lassie, Kefir and Kumiss, cheese, yoghurt.

Module IV

Milk borne disease, microbial diseases of dairy cattle and its control measures.

6 Hrs

6 Hrs

10 Hrs

8 Hrs

References

- 1. Blank F.C., Hand book of food nutrition
- Elmer H. Marth ,Marth H. Marth , Elmer H. Marth -Applied Dairy Microbiology, Second Edition.
- 3. James M. Jay, Modern food microbiology, 4th ed.
- Rameshwar Singh, S.K.Tomar&GunjanGoel, Applied Dairy and Food Microbiology Robinson -Dairy Microbiology Handbook, Third Edition: The Microbiology Of Milk And Milk Products.
- 5. Shakuntala N, Manay, M. Shadaksharaswamy, Food facts and Principles 2nd ed. New Age International publishers
- Shakuntala N, Manay, M. Shadaksharaswamy, Food facts and Principles 2nd ed. New Age International publishers
- 7. W.C. Frasier & Westhoff, Food Microbiology,
- 8. Winton and Winton, Milk and Milk Products.

PRACTICAL-3

COURSE TITLE – DAIRY MICROBIOLOGY

54 hrs

- 1. Qualitative analysis of milk by standard plate count method .
- 2. Isolation of coliforms from milk.
- 3. Detection of number of bacteria in milk by breed count.
- 4. Quality testing of milk by rezazurin test
- 5. Methylene blue reduction test for microbial contamination of milk.
- 6. Determination of phosphatase activity of milk
- 7. Detection of mastitis through milk test .
- 8. Isolation of Lactobacilli and Staphylococcus from curd
- 9. Fermented Dairy products

SEMESTER 3

ZF3VOT06 - VOCATIONAL COURSE 6 INDUSTRIAL MICROBIOLOGY

Objectives

- 1. Discuss the genetic manipulation of microorganism to construct strains that better meet the needs of an industrial or biotechnological process
- Discuss the preservation of microorganisms 2.
- 3. To study the major products or uses of industrial microbiology and biotechnology

Module I

History, development and scope of industrial microbiology, Screening of industrially important microorganisms- Primary and Secondary Screening. Strain Improvement - Mutation, Recombination, and protoplast Fusion.

Module II

Differentiate solid state and Basic Design & instrumentation of Fermenter, types of fermentation (solid state & submerged), types of fermenter - airlift fermenter, CSTR, tubular Fermenter, fluidized bed Fermenter.

Module III

Development of inoculum for industrial fermentation. Fermentation media, Carbon source, nitrogenous materials and antifoams.

Industrial sterilization- batch and continuous sterilization.

Module IV

Primary and secondary metabolites. Preservation of microbes - serial subculture, preservation by overlying culture with mineral oil, lypohilization, storage of microbes at a very low temperature or in liquid nitrogen. Methods for preservation of fungi.

Module V

Fermentation process: Surface, Submerged and solid state fermentation .Production of organic acids; acetic acid, citric acid, lactic acid. Production of amino acids: lysine and glutamic acid, production of enzymes: proteases and amylases. Production of antibiotics: Penicillin, Streptomycin, Production of vitamins- Vitamin B12 & riboflavin

6 Hrs

8 Hrs

8 Hrs

6 Hrs

8 Hrs

36 Hrs 2 Credits

References

- 1. A.H. Patel, Industrial microbiology, Mac Millan India Ltd.
- 2. K. Sukesh, M.M. Joe&P K Sivakumar- An Introduction to Industrial Microbiology,
- 3. L.E. Cesida, Industrial Microbiology, New Age International Publishers.
- Michael J. Waites, Neil L. Morgan, <u>John S. Rockey</u>Industrial Microbiology: An Introduction
- 5. Prescott, Harley & Klein Microbiology, 7th edition, Mac Graw Hill International edition.
- 6. Peter F. Stanbury, Principles of fermentation technology, Elsevier Publicatio

PRACTICAL 4

COURSE TITLE – INDUSTRIAL MICROBIOLOGY

54 hrs

0 Credits

- 1. Crowded plate technique for screening microbial antibiotics.
- 2. Solid state fermentation Mushroom production
- 3. Submerged fermentation- Study of alcoholic fermentation of fruit juice by yeast.
- 4. Immobilization of yeast cells

5. Preservation techniques:

- a. Serial sub culturing
- b. Over laying with mineral oil
- c. Lyophilization
- d. Liquid nitrogen storage.
- e. Methods for the storage of Fungi

SEMESTER 4 ZF4VOT07 - VOCATIONAL COURSE 7

BASIC FOOD MICROBIOLOGY

Objectives

- 1. To study interaction between micro-organisms and food .
- 2. To discusses the factors affecting the presence of micro-organisms in foods and their capacity to survive and grow.
- 3. To study recent developments in procedures used to assay and control the microbiological quality of food.

Module I

Food as a substrate for microorganisms. Factors affecting microbial growth in food, extrinsic -Temperature, Relative humidity and concentration of gases and intrinsic - hydrogen ion concentration (pH), water activity, oxidation reduction potential, nutrient content, inhibitory substances and biological structure.

Module II

Microorganisms - important in food microbiology, Bacterial - morphological, cultural and physiological characteristics important in food bacteriology, Important groups of bacteria associated with various foods. Molds and yeast associated with different foods.

Module III

Source of contamination of food:-Preharvest - from green plants and fruits, animals, soil, air, sewage, water and Post harvest:-processing equipment, transport, workers, during handling and processing General principles underlying spoilage of food; Chemical changes caused by microorganisms: Causes of spoilage, classification of food by ease of spoilage.

Module IV

Principles of food preservation, Asepsis, removal of microorganism, maintenance of anaerobic conditions, preservation by the use of high temperature, low temperature, drying, food additives and irradiation.

Module V

7 Hrs

7 Hrs

7 Hrs

8 Hrs

36 Hrs

2 Credits

Methods for the microbiological examination of foods: indicator organisms, direct examination, culture techniques, Enumeration methods - plate counts, most probable number counts: dye reduction test. Rapid methods for the detection of specific organism and toxins, immunological methods

References

- 1. George J. Banwart, Basic Food Microbiology
- 2. James M. Jay, Modern food microbiology, Van Nostand Reinhold Company
- 3. M.R. Adams, M.O. Moss, Food microbiology, New Age International (P) Ltd. Publishers
- 4. Prescott, Harleg, Klein, Microbiology, 7th ed. Mac Graw Hill International edition.
- 5. W.C. Frazier and Westhoff, .Food Microbiology.

COURSE TITLE – BASIC FOOD MICROBIOLOGY PRACTICAL 5

54 hrs

- I. Isolation and Enumeration of bacteria from spoiled food(vegetables, meat, fish).
- II. Biochemical tests used for identification of Bacteria.
 - a. IMVIC Test
 - b. Sugar Fermentation tests (GLSM)
 - c. Urease test
 - d. Catalase test
 - e. Oxidase test
 - f. Litmus milk reaction
 - g. TSI test
- III. Identification of bacteria from spoiled food samples.
- IV. Selective isolation and identification of
 - a) Staphylococcus
 - b) Salmonella
 - c) Vibrio

SEMESTER 4

ZF4VOT08 - VOCATIONAL COURSE 8

FOOD MICROBIOLOGY AND FERMENTED FOODS

36 Hrs

2 Credits

Objectives

1. To study interaction between micro-organisms and food – spoilage.

- 2. To study Foodborne illness
- 3. Production of different fermented Foods.

Module 1

Microbiology of cereal grains, flours and bread.Contamination, preservation and spoilage of vegetables and fruits, egg, meats and fish .

Module II

Fermented foods: oriental fermented foods-Tempeh, soysause, miso, idli, natto, Minchin and poi.Fermented vegetables - sauerkraut and Kimchi., Fermented meat and fish.

Microbial role in production of Bread, wine and malt beverages, single cell protein Mushroom production.

Module III

Food borne Diseases- Food poisoning, Bacterial food born infections (*Clostridium perfringens, Salmonella, Shigella, Campylobactor, Listeria, Vibrio, E.coli and Streptococcus faecalis)* and intoxication (*Staphylococcus aureus* and *Clostridium botulinm*). Mycotoxins-Aflatoxin, Patulin, Ochratoxin, Luteoskyrin and ATA.

Module IV

Food sanitation:-Principles of food sanitation, hygiene and safety. Sanitising equipments, cleaning agents and sanitising chemicals.

Module V

Hazard Analysis Critical Control Points (HACCP).

References

- 1. Banwart, Basic Food Microbiology,
- 2. Khetarpaul, Neelam, Food Microbiology.

• • • •

7 Hrs

10 Hrs

10 Hrs

4 Hrs

- 3. M.R. Adams, M.O. Moss, Food microbiology, New Age International (P) Ltd.
- 4. Prescott, Harleg, Klein, Microbiology, 7th ed. Mac Graw Hill International edition.
- 5. W.C. Frazier and Westhoff, Food Microbiology
- 6. W.M. Foster, Food Microbiology.

COURSE TITLE – BASIC FOOD MICROBIOLOGY FOOD MICROBIOLOGY AND FERMENTED FOODS PRACTICAL 6

54 hrs

- 1. Isolation and enumeration of fungi from spoiled food samples (vegetables, meat, fish)
- 2. Identification of fungi from spoiled food.
- 3. Lactophenol cotton blue staining of fungi.
- 4. Slide culture technique for identification of fungi.
- 5. Effect of incubation temperature on microbial growth.
- 6. Measurement of TDP
- 7. Measurement of TDT
- 8. Estimation of total and volatile acidity in alcoholic beverages.
- 9. Fermented food products

MODEL II VOCATIONAL COURSE

3. MEDICAL MICROBIOLOGY

SYLLABUS -

THEORY & PRACTICALS

VOCATIONAL SUBJECT: MEDICAL MICROBIOLOGY

VOCATIONAL COURSES

COURSE I ZMIV0T01	: Fundamentals of Microbiology		
COURSE II ZM1V0T02	: Basics of Microbial Physiology & Genetics		
Practical I	: Fundamentals of Microbiology and Basics of Microbial Physiology & Genetics		
COURSE III ZM2V0T03	: Parasitology		
COURSE IV ZM2V0T04	: Medical Virology		
Practical II	: Parasitology & Medical Virology		
COURSE V ZM3V0T05	: Medical Mycology		
COURSE VI ZM3V0T06	: Diagnostic Microbiology		
Practical III	: Medical Mycology		
Practical IV	: Diagnostic Microbiology		
COURSE VII ZM4V0T07	: Medical Bacteriology		
COURSE VII ZM4V0T08	I : Clinical Microbiology		
Practical V: N	Iedical Bacteriology		
Practical VI : Clinical Microbiology			

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SEMESTER I

COURSE I – FUNDAMENTALS OF MICROBIOLOGY

36 Hrs

Objectives

- 1. To introduce students to the fascinating world of microbes
- 2. Give an understanding of the scope of Microbiology
- 3. .To impart knowledge on major categories of microorganisms and understand their classification and appreciate their diversity.
- 4. To impart an awareness on how microbes can be controlled
- 5. To describe the parts and working principle of different microscopes and specimen preparation.

Module I

Definition of Microbiology, History: Early observation of microorganisms-Leewenhoek and Robert Hook; Spontaneous generation theory and refutation of spontaneous generation theory; Contributions of Robert Koch, Louis Pasteur, Lister, Edward Jenner, Beijernick and Winogradsky, discovery of antibiotics. Scope of microbiology

Module II

Microbial systematics -evolution of diverse microbial taxonomy as classification-taxonomic hierarchies- classical approach, numerical taxonomy, molecular based classification (16s rRNA-bacteria, Cytochrome oxidae I gene (COI), mitochondrial gene, ITS- in other protists in brief). Phylogenetic groups of bacteria- Archeae, Eukaryae-Bacterial taxonomy and nomenclature-Bergey's Manual

Module III

Control of microorganisms- Sterilization and disinfection .Control agents physical (moist and dry heat, by filtration, by irradiation) & chemical methods (alcohols, phenol, detergents, halogens, dyes);.Antibiotics- classification based on mode of action with eg. Instruments used microbial control – autoclave, hot air oven, laminar air flow

Module IV

10 Hrs

6 Hrs

10 Hrs

10 Hrs

Microscopy: - light microcopy, bright field, Dark field, phase contrast microscopy, fluorescence, Electron microscopy- SEM, TEM, STEM, Newer techniques in microscopy- confocal, scanning probe microscopy, Specimen preparation for light microscopy – wet mount preparation, hanging drop method, smear preparation. Bacterial staining – types positive and negative staining, simple and differential staining, Specimen preparation for electron microscopy.

References

Alcamo, Fundamentals of microbiology, 5th Edition,

Ananthanarayan & Panicker. Text book of Microbiology, 7th Edition, Orient Longman.

Jacquelyn G. Black, Microbiology: Principles and Explorations, 9th Edition, Willey Publication

Joanne M. Willey, Prescott, Harley & Klein's Microbiology, 7th Edition by McGraw & Hill Publisher

M.J. Pelczar, E.C.S Chan & N.R. Kreig, Microbiology, 5th Edition, Tata McGraw Hill Edition. Michael T. Madigan, Brock Biology of Microorganisms, 13th Edition, Amazon Publication

R.P. Singh, Microbiology, 1st Edition, Kalyani Publishers.

SEMESTER I

BASICS OF MICROBIAL PHYSIOLOGY AND GENETICS 36 Hrs

Objectives

- 1. To describe the differences in procaryotic and eukaryotic cell morphology and structure
- 2. To give a brief understanding of microbial physiology
- **3.** To describe some of the general methods used in the study and culture of microorganisms
- 4. To introduce microbial genetics

Module I

10 Hrs

Morphology and fine structure of bacteria, size, shape and arrangements. Flagella, pili, capsule – Indian ink staining, cell wall and its composition- Gram's staining, acid fast staining, Cytoplasmic membrane, protoplasts, spheroplasts, intracellular membrane systems, cytoplasm, vacuoles, nuclear material, bacterial spores- spore staining, cell inclusions.

Module II

8 Hrs

Microbial Nutrition- Classification based on nutritional requirements. Uptake of nutrientsactive and passive transport. Bacterial growth, Bacterial growth curve. Batch, Fed batch, continuous culture of microbes, Influence of environmental factors on microbial growth. Instruments involved in growth studies- Instruments- Incubator, colony counter

Measurement of microbial size- micrometry; Measurement of microbial numbers –viable and non viable counts **8 Hrs**.

Module III

Culture Media- definition, types of media- solid semi-solid and liquid media, Basal media, differential, selective, enrichment, enriched and transport media. Cultivation of Bacteria – Aerobic methods- streak plate, lawn culture, pour plate, spread plate, stab , liquid culture & Anaerobic culture methods-displacement O₂ with other gases, Pyrogallic acid and NaOH (chemical/biological method),anaerobic jar,Gaspak

Module IV

9 Hrs

Bacterial genetics -Introduction and history-Discovery of DNA as genetic material, DNA double helix-structure; Extrachromosomal genetic element- Plasmids-episome, compatible and incompatible, F factor, R plasmids (resistance transfer factor), Col plasmids. Reasons for variation- Bacterial recombination- conjugation (F+ x F- mating), transformation, transduction, transposons; Mutation (in brief); Transposable genetic element

References

Benson, Microbiological application laboratory manual in general microbiology, 8th Edition, McGraw Hill Publication.

Joanne M. Willey, Prescott, Harley & Klein's Microbiology, 7th Edition by McGraw & Hill Publisher

Jacquelyn G. Black, Microbiology: Principles and Explorations, 9th Edition, Willey Publication

M.A. Subramanian , Biophysics principles and techniques, MJP Publishers Microbiology, 5th Edition, M. J. Pelczar, E.C.S Chan & N.R. Kreig, Tata McGraw Hill Edition David Freifelder, Molecular biology, Narosa Publishing House

Michael T. Madigan, Brock Biology of Microorganisms, 13th Edition, Amazon Publication

SEMESTER I

PRACTICAL I- FUNDAMENTALS OF MICROBIOLOGY & BASICS OF MICROBIAL PHYSIOLOGY & GENETICS

36 Hrs

1.	Cleaning and sterilization of glassware, Autoclave, hot air oven, incubator and		
	laminar air flow	(3 Hrs)	
2.	Instrumentation and working principle of		

Laminar air flow Bench, Autoclave, Hot air oven, Colony counter (2 Hrs)

- 3. Preparation of Solid and liquid media for microbial cultures.
 - a. Liquid media (1) peptone water/Glucose broth (2) Nutrient broth (4 Hrs)
 - b. Solid media (1) Nutrient agar (2) Mac Conkey's agar (3) Blood agar
 - c. Semi Solid agar

4. Culture methods (a) streak culture (b) Lawn culture (c) Stab culture (d) Pour & spread plate culture (e) Liquid culture. (7 Hrs)
5. Isolation of pure colonies (a) streak plate method (b) pour plate method c)

- spread plate method(8 Hrs)6. Bacterial Staining Method Simple Staining, Gram's staining(4 Hrs)
- 7. Examination of microbes in Living condition (4 Hrs)(a) wet mount (b) Hanging drop method for demonstrating motility of bacteria
- 8. Measurement of bacterial growth by optical density method (4 Hrs)

SEMESTER II

COURSE III PARASITOLOGY

36 Hrs

Credits 2

Objectives

- 1 To understand the basic science of medial parasitology
- 2 To define the role of vectors in parasitology

- 3 To study the morphology, life cycle and pathogenesis of protozoan parasites
- 4 To study the morphology, life cycle and pathogenesis of protozoan parasites
- 5 To recognize the general outlines of parasite identification, control and prevention

General Introduction to parasitology- Type of parasites, types of host, sources & modes of parasitic infection. Classification of protozoan and helminthes. Collection and preservation of specimens for parasitological examination, transport of specimens, Examination of blood parasites thick and thin smears for malarial, filarial and other parasites

MODULE II

MODULE I

Common Vectors in Parasitology

Classification of arthropods of public health importance,

Role of Arthropods in the transmission of disease and their control measures (Mosquito-Aedes, Anopheles, Culex), Ticks, Flea)

MODULE III

Morphology, life cycle & pathogenesis (in brief), laboratory diagnosis of Protozoa: Entamoeba, Trichomonas, , Trypanosoma, Leishmania, Giardia, Plasmodium, Toxoplasma.

MODULE IV

Morphology, life cycle & pathogenesis (in brief), laboratory diagnosis each of the helminthes : a) Platyhelminthes: *Taenia, Echinococcus, Schistosoma, Fasciola*, (b) Nemathelminthes: *Ascaris, Ancylostoma, Trichinella, Enterobius, Trichurias, Wuchereria, Dracunculus*

References

C. H. Parija Text book of medical parasitology

K. P. Srivastava, A Text book of applied entomology, Kalyani Publishers

Panicker's Text book of medical parasitology, 7th edition C.K.Jayaram Panicker, Jaypee brother's medical publishers (p) Ltd.

Park's text book of preventive and social medicine, 22nd edition, Banasirdas Bhanot Publishers

8 Hrs

6 Hrs

10 Hrs

Subash C. Parija Text Book of Medical parasitology All India Publishers & **Distributors Publisher** Monica Cheesbrough, Medical Laboratory manual for Tropical Countries Microbiology Vol.I & II ELEBS.

SEMESTER II

COURSE IV MEDICAL VIROLOGY

36 Hrs

Credits2

Objectives

- 1. To develop a basic knowledge of virus morphology, classification and replication
- 2. To introduce the methods of cultivation of virus
- 3. To promote understanding of pathogenesis, diagnosis, prevention and control of human viruses

Module I

Viruses- Introduction. General characteristics of Viruses, Morphology-Size, structure & shape. Chemical properties. Bacteriophage-structure. Classification and nomenclature of viruses. Viroids & Prions. Steps in replication of Viruses. One-step growth curve. Lytic and lysogenic cycle.

Module II

Cultivation of Animal Viruses-animal inoculation, embryonated eggs, Tissue culture-organ culture, explants culture and cell culture a) primary cell culture, b) diploid cell culture d)continuous culture. Detection of growth of virus in cell culture-cytopathic effect (CPE), metabolic inhibition, hemadsorption, interference, transformation, immunoflurescence. Viral assay- electron microscopy and hemagglutination, assay of infectivity. Bacteriophageplaque assay-PFU. Antiviral agents-based on action with examples

Module III

Morphology, Pathogenicity, clinical features (in brief), lab diagnosis and treatment of each of these viruses:

Herpes virus (HSV, Varicella Zoster), Orthomyxovirus (influenza), Paramyxoviruses, (mumps,

measles) Rubella virus, Hepatitis virus (A, B, C), Rhabdo virus, AIDS virus, Polio virus,

Papiloma, Rota viruses, Corona virus-SARS

Module IV

5 Hrs

13 Hrs

8 Hrs

Arboviruses. Definition, Pathogenesis- 3 main syndromes: fever with and without rash and arthraglia, encephalitis, hemorrhagic fever (Brief mention of Chikun gunya, dengue, yellow fever, Zika, Ebola).

Oncogenic viruses-Definition and mechanism in brief, Examples of DNA oncogenic virus -Papova, Pox, Herpes and hepatitis B virus, RNA-retovirus (discussed in brief).

References

Ananthanarayan R. and C. K. J. Paniker. Text book of Microbiology, Sixth edition Gabriel Virella, B.I. Microbiology and Infectious diseases, 3rd ed. Waery Publications Philip A. Thomas, Clinical Microbiology, Orient Longman Pvt. Ltd.

Practical II PARASITOLOGY AND MEDICAL VIROLOGY

36 hrs

1.	Detection of parasite by lodine mount preparation	(5 Hrs)		
2.	Detection of parasite by Concentration technique (floatation-Z	inc Sulphate		
	method, sedimentation-formol ether)	(6 Hrs)		
3.	Examination of blood for parasites- Thin blood film	(8 Hrs)		
4.	I. Identification of Vectors of clinical importance- Mosquito(Anopheles, Culex),			
	Fleas, Ticks	(2 Hrs)		
5.	Identification of helminthic parasite- Ascaris, Ancylostoma, Taenia	(2 Hrs)		
6.	Introduction to cultivation techniques used in virology laboratory	(3 Hrs)		
7.	Method of cultivation of virus using embryonated eggs - Cho	orio Alantoic		
	Membrane (CAM)	(6 Hrs)		
8.	Demonstration of haemagglutination	(4 Hrs)		
	References			
	 N.Kannan, Laboratory manual in general microbiology, Paramount Publications 	Palani		
	2. Subash Chandra parija textbook of practical microbiology, Ahuja			

Publishing House, First edition

SEMESTER III

COURSE V MEDICAL MYCOLOGY

Objectives

- 1. To study the classification of fungus
- 2. To familiarize with routine mycological techniques, culture media and identification procedures
- To promote understanding of pathogenesis, diagnosis, prevention and control of medically important fungus

MODULE I

Classification of fungi; collection & transport of specimens, Examinations of fungus- mounting fluids and stains, Routine mycological techniques & Culture Media used for cultivation (Sabouraud's Dextrose Agar, Potato Dextrose Agar, Brain Heart Infusion broth & agar, Czapek Dox Agar, Corn meal agar, Bird Seed agar), Maintenance of fungus culture.

MODULE II

Superficial, Cutaneous& subcutaneous mycoses:

Superficial- distribution, etiological agents, clinical features, diagnosis, treatment of *Tinea versicolar, Tinea nigra, piedra (Black &White)*,

Cutaneous- Dermatophytoses (Causative agent, pathogensesity, lab diagnosis& treatment)

Subcutaneous: distribution, etiological agents, clinical features, diagnosis, treatment of Mycetoma, Sporotrichosis, Chromoblastomycosis,

MODULE III

Systemic mycoses- Causative agent, pathogenicity, clinical features, lab diagnosis & treatment of - Blastomycoses, Paracoccidioidomycoses, Coccidioidomycoses
MODULE IV 8 Hrs

10 Hrs

9 Hrs

9 Hrs

36 Hrs Credits2

Opportunistic mycoses- Causative agent, pathogensesity, clinical features, lab diagnosis & treatment of - Aspergillosis, Penicilliosis, Zygomycoses (Mucor, Rhizopus). Candidiasis

References

Jagadish Chander Text book of Medical mycology

Ananthanarayan R. and C.K.J. Paniker. Text book of Microbiology, Sixth edition

Medical Mycology by Rippon

P.Sivamani, Medical Mycology, 1st edition Siva Publications

Practical III MEDICAL MYCOLOGY

1.	Microbiology laboratory safety	(2 Hrs)
2.	Collection of samples for fungal infections.	(2 Hrs)
3.	Slide culture method for cultivation of fungus	(15 Hrs)
4.	Study of cultural characteristics of fungi on SDA- Aspergillus, Penici	llium ,
	Rhizopus ,Mucor	(6 Hrs)
5.	Study of morphology of fungi by lactophenol cotton blue - Aspergill	us,
	Penicillium , Rhizopus ,Mucor	(15 Hrs)
6.	Study of morphology of fungi by cellophane tape method	(4 Hrs)
7.	Germ tube test for the identification of Candida albicans	(4 Hrs)
8.	Demonstration of hair bait technique	(6 Hrs)
	References	

- 1. N.Kannan, Laboratory manual in general microbiology, Palani Paramount Publications
- 2. Subash Chandra parija textbook of practical microbiology, Ahuja Publishing House, First edition

Module I

SEMESTER III

Laboratory safety-Good lab hygiene, Personal barrier protection-gloves, outerwear, respiratory protection, eyes & face. Warning signage. Microbiological hazards-Biological Safety cabinet: Class I, II & III, Biosafety levels, Universal precautions, Decontamination, Hazardous waste-Infectious waste, Sharp waste and waste disposal.

Credits 2

Module II

36 Hrs

Microbiological methods- Morphology, Cultural characteristics, Biochemical characteristic-Indole, Methyl red, Voges Prauskauer, Citrate, Sugar fermentation, TSI, Oxidase, Catalase, Coagulase, DNAase, Urease, Gelatinase, Cellulase. H₂S production. Bacitracin, Optochin sensitivity, Antibiotic susceptibility assay- Kirby -Bauer method of Disk Diffusion, Tube dilution technique

Module III

Immunological/ Serological diagnosis- Definition of antigen, antibody, Ag-Ab reactionprecipitation and agglutination. Immunological detection methods- Immunodiffusion-Ouchterlony technique, Immunoelectrophoresis- Couter Immuno Electrophoresis (CIA), RadioImmuno Assay, ELISA, WIDAL, VDRL, ASO Coombs Test, Haemmagglutination inhibition, Quellung reaction, Mantoux test, Weil-Felix test

Module IV

Automated Methods for Diagnostic microbiology: Principles employed by common

automated systems for detection and identification of viable pathogens - Turbidity as an indicator of growth, Colorimetric and pattern recognition methods for microbial identification -

Flurophore- labelled/ oxidation-reduction substrate Vitek bacterial identification system,

metabolism as indicator of growth & substrate utilization-Biolog identification, API 20 E

strips, measurement of CO₂ as product of metabolic activity- BACTEC system, analysis of

Fatty acid profile – using GLC-MIDI Sherloch Microbial Identification Systems,

Bioluminescence assay, electrical impedance.

COURSE VI DIAGNOSTIC MICROBIOLOGY

7 Hrs

12 Hrs

5 Hrs

Module V

Molecular techniques: DNA probes, Blotting techniques-Western blotting, PCR, Emerging techniques in microbiology laboratory- MALDI-TOF Mass Spectrometry –description of this technique in brief.

Core Readings

Shanson D.C., Speller D.C. E. Microbiology in clinical practice,III edition, Butterworth & Heinemann Publication
Kenneth D M Clatchey Clinical Laboratory Medicine 2nd Edition
Carry-Ann D Burnham . Automation and Emerging Technology in Clinical Microbiology
Paul G. Engelkirk, Janet L. Duben-Engelkirk Diagnosis of infectious diseases: Essentials of Diagnostics.

Practical IV DIAGNOSTIC MICROBIOLOGY

Biochemical tests for the identification of microbes

1. Fermentation of carbohydrates (Glucose, lactose, Mannit	ol) (4 Hrs)
2. IMViC tes	(6 Hrs)
3. Urease test	(2 Hrs)
4. Catalase test	(4 Hrs)
5. Oxidase test	(4 Hrs)
6. Coagulase test	(4 Hrs)

7. Triple sugar iron agar test for the identification of E. coli, Klebsiella, Proteus and

Pseudomonas	(4 Hrs)
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Serological tests for identification of microbes

8.	VDRL	(6 Hrs))
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9. Widal	(12 Hrs)
10. Demonstration of immunodiffusion method	(4 Hrs)
11. Perform ASO	(4 Hrs)

References

- 1. N.Kannan, Laboratory manual in general microbiology, Palani Paramount Publications
- 2. Subash Chandra parija textbook of practical microbiology, Ahuja Publishing House, First edition

SEMESTER IV

COURSE VII MEDICAL BACTERIOLOGY

36 hrs Credits 2

Objectives

1 To develop a knowledge of medically important bacteria and their relevance of

infectious diseases;

2.To understand the principles of prevention and treatment of pathogenic

microorganism infection in humans.

Module I

Gram Positive & Negative cocci

Morphology, cultural& biochemical characteristics, antigenic properties, toxins/virulence factors(in brief), pathogenecity, lab diagnosis, prophylaxis & treatment - *Staphylococcus aureus*, *Streptococci* (*Str. pyogenes* and *Str. pneumonia*), Neisseriae (*N. meningitides* and *N. gonorrhoeae*)

Module II

Gram Positive Rods

Morphology, cultural& biochemical characteristics, antigenic properties, toxins/virulence factors(in brief), pathogenecity, lab diagnosis, prophylaxis & treatment *Bacillus anthracis, Corynebacterium diphtheriae, Clostridium tetani*

6 Hrs

Module III

Gram Negative Rods

Morphology, cultural& biochemical characteristics, antigenic properties, toxins/virulence factors(in brief), pathogenecity, lab diagnosis, prophylaxis & treatment

Escherichia coli, Klebsiella pneumoniae, Salmonella typhi, Shigella, Bordetella pertussis, Pseudomonas aeruginosa, Vibrio cholerae

Module IV

12 Hrs

Branching, Spiral, Pleomorphic & cell wall less bacteria-

Mycobacterium tuberculosis, Actinomycetes, Treponema pallidum, Haemophilus influenza type B, Rickettsiae (Typhus fever & Spotted fever group), Mycoplasma pneumoniae.

References

Ananthanarayan R. and C.K.J. Paniker. Text book of Microbiology, Sixth edition

Chakraborthy P., A textbook of Microbiology, 1st edition, , New Central Book Agency (P) Ltd.

Gabriel Virella, B.I. Microbiology and Infectious diseases, 3rd ed. Waery Publications

Practical V SEMESTER MEDICAL BACTERIOLOGY 54	hrs
Identification of bacteria based on morphology1. Differential staining- Spore staining for endospore	(10 Hrs)
2. Negative staining for capsulated organism	(8 Hrs)
3. Metachromatic granule staining for granules	(10 Hrs)
Identification of bacteria based on colony morphology	
4. Cultural characteristics on NA and Mac Conkey agar	(4 Hrs)
5. Demonstration of haemolytic property on Blood agar	(4 Hrs)
6. Antimicrobial susceptibility test	(8 Hrs)
7. Identification of Gram positive bacteria (Staphylococcus aureus)	(5 Hrs)

8. Identification of Gram negative bacteria (Escherichia coli, Klebsiella

pneumoniae)

References

- 1. N.Kannan, Laboratory manual in general microbiology, Palani Paramount Publications
- 2. Subash Chandra parija textbook of practical microbiology, Ahuja Publishing House, First edition

SEMESTER IV COURSE VIII CLINICAL MICROBIOLOGY

36 Hrs

(5 Hrs)

Credits 2

Objectives

1.To develop a basic understanding of laboratory safety, specimen collection and transport2 To describe the epidemiology, clinical manifestations, pathogenesis, and laboratory diagnosisof the following diseases caused by different microbial pathogens

Module 1

Microbiology laboratory safety, General concepts for specimen collection and transport of clinical specimens. Diagnostic methods in clinical microbiology and recent advances in diagnosis (in brief)

Module 2

Symptoms, Causative agents, clinical features, epidemiology and treatment of important Respiratory tract infections: Infections of the upper and lower respiratory tract-

Bacterial- Strep throat, Diphtheria, pneumonia (pneumococcal, *Klebsiella, Mycoplasma*), Pertussis, tuberculosis

Viral- Common cold- rhinovirus, Adenoviral pharyngitis, influenza, Respiratory Syncytial virus infections, Corona virus (SARS)

Fungal- Oral thrush

Module 3

Symptoms, Causative agents, clinical features, epidemiology and treatment of important gastrointestinal tract infections;

187

4 Hrs

10 Hrs

8 Hrs

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Bacterial-Cholera, Shigelllosis, *E. coli* gastroenteritis, Salmonellosis, Campylobacteriosis
Viral- Rota viral gastroenteritis, Hepatitis A, B, C
Protozoan- Giardiasis, Amoebiasis

Module 4

Symptoms, Causative agents, clinical features, epidemiology and treatment of important Urinary tract infection- Bacterial- bacterial cystitis, (E. coli, Klebsiella, Proteus, Staphylococcus, Pseudomonas), Fungal- Candida

Symptoms, Causative agents, clinical features, epidemiology, lab diagnosis and treatment of **Sexually transmitted diseases Non veneral**- bacterial vaginosis, vulvo vaginal Candidiasis; **Veneral**- Bacterial- Gonorrhoea, syphilis, Chlamydial genital system infection; **Viral**- AIDS, Papilloma virus, Genital Herpes Simplex

Module 5

4 Hrs

Important Infections of the nerve system- **Bacterial**- meningococcal meningitis, Hansen's disease (leprosy), Botulism

Viral- Viral meningitis (HSV, Mumps), Polio (infantile paralysis), rabies

References

Ananthanaryanan R. and C.K.J. Paniker Text book of Microbiology, 9th edition, University Press (India) Pvt. Ltd. Publisher

Elmer W. Koneman Color Atlas & Textbook of Diagnostic Microbiology 5th edition, Lippincott Publication

Eugene W. Nester, Microbiology a Human perspective, 4th edition McGraw Hill Publication.

Gabriel Virella, B.I. Microbiology and Infectious diseases, 3rd edition, Waverly Publications

Patricia M. Tille, Bailey & Scott's Diagnostic Microbiology 13th edition, Elsevier Publication Philip A. Thomas, Clinical Microbiology, Orient Longman Pvt. Ltd.

Richard V Goering, Mim's Medical Microbiology 5th edition, Elsevier Publication

Practical VI CLINICAL MICROBIOLOGY

54 Hrs

1.	Micro	biology of laboratory safety	(1Hr)
2.	Gener	al concept for specimen collection & handling	(3 Hrs)
3.	Semi o	quantitative method for analysis of urine	(8 Hrs)
4.	Exami	nation of clinical sample- urine	
	a.	Examination of specimen microscopically- wet mount,	Gram
		staining	(4 Hrs)
	b.	Culturing of specimen	(4 Hrs)
	c.	Biochemical identification of pathogen	(4 Hrs)
	d.	Antibiotic susceptibility test	(4 Hrs)
5.	Exami	nation of clinical sample- Sputum	
	a.	Examination of specimen microscopically- wet mount,	Gram
		staining	(4 Hrs)
	b.	Culturing of specimen	(4 Hrs)
	C.	Biochemical identification of pathogen	(4 Hrs)
	d.	Antibiotic susceptibility test	(4 Hrs)
6. I	dentifi	cation of normal flora of skin	(5 Hrs)
7.	Identifi	cation of normal flora of mouth	(5 Hrs)

References

- N.Kannan, Laboratory manual in general microbiology, Palani Paramount Publications
- Subash Chandra parija textbook of practical microbiology, Ahuja Publishing House, First edition

15. B.Sc ZOOLOGY PROGRAMME-MODEL – III

B.Sc. (BIOLOGICAL TECHNIQUES AND SPECIMEN PREPARATION) (B.Sc. BT & SP) INDUSTRIAL MICROBIOLOGY(DOUBLE CORE)

1. B.Sc. BIOLOGICAL TECHNIQUES AND SPECIMEN PREPARATION (B.Sc. BT & SP)

Total Credits 120 Total Instructional Hours 150

a. SCHEME OF DISTRIBUTION OF HOURS & EXAMINATION

i. THEORY:

Theory Examinations will be conducted by the University at the end of the respective semester in which the course is conducted. Duration 3 Hrs (Internal External ratio =1:4)

	Course	Course Title		Hrs per	Credits	Marks ratio	
No	Code	Course little	Course Category	week		Intl	Extl
1		English- (From Board of studies - English)	Common Course	5	4	1	4
2	ZB1CRT01	Introduction to Biological Sciences	Core Course 1 : Theory	2	2	1	4
3		Introduction to Biological Sciences	Core Course 1 : Practical	2	0	0	0
4	ZB1CRT02	Collection and Preservation of Biological Specimen 1 (Plants)	Core Course 2 : Theory	2	2	1	4
5		Collection and Preservation of Biological Specimen 1 (Plants)	Core Course 2 : Practical	2	0	0	0
6	ZB1CRT03	Collection and Preservation of Biological Specimen 2 (Animals)	Core Course 3 : Theory	2	2	1	4
7		Collection and Preservation of Biological Specimen 2 (Animals)	Core Course 3 : Practical	2	0	0	0
8		Biochemistry-1 (From Board of Studies" Biochemistry)	Complementary I : Theory	2	2	1	4
9		Practical	Complementary I :	2	0	0	0

	Practical				
Zoology -1 (From B of Studies- Zoology	oard Complementary II : Theory	2	2	1	4
Practical	Complementary II : Practical	2	0	0	0
Total			14		

	Course		Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1		English (From Board of studies – English	Common Course	5	4	1	4
3	ZB2CRT04	General Biological Techniques	Core Course 4 : Theory	2	2	1	4
4		General Biological Techniques	Core Course 4 : Practical	2	2	1	4
5	ZB2CRT05	Teaching Laboratory Techniques	Core Course 5 : Theory	2	2	1	4
6		Teaching Laboratory Techniques	Core Course 5 : Practical	2	2	1	4
7	ZB2CRT06	Food Microbiology & Biotechnology	Core Course 6 : Theory	2	2	1	4
8		Food Microbiology & Biotechnology	Core Course 6 : Practical	2	2	1	4
8		Biochemistry-2 (From Board of Studies - Biochemistry)	Complementary I : Theory	2	2	1	4
9		Practical	Complementary I : Practical	2	2	1	4
		Zoology -2 (From Board Of Studies- Zoology)	Complementary II : Theory	2	2	1	4
		Practical	Complementary II : Practical	2	2	1	4
		Total		25 hrs	24		

	Course		Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1	ZB3CRT07	Physiology With Clinical Correlation	Core Course 7 : Theory	3	3	1	4
2		Physiology With Clinical Correlation	Core Course 7 : Practical	2	0	0	0
3	ZB3CRT08	Clinical Chemistry an Clinical Microbiology	Core Course 8 : Theory	3	3	1	4
4		Clinical Chemistry and Clinical Microbiology	Core Course 8 : Practical	2	0	0	0
5	ZB3CRT09	Tissue Culture and Gene Manipulation	Core Course 9 : Theory	3	3	1	4
6		Tissue Culture and Gene Manipulation	Core Course 9 : Practical	2	0	0	0
7		Biochemistry-3(From Board Of Studies" Biochemistry)	Complementary I : Theory	3	3	1	4
8		Practical	Complementary I : Practical	2	0	0	0
9		Zoology -3 (From Board Of Studies Zoology	Complementary II : Theory	3	3	1	4
10		Practical	Complementary II : Practical	2	0	0	0
		Total		25 hrs	15		

	Course		Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Marks ratio Intl Extl 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4	Extl
1	ZB4CRT10	Radiology and Advanced Instrumentation Techniques	Core Course 10 : Theory	3	3	1	4
2		Radiology and Advanced Instrumentation Techniques	Core Course 10 : Practical	2	2	1	4
3	ZB4CRT11	Entrepreneurship Development and Marketing	Core Course 11 : Theory	3	3	1	4
4		Entrepreneurship Development and Marketing	Core Course 11 : Practical	2	2	1	4
5	ZY4CRT04	Research methodology,Biophysics & Biostatistics (Core Course IVof <i>Board of</i> <i>Studies - Zoology</i>)	Core Course 12 : Theory	3	3	1	4
6		Research methodology, Biophysics &Biostatistics	Core Course 12 : Practical	2	2	1	4
7		Biochemistry - 4 (From Board of Studies - Biochemistry)	Complementary I : Theory	3	3	1	4
8		Practical	Complementary I : Practical	2	2	1	4
9		Zoology - 4 (From Board of Studies – Zoology	Complementary II : Theory	3	3	1	4
10		Practical	Complementary II : Practical	2	2	1	4
		Total		25 hrs	25		

	Course Code	Course Title	Course	Hrs per		Marks ratio	
No			week	Credits	Intl	Extl	
1	ZY5CRT05	Environmental Biology & Human rights	Core Course 13 : Theory	3	3	1	4
2	ZY5CRPO5	Environmental Biology & Human rights	Core Course 13 : Practical	2	0	0	0
3	ZY5CRT06	Cell Biology & Genetics	Core Course 14 : Theory	3	3	1	4
4	ZY5CRPO6	Cell Biology & Genetics	Core Course 14 : Practical	2	0	0	0
5	ZY5CRT07	Evolution, Ethology & Zoogeography	Core Course 15 : Theory	3	3	1	4
6	ZY5CRPO7	Evolution, Ethology & Zoogeography	Core Course 15 : Practical	2	0	0	0
7	ZY5CRT08	Human Physiology, Biochemistry & Endocrinology	Core Course 16 : Theory	3	3	1	4
8	ZY5CRPO8	Human Physiology, Biochemistry & Endocrinology	Core Course 16 : Practical	2	0	0	0
	ZY5OPT01	1 – Vocational Zoology (Apiculture, Vermiculture, Ornamental fish culture)	Open Courses for other				
	ZY5OPT02	2 – Public health and Nutrition	streams (Select any one out of	4	3	1	4
9	ZY5OPT03	3 – Man, nature & Sustainable Development	three)	three)			
		Project work & Field Visit/Study Tour, Visit to research institutes, Group activity	Project work	1	0		
	Total			25 hrs	15		

	Course Code	Course Title	Course	Hrs per week	Credits	Marks ratio	
No			Category			Intl	Extl
1	ZY6CRT09	Developmental Biology	Core Course 17 : Theory	3	3	1	4
2	ZY6CRP09	Developmental Biology	Core Course 17 : Practical	2	2	1	4
3	ZY6CRT10	Microbiology & Immunology	Core Course 18 : Theory	3	3	1	4
4	ZY6CRP10	Microbiology & Immunology	Core Course 18 : Practical	2	2	1	4
5	ZY6CRT11	Biotechnology, Bioinformatics and Molecular Biology	Core Course 19 : Theory	3	3	1	4
6	ZY6CRP11	Biotechnology, Bioinformatics and Molecular Biology	Core Course 19 : Practical	4	2	1	4
7	ZY6CRT12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course 20 : Theory	3	3	1	4
8	ZY6CRP12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course 20 : Practical	2	2	1	4
	ZY6CBT01	Elective 1: Ecotourism & Sustainable Development					
	ZY6CBT02	Elective 2: Agricultural pest management	Choice Based Core Elective				
9	ZY6CBT03	Elective 3: Vector & Vector bourne Diseases	Courses (Select any one out of	3	2	1	4
	ZY6CBT04	Elective 4: Nutrition, Health & life style management	four)				
10	ZB6PRP01	Project work & Field Visit/Study Tour, Visit to research institutes , Group activity	Project		2		
11	ZB6OJP01	OJ (64 Hrs + 36 Hrs)	OJ		3		
	Total			25 hrs	27		

ii. SCHEME FOR PRACTICAL EXAMINATION FOR BSc (Biological Techniques and Specimen Preparation) BSc (BT&SP)

University Practical Examinations will be conducted at the end of even semesters (Semester II, IV and VI).

			E	Mark	s-ratio	
Semester	Code	Course	Exam duration	Internal (I)	External (E)	Credits
	ZB2CRP01	Practical 1 (Core 1 & 4)	3 hrs	1	4	2
	ZB2CRP02	Practical 2 (Core 2 & 5)	3 hrs	1	4	2
2	ZB2CRP03	Practical 3 (Core 3 & 6)	3 hrs	1	4	2
		Complimentary 1 Biochemistry	3 hrs	1	4	2
		Complimentary 2 Zoology	3hrs	1	4	2
	ZB4CRP04	Practical 4 (Core 7 & 10)	3 hrs	1	4	2
	ZB4CRP05	Practical 5 (Core 8 & 11)	3 hrs	1	4	2
4	ZB4CRP06	Practical 6 (Core 9 & 12)	3 hrs	1	4	2
		Complimentary 1 Biochemistry	3 hrs	1	4	2
		Complimentary 2 Zoology	3hrs	1	4	2
	ZY6CRP03	Practical 7 (Core 13 & 17)	3 hrs	1	4	2
	ZY6CRP04	Practical 8 (Core 14 & 18)	3 hrs	1	4	2
6	ZY6CRP05	Practical 9 (Core 15 & 19)	3 hrs	1	4	2
	ZY6CRP06	Practical 10 (Core 16 & 20)	3 hrs	1	4	2

b. SYLLABUS THEORY & PRACTICALS

(BIOLOGICAL TECHNIQUES AND SPECIMEN PREPARATION) (B.Sc. BT & SP)

SEMESTER I

ZB1CRT01 - CORE COURSE 1 INTRODUCTION TO BIOLOGICAL SCIENCES

36 hrs

Credits 2

OBJECTIVES

- 1. To develop proper scientific mind, culture and work habits
- 2. To emphasize the central role that biological sciences plays in the life of all organisms
- 3. To introduce the student to some of the present and future applications of bio-sciences

Module 1

What is biology: Salient features of life; Importance of biology on the frontiers of science and technology, History of Biology, Biology in ancient times, Landmarks in the progress of Biology, Branches of Biology

Module 2

Introduction to the world of living organisms: Outline classification of living organism, levels of biological organization, Broad overview of life on earth; history and evolution of life on earth, Theories of evolution, origin and progression of life on earth.

Module 3

Chemicals of life: Elements found in living organisms, acids and bases, An overview of Carbohydrates, proteins, lipids,nucleic acids, vitamins and minerals including functions, physical, chemical properties, basic units, types, Physical chemistry and significance of water, interactions between molecules.

Module 4

Homeostasis:Definition, Concept and importance in biological system. Control systems in biology, general idea of biological negative feedback mechanisms and temperature regulation.

Module 5

Nature and Scope of biology: Branches of biology- Botany, Zoology, Cell biology, Molecular biology, Developmental biology, Marine biology, Ecology, Physiology, Anatomy, Morphology, Genetics, Biochemistry, Microbiology, Biotechnology

10 Hrs

6 Hrs

6 Hrs

8 Hrs

Rferences

- 1. Debbies Holmes, Peter Moody and Diana Dine (2006) Research methods for the Biosciences. International student Edition: Oxford University Press. P. 288-299.
- Bowler Peter J. And Iwan Rhys Morus (2005) Making Modern Science: A Historical Survey. University of Chicago Press, Chicago, IL:
- N. Campbell and J. Reece (2005) Principles of Biology: Interactive textbook from Nature Education Biology: 7th edition, Pearson, Benjamin, Cummings
- Chakrabarti B K, Ghosh H N & Sahana S N (1984): Human Physiology, The New Book Stall, Calcutta, India
- Ernst Mayr (1982) The Growth of Biological Thought: Diversity, Evolution and Inheritance.Published by Harvard University Press.
- Ernst Myer. (1997). This is Biology: The Science of the living World.University Press, Hyderabad, India
- Kuhn, Thomas. (1996) The Structure of Scientific Revolutions3rd ed.: University of Chicago Press, Chicago, IL
- Knudsen, J.W (1966)*Biological Techniques* Harper InternationalEdition by Harper & Row
- 9. Marie, M (2005) Animal Bioethics: Principles and Teaching Methods Wageningen Academic Publishers
- D.E. Metzler (2003) Biochemistry: The chemical reactions of living cells: Volumes I & II, 2nd edition, Academic Press
- Roger Eckert; D Randall; George Augustine (1988) Animal Physiology, Mechanism and Adaptations, W.H Freeman, NewYork
- 12. Taylor et al., (2008) Biological Science Cambridge University Press,
- Thomas, A.P (2009) Biology Perspectives and Methods. Green Leaf Publishers, Kottayam.
- K. Vijayakumaran Nair & Biju Dharmapalan (2010) Methodology and Perspectives of Science; Publisher: Trivandrum: Academica.

SEMESTER I

CORE COURSE 1 INTRODUCTION TO BIOLOGICAL SCIENCES (PRACTICAL)

36 Hrs

0 Credit

	0 Ci cuit
1. Simple identification of any 5 vertebrates and 5 inverte	ebrates upto species and
differences in classification of vertebrates and invertebrates	6 Hrs
2. Identification of a) plant c	
3. ell (onion cell) b) animal cell (cheek cell) (5 hrs)	
4. Study on food chain and food web	5 Hrs
5. Physiological Measurements: Blood Pressure (normal & unde	er stress) and Temperature
	8 Hrs
6. pH measurements	6 Hrs
a) Measurement of pH of different solutions, like aerated drinks,	fruit juices, shampoos and
soaps (use dilute solutions of soaps and shampoos to prevent	damage to the glass
electrode) using pH-meter	
b) Determination of pH of water	

7. Titration experiments: estimation of acids and bases

SEMESTER I

ZB1CRT02 - CORE COURSE 2 COLLECTION AND PRESERVATION OF BIOLOGICAL SPECIMEN 1 (PLANTS)

36 Hrs 2 Credits

Objectives

- 1. To introduce the student to some of the collection and preservation of plant specimens
- 2. To develop critical thinking skill and research aptitude among students, by introducing the frontier areas of the biological science

Module 1

Broad classification of plants: where and how to collect plants, Overview of economically important plants, preparation and storage of herbarium sheets; preparation of dry specimens for

12 Hrs

display boxes; preparation of museum specimens. Modelling materials: characteristics of teaching models, proportions, durability, attractiveness, innovativeness.

Module 2

Anatomy of Plants: Special features of anatomical sections of monocot and dicot stems and roots; Double staining methods; Special staining methods; Preparation and storage of permanent slides.

Module 3

Cell division stages: Stages of Mitosis and Meiosis in Plants; Sources of materials; Preparation of permanent slides showing stages of division; Use of chemicals to arrest division; Special stains and their preparation.

Module 4

Distribution of plants: methods of survey, different approaches of sampling, determination of frequency dominance

References

- 1. Aggarwal S.K (2009)Foundation Course in Biology Ane's Students Edition
- Cappucchino J.G., and Sherman, N. Microbiology A Laboratory Manual3rd Ed. The Benjamin/Cummings Publishing Co.
- 3. Dubey, R.C. and Maheshwari, D.K. (2002) Practical Microbiology S.Chand& Company Ltd.
- Eldon D. Enger, Frederick C. Ross and David Bailey (2008)(Eleventh Edition)Concepts in Biology. Tata – McGraw Hill, New Delhi
- 5. Talaro, K.P., and Talaro, A. 2002. Foundations in Microbiology 4th Ed. McGraw Hill.
- 6. Taylor, Green, Stout (2008) Biological Science, Cambridge University press.

SEMESTER 1

CORE COURSE 2 COLLECTION AND PRESERVATION OF BIOLOGICAL SPECIMEN 1(PLANTS) (PRACTICALS)

36 Hrs 0 Credit

1.	Preparation of herbarium sheets	(5 Hrs)
2.	Identification of mitotic stages: onion root tip	(5 Hrs)

9 Hrs

9 Hrs

3.	Preparation of museum specimens	(5 Hrs)
4.	Preparation of display boxes of dry plant and plant product mounts	(5 Hrs)
5.	Preparation of whole mounts	(4 Hrs)
6.	Collection and preservation of materials for anatomical and cytological studies	(2 Hrs)
7.	Preparation of teaching models [Plaster of Paris, Epoxy Resin, Clay]	(10 Hrs)

ZB1CRT03 - CORE COURSE 3 **COLLECTION AND PRESERVATION OF BIOLOGICAL SPECIMEN 2 (ANIMALS)**

36 Hrs 2 Credits

Objectives

- 1. To introduce the student to some of the collection and preservation of animal specimens
- 2. To develop critical thinking skill and research aptitude among students, by introducing the frontier areas of the biological science

Module 1

Collection and preservation of invertebrates: (Protista, porifera, coelenterata, ctenophora, Platyhelminthes, Nematoda, Annelida, Arthropoda, Mollusca, Echinodermata, hemichordata

Module 2

Collection and preservation of vertebrates: Pisces, aves, amphibians, reptiles and mammals

Module 3

Life cycles: study on life cycle of a selected animal from each phylum (Protista, porifera, coelenterata, ctenophora, Platyhelminthes, Nematoda, Annelida, Arthropoda, Mollusca, Echinodermata, hemichordate, Pisces, aves, amphibians, reptiles and mammals)

Module 4

Preparation of specimens: Preparation of museum specimens, articulated skeletons, Dermestid technique, Alizarin preparation and resin-embedded specimens, Preparation of sections involving microtome and cryostat, Taxidermy.

6 Hrs

8 Hrs

6 Hrs

Module 5

Demographic Techniques and Population Patterns of animals: Techniques Used to Quantify Population Density, Patterns of Spacing, Fragmented Habitats, Spatial Arrangement of Habitats and Organisms.

References

- 1. Blamire, J. (1994)Exploring Life- The Principles of Biology, Wm. C.Brown Publishers
- Campbell, N.A., and Reece, J.B (2005)Biology. 7th (International) Ed. Pearson-Benjamin-Cummings
- Green, N.P.O., Stout, G. W. & Taylor, D.J (1990) Biological Science2ndEd. Cambridge Low Price Edition, Cambridge University Press.
- Knudsen, J.W (1966) Biological TechniquesHarper InternationalEdition by Harper & Row
- Hickman, C.P., Roberts, L.S. and Larson, A (2003) Animal Diversity3rdEd. Mc Graw Hill
- 6. Miller, S.A., and Harley J.P. (2005)Zoology. 6th Ed. Mc Graw Hill

SEMESTER I

CORE COURSE 3 COLLECTION AND PRESERVATION OF BIOLOGICAL SPECIMEN 2 (ANIMALS) (PRACTICALS)

36 Hrs

0 Credit

1. Whole mount preparation of small animals and parts of animals	(9 Hrs)
2. Alizarin preparation of small invertebrates	(4 Hrs)
3. Preparation of articulated skeletons	(6 Hrs)
4. Preparation of resin embedded specimens	(4 Hrs)
5. Demonstration of Taxidermy	(5 Hrs)
6. Preparation of specimens by each student from a given phylum	(4 Hrs)
7. Frequency distribution of animals in a specific area of campus	(4 Hrs)

ZB2CRT04 - CORE COURSE 4 **GENERAL BIOLOGICAL TECHNIQUES**

36 Hrs 2 Credits

Objectives

Module 1

- 1. To impart a knowledge and understanding of biological experimental techniques, includingpractical laboratory skills To familiarize with the basic tools and techniques of scientific study with emphasis on 2.
- biological sciences

Microscopy: Magnification and Resolution, bright field, dark field , phase contrast, stereoscopic, fluorescence, polarization microscopy, electron microscopy-SEM, TEM.

Ocular and stage micrometers; Hemocytometer; Camera lucida

Module 2

Seperation techniques: Centrifuge, chromatography, electrophoresis Module 3

Analytical techniques: Colorimeter, pH meter, Spectrophotometer, x-ray crystallography.

Module 4

Micrbiological Techniques: Identification of common microorganisms, culture techniques; Types of solid and liquid culture media for bacteria, fungi, algae and protozoa (at least 2 for each); Sterilization methods, Cell counting and methods for measuring microbial growth; Storage and maintenance of Stock cultures.

References

- 1. Bhaskaran, K.K (1986) Microtechnique and Histochemistry. Evershine Press, Vellangalloor
- 2. Cappucchino, J.G., and Sherman N. Microbiology A Laboratory Manual3rd Ed. The Benjamin/Cummings Publishing Co
- 3. Dubey, R.C. and Maheshwari, D.K (2002) Practical Microbiology S.Chand& Company Ltd.
- 4. Junqueira, L.C., and Carneiro, J (2005)Basic Histology11th Ed. Mc GrawHill
- 5. Talaro, K.P., and Talaro, A (2002)Foundations in Microbiology4th Ed. McGraw Hill.

12 Hrs

6 Hrs

6 Hrs

ZB2CRP01 - CORE COURSE 4 GENERAL BIOLOGICAL TECHNIQUES (PRACTICALS)

	36 Hrs Credit 2
1. Light microscope: its parts and their description	(3 Hrs)
2. Use of ocular and stage micrometers for measurement of width of hair etc.	(5 Hrs)
3. Camera lucida	(4 Hrs)
4. Chromatography	(10 Hrs)
5. Instrumentation-pH meter, Colorimeter, centrifuge, electrophoresis	(8 Hrs)
6. Counting cells in hemocytometer; Growth Curve	(6 Hrs)

SEMESTER 2

ZB2CRT05 - CORE COURSE 5 TEACHING LABORATORY TECHNIQUES

36 Hrs

Credits 2

26 11.

Objectives

- 1. To impart a knowledge and understanding of biological experimental techniques, including practical laboratory skills.
- 2. To learn about laboratory techniques, water, soil and air analyses.

Module 1

Organization of a teaching laboratory: equipment, reagents, glass wares, specimens, purchase and maintenance of stock register

Module 2

Distillation of water: Types of distillation stills [metal, solar, glass still],Cleaning agents for glassware, Methods of sterilization and storage of glassware.

Module 3

10 Hrs

8 Hrs

Water pollution:Abiotic and biotic pollutants of water and their indicators; Assay techniques (any 5).

Module 4

Air pollution:Introduction to air pollution, various factors contributing to air pollution, common air pollutants and sources of air pollutants, technology for air pollution control, Assay techniques(any 5).

Module 5

Soil pollution: Introduction to soil pollution, types and sources, main causes and effects, control measures of soil pollution. Assay techniques(any 5).

References

- 1. Arms, K. (1990)Environmental Science, Saunders College Publishing
- 2. Christopher.F Forster, D.A.John Wase, (1987) Environmental Biotechnology, Ellis Harwood.
- 3. Joseph C. Daniel(1999), Environmental aspects of Microbiology, Bright Sun publication
- 4. Khopkar, S.M (1993) Environmental Pollution Analysis, New Age International (P) Limited Publishers
- 5. Robert Jennings Heinsohn, Robert Lynn Kabe(1999) Sources and Control of Air Pollution, Prentice Hall
- 6. Sharma, P.D (1994)Ecology and Environment6th ed. Rastogi Publications
- 7. R. P. Singh (2012) Microbiology, Kalyani Publishers

SEMESTER 2

ZB2CRP02 - CORE COURSE 5 GENERAL LABORATORY TECHNIQUES (PRACTICALS)

36 Hrs

2 Credits

1. Components of distillation stills and ion exchanger	(2 Hrs)
2. Cleaning of dirty glass wares using various cleaning agents.	(4 Hrs)
3. Sterilization of glass wares [using hot air oven and autoclave	(4 Hrs)
4. Analysis of water pollutants:	
Determination of hardness of water	(3 Hrs)
Determination of residual chlorine of water	(3 Hrs)

6 Hrs

Estimation of COD	(3 Hrs)
Estimation of BOD	(3 Hrs)
Bacteriological examination of water by MPN method	(6 Hrs)
5. Analysis of soil pollutants: Determination of total bacterial populations from so	il (4 Hrs)
6. Analysis of air pollutants: Enumeration of microorganisms from air	(4 Hrs)

ZB2CRT06 - CORE COURSE 6 FOOD MICROBIOLOGY & BIOTECHNOLOGY

36 Hrs

2 Credits

OBJECTIVES

- 1. To make aware of different useful microorganisms, their role in food processing and preservation.
- 2. To understand the factors and predict microorganisms, which can cause foodspoilage.
- 3. To understand the causes of foodborne diseases.
- 4. To give a brief outline of food production through biotechnology

Module 1

Roles of microbes in food production: Bread, wine, curd, yoghurt, cheese, , food and fodder yeast. Mushroom production.

Module 2

Preservation and spoilage of food: Principles underlying spoilage of food, causes of spoilage, classification of food by ease of spoilage.

Control of microbes in food; Sterilization- Preservation by the use of high temperature, low temperature, drying. Aseptic packaging, Canning

Module 3

Food borne diseases: Important groups of bacteria associated with various foods. Food poisoning , food borne infections and intoxication. Brief description on *Clostridium botulinum*, *Vibrio, Salmonella, Hepatitis A, E. coli* 0157:H7,*Listeria*.

Module 4

Food additives and adulteration: Uses of food additives: Non preservative, preservative. Food adulterants & Prevention of Food Adulteration: common adulterants for foods like milk and milk products, atta, edible oils, cereals, condiments and curry powder.

208

8 Hrs

8 Hrs

6 Hrs

1.6

Module 5

6 Hrs

Biotechnology in food production:Transgenic plants-Flavr- savr tomatoes; Nutritionally enriched foods:Golden rice, "heart healthy" canola oil.GM foods- advantages and disadvantages. Single cell protein, algae as food. Biotechnological potential of microalgae, food, feed and fuel production of pharmaceutically valuable microalgae.

References

- M.R. Adams, M.O. Moss, Food microbiology, New Age International (P) Ltd. Publishers.
- Bala Subramanian D., C.F & Bryle & K. Dharmarajan J. Green Kunthala Jayaraman (2007)Concept in Biotechnology. University Press
- Colin Ratledge & Bijorn Kristiansen (2008)Basic Biotechnology 3 rd ed. Cambridge University
- 4. W.C. Frazier and Westhoff, . Food Microbiology.
- Lindsay, (1988) Willis Biotechnology, Challenges for the flavour and food industries", Elsevier Applied Science.
- 6. George J.B., (1987) Basic Food Microbiology, CBS Publishers & Distributors,
- Gavin Brooks (1998) Biotechnology in Healthcare: An Introduction to Biopharmaceuticals Pharmaceutical Press.
- Janarthanan S & Vincent S (2007) Practical Biotechnology, Method of Protocols. University Pres.
- 9. John E. Smith(2005) Biotechnology Cambridge Low priced ed. (ThirdEd)
- 10. Prescott, Harleg, Klein, Microbiology, 7th ed. Mac Graw Hill International edition.
- Roger A., Gorden B., and John T., (1989). Food Biotechnology Cambridge University Press
- 12. Ronald H. Schmidt and Gary E. Rodrick. (2002). Food Safety Handbook. Wiley; 1st edition. Part 1: Definition of food safety and characterization of food hazards.
- 13. Ronald H. Schmidt and Gary E. Rodrick (2002) Food Safety Handbook. Wiley; 1st edition. Part 2: Prevalence of foodborne pathogens
- 14. Singh B.D. (2002) Biotechnology Kalyan Publishers Nw Delhi.
- 15. N Shakuntala Manay, M. Shadakshara Swamy Food-Facts and Principles II Ed. New Age International Pub.

16. Som Nath Mahindru (2009). Food additives: characteristics, detection and estimationNew Delhi APH Publ. Corp.

SEMESTER 2

ZB2CRP03 - CORE COURSE 6 FOOD MICROBIOLOGY & BIOTECHNOLOGY (PRACTICALS)

36 Hrs

2 Credits

1.	Preparation of fermented foods: curd, yogurt.	(6 Hrs)
2.	Isolation and Enumeration of bacteria from spoiled food (vegetables,	meat and fish)
		(8 Hrs)
3.	Identification of bacteria from spoiled food samples.	(6 Hrs)
4.	Qualitative analysis of milk by standard plate count method.	(6 Hrs)
5.	Methylene blue reduction test for microbial contamination of milk.	(5 Hrs)
6.	Food adulteration detection by physical and chemical tests.	(5 Hrs)
	Physical tests; Tea leaves, Black pepper, Cumin seeds, Cloves, Arhar	dal (Toor dal)
	Chemical tests; Coffee powder, Turmeric powder, Coconut oil, Ghee,	Jaggery, Sugar

SEMESTER 3

ZB3CRT07 - CORE COURSE 7 PHYSIOLOGY WITH CLINICAL CORRELATION

54 Hrs 3 Credits

Objectives

- 1. To inspire the students in learning the frontier areas of biological sciences
- 2. To appreciate the correlation between structure and function of organisms
- **3**. To make them aware of the different body systems and the need for maintaining good health through appropriate life style.

Module 1

Basics of human physiology: levels of structural organisation- chemical level, cellular level (in brief), tissue level- epithelial tissue, connective tissue, membranes.

Module 2

Muscular system: types of muscular tissue, ultra structure of myofibril, sliding filament mechanism, neuromuscular junction, muscle metabolism, common clinical abnormalities (any 8).

Respiratory system: functional organization, common clinical abnormalities

Module 3

Nervous system: structure of neuron, electrical signals in neuron, signal transmission, anatomy of brain and anatomy of spinal cord, spinal cord physiology, common clinical abnormalities (any 8).

Module 4

Endocrine system: anatomy of endocrine glands, hormone activity, mechanism of hormone action, functions and common clinical abnormalities (any 8).

Module 5

Cardiovascular system: anatomy of Heart and circulation of blood, cardiac conducting system, ECG, cardiac cycle, cardiac output, composition of blood, blood clotting and blood groups ,blood pressure and Common clinical abnormalities (any 8)

References

- 1. Fox, S.I.(2006) Human Physiology9th ed. McGraw Hill International Edition
- 2. Guyton and Hall (2006) Text book of Medical Physiology
- 3. Seeley, R.R., Stephens, T.D., and Tate, P(2006) Anatomy and Physiology7th ed. McGraw Hill International Edition
- 4. Thibodeau, G.A., and Patton, K.T(2007)Anthon's Textbook of Anatomy and Physiology. 18th ed. Mosby
- Tortora, G.J., and Derrickson, B (2006) Principles of Anatomy and Physiology11th ed. John Wiley & Sons, Inc.

2 Hrs

6 Hrs

10 Hrs

8 Hrs

CORE COURSE7 PHYSIOLOGY WITH CLINICAL CORRELATION (PRACTICALS)

36 Hrs 0 Credit

1. Determination of O ₂ uptake by cockroach [Respirometer]	(3 Hrs)
2. Effect of adrenalin and noradrenalin on the heart beat of frog (demo)	(2 Hrs)
 Preparation of human blood smear and identification of leucocytes Determination of differential WBC count 	(6 Hrs) (6 Hrs)
5. Estimation of haemoglobin	(4 Hrs)
6. Demonstration of hemin crystals	(3 Hr)
7. ESR	(4 Hrs)
8. Blood grouping (ABO, Rh).	(2 Hrs)
9. Bleeding time and Clotting time	(6 Hrs)

SEMESTER 3

ZB3CRT08 - CORE COURSE 8 CLINICAL CHEMISTRY AND CLINICAL MICROBIOLOGY

54 Hrs 3 Credits

Objectives

- 1. To inspire the students in learning the frontier areas of biological sciences
- 2. To expose the students to fundamentals in clinical chemistry and to make them appreciate the relevance of the subject in biological studies.
- 3. To make them aware of the pathogens, health related problems, their origin and treatment.

Module 1

Lifestyle diseases: AIDS, Diabetes Mellitus, Obesity, Cancer, Cardiovascular diseases, kidney disorders, liver disorders.

Module 2

12 Hrs

Functions of various organs and their clinical assessment (Brief treatment only but emphasizing the biochemical aspect): e.g., liver, kidney, heart, pancreas, endocrine glands, lung, brain. Biochemical changes in the organs under pathological conditions.

Module 3

Routine biochemical tests of blood sugar, cholesterol and NPN.

Module 4

Microorganisms of medical importance- Symptoms, causative agents, clinical features, laboratory diagnosis and treatment of important

Bacterial diseases- Diphtheria, Pneumonia, Cholera, Tuberculosis, Salmonellosis, Typhoid

Viral diseases - Common cold, Respiratory Syncytial virus infections, Corona virus (SARS), AIDS

Fungal diseases - Oral thrush, Aspergillosis

Module 5

Parasitic diseases- Symptoms, causative agents, clinical features, laboratory diagnosis and treatment of Malaria, Filariasis, Giardiasis, Amoebiasis,

References

- Ananthanaryanan R. and C.K.J. Paniker (2009) Text book of Microbiology, 9th edition, University Press (India) Pvt. Ltd. Publisher
- Elmer W. Koneman (2006) Color Atlas & Textbook of Diagnostic Microbiology 5th edition, Lippincott Publication
- Cheesbrough, M. (1998) District Laboratory Practice in Tropical CountriesPart 1. Cambridge Low Price Edition. Cambridge University Press
- Cheesbrough, M. (1998) District Laboratory Practice in Tropical CountriesPart 2. Cambridge Low Price Edition. Cambridge University Press
- 5. Mukherjee, K.L. (ed,) (1988) Medical Laboratory Technology Vol. 1. TataMcGraw Hill
- 6. Mukherjee, K.L. (ed,)(1988) Medical Laboratory Technology Vol. 2. TataMcGraw Hill
- 7. Mukherjee, K.L. (ed,)(1988) Medical Laboratory Technology Vol. 3. TataMcGraw Hill
- 8. Philip A. Thomas (2007) Clinical Microbiology, Orient Longman Pvt. Ltd.
- 9. Talaro, K.P., and Talaro, A. (2002). Foundations in Microbiology4th ed. McGraw Hill

14 Hrs

4 Hrs

SEMESTER III

CORE COURSE 8 CLINICAL CHEMISTRY AND CLINICAL MICROBIOLOGY (PRACTICALS)

36 Hrs 0 Credit

1.	Estimation of: blood glucose, total protein in serum, serum albumin,	bloodurea,
	creatinine in blood, serum bilirubin, serumcholesterol, serum alkaline phospha	atase, serum
	acid phosphatase.	(14 Hrs)
2.	Media preparation, Inoculation, and maintenance of bacteria.	(7 Hrs)
3.	Gram staining	(2 Hrs)
4.	Acid fast staining	(2 Hrs)
5.	Widal Test	(2 Hrs)
6.	Identification of microorganisms (bacteria, fungi) of clinical significance	(9 Hrs)

SEMESTER 3

ZB3CRT09 - CORE COURSE 9 TISSUE CULTURE & GENE MANIPULATION 54 Hrs

3 Credits

OBJECTIVES

- 1. To emphasize the central role that genetics plays in the life of all organisms
- 2. To learn about the tissue culture techniques
- 3. To introduce the student to some of the present and future applications of bio-sciences
- 4. To develop critical thinking skill and research aptitude among students, by introducing the frontier areas of the biological science.

Module 1

12 Hrs

Manipulating DNA: in microbes, plants and animals – overviews

Restriction endonucleases, ligases, cloning vectors [plasmids & phage DNA]

Isolation of genomic DNA, Mechanism of gene transfer and methods, Identification and selection of recombinants.

Recombinant DNA technologyand its applications.

Module 2	12 Hrs	
Gene Cloning:Introduction and overview of Gene cloning; Techniques, principles and applications		
Module 3	8 Hrs	
Plant tissue culture: mediaand composition. Characteristic of plant cells in culture, callus,meristem,anther,embryo,ovule,ovary and endosperm culture		
Module 4	12 Hrs	
Germ plasm: Storage, somatic hybridization		

Module 5 10 Hrs

Animal cell culture: growth media and maintenance of culture. Characteristics of animal cells in culture, hybridoma technology.

References

- 1. Brown, T.A (2007)Genomes 3. GS Garland Science
- 2. Glick B.R., Pasternak, J.J. and Patten, C.L (2010) Molecular Biotechnology: Principles and Applications of Recombinant DNA 4th ed., ASM Press (Washington DC).
- 3. James D. Watson (1993) Recombinant DNA: 2nd Edition
- Prakash, M., and Arora, C.K. (1998) Cell and Tissue CultureAnmol Publications Pvt. Ltd.
- 5. Rema, L.P (2006). Applied BiotechnologyMJP Publishers
- Surzycki, S. (2003) Human Molecular Biology Laboratory Manual, Blackwell Publishing
- 7. Watson, J.D., Caudy, A.A., Myers, R.M. and Witkowski, J.A., (2007) Recombinant DNA:Genes and Genomes- A Short Course, Cold SpringHarbor Laboratory Press

CORE COURSE 9 TISSUE CULTURE AND GENE MANIPULATION (PRACTICALS)

36 Hrs

	0 Credit
1. Media formulation for plant tissue culture	
2. Surface sterilization.	(2 Hrs)
3. Callus induction.	(2 Hrs)
4. Auxillary bud culture.	(2 Hrs)
5. Isolation of protoplast.	(4 Hrs)
6. Isolation of genomic DNA and its quantification.	(10 Hrs)
7. Isolation of plasmid DNA.	(6 Hrs)
8. Restriction digestion, ligation, bacterial transformation.	

SEMESTER 4

ZB4CRT10 - CORE COURSE10 RADIOLOGY AND ADVANCED INSTRUMENTATION TECHNIQUES

54 Hrs 3 Credits

Objectives

- 1. To introduce the student to some of the radiological techniques and its applications
- 2. To develop an awareness about the harmful effects of radiation

Module 1

Radioactive materials: Types and sources of radiation. Effect of various types of radiation on biological systems. Radioactive emissions.

Module 2

10 Hrs

Isotopes: Definition, isotopes of common biological use, techniques for detection of isotopes and applications - [Autoradiography, Geiger counting technique, liquid scintillation, Gamma counter]

Use of radio isotopes in biological research, auto-radiography- pulse chase experiment.

Isotope dilution technique

Module 3

Management of radioactive wastes: waste disposal and cleaning of contaminated glassware. Precautions for handling, safety in use of radiation sources and radio isotopes.

Module 4

Newer techniques in microscopy:Confocal, Scanning probe microscopy, Flowcytometry & cell sorting.

Module 5

Chromatographic techniques: Theory, methods and application of paper, gas, affinity, ion exchange chromatography, TLC, HPLC, Gel filtration.

Electrophoresis: Theory, methods and applications of gelelectrophoresis: AGE, PAGE,

Polymerase chain reaction, DNA sequencing, DNA fingerprinting.

References

- Jones, M., Jones, Geoff, G. and Marchington, P (1999) Physics 2nd ed. Cambridge University Press
- Jones, M., Jones, Geoff, G, and Acaster D. (1999)Chemistry 2nd ed. Cambridge University Press
- 3. Blei, I and Odian, G (2006)General, Organic and Biochemistry- Connecting Chemistry to your Life 2nd Ed. W.H. Freeman and Company
- Kotz, J.C., and Treichel, P(1999) Chemistry and Chemical Reactivity 4th ed. Saunders College Publishing
- Wilson, K., and Walker, J (2000) Practical Biochemistry- Principles and Techniques5th ed. Cambridge Low Price Editions, Cambridge University Press

15 Hrs

10 Hrs

SEMESTER 4

ZB4CRP04 - CORE COURSE10 RADIOLOGY AND ADVANCED INSTRUMENTATIONTECHNIQUES (PRACTICALS)

36 Hrs 2 Credits

	Types and effects of various radiations. Isotope dilution techniques. (Visit	
	to a radioisotope lab.)	(14 Hrs)
~	2. Problems in radiology [on half cycle, quantity, disposal]	(14 Hrs)
	3. Agarose gelelectrophoresis: AGE	(8 Hrs)

SEMESTER 4

ZB4CRT11 - CORE COURSE 11 ENTREPRENEURSHIP DEVELOPMENT AND MARKETING

54 Hrs 3 Credits

Objectives

1. To understand the importance of marketing strategies and effects on entrepreneurial development.

Institutions, financing procedure and financial incentives.			
Module 2 10 Hrs			
Resource management: man, machine and materials, quality control/ quality assurance and			
testing of products			
Module 3 12 Hrs			
Elements of marketing & sales management [Nature of product and marketstrategy, packaging			
and advertising, after sales service]			
Module 4 12 Hrs			
Income tax, sales tax and excise rules			
Module 5 12 Hrs			

Need, scope and approaches for project formulation, structure of project reports.Project implementation, project report and appraisal. Network analysis –PERT,CPM Entrepreneur traits of entrepreneur

References

1. Khanna, O.P. and Sarup A.(1999)Industrial Engineering and Management, Dhanpat Rai Publications (P) Ltd.

2. Khanna, O.P (1999). Work Study, Dhanpat Rai Publications (P) Ltd.

3. Khanna, O.P (1999)Textbook of Mecahnical Estimating and Costing, Dhanpat Rai Publications (P) Ltd.

SEMESTER 4

CORE COURSE 11 ENTREPRENEURSHIP DEVELOPMENT AND MARKETING (PRACTICALS)

36 Hrs 2 Credits

1. Preparation and analysis of a project(14 Hrs)

2. Entrepreneurial motivation training through games, role playing, discussions and exercises

(8 Hrs) (14 Hrs)

3. Preparation of report on an industry/firm

SEMESTER IV

CORE COURSE 12

RESEARCH METHODOLOGY, BIOPHYSICS AND BIOSTATISTICS

54 Hrs

3 Credits

Objectives

- 1. To familiarise the learner the basic concept of scientific method in research process.
- 2. To have a knowledge on various research designs.
- 3. To develop skill in research communication and scientific documentation.
- 4. To create awareness about the laws and ethical values in biology.
- 5. To equip the students with the basic techniques of animal rearing collection and preservation
- 6. To help the student to apply statistical methods in biological studies.

RESEARCH METHODOLOGY

Module I

Basic concepts of research: Meaning, Objectives, Approaches, Types of research.

Research Process: Scientific method in research (eight steps).

Importance of literature reviewing in defining a problem,

Identifying gap areas from literature review.

Research Communication and scientific documentation: Project proposal writing,

Research report writing, (Structure of a scientific paper), Thesis, dissertation, research article.

Presentation techniques: Oral presentation, Assignment, Seminar, Debate, Workshop,

Colloquium, Conference

Sources of Information: Primary and secondary sources. Library- Books, Journals, Periodicals, Reviews, Internet.

Search engines Online libraries, e-Books, e-Encyclopedia, Institutional Websites. Plagiarism

Module II

12 Hrs

Animal Collection – Tools & techniques

Sampling techniques

Quadrate Line transect Measurements Density Abundance Frequency Biodiversity indices – concepts Simpson index

Collection methods, techniques and equipments

Plankton Insects Fish Bird Preservation techniques – Taxidermy Rearing techniques Laboratory and field.

Units of measurements- units, SI system, Equivalent weight, normality, molarity

Biophysics

Module III

14 Hrs

Basic understanding on principle and uses of the following:

Microscopy

(a) Light microscopy, Bright field (Compound Microscope), Phase contrast, Dark field microscopy, Fluorescence, Polorization microscopy, Video microscopy.

(b) Electron - Scanning (SEM), Transmission (TEM) and STEM

Micrometry – Stage and Eyepiece micrometers

Camera Lucida Instrumentation pH Meter Separation Techniques: Centrifuge, Chromatography, Electrophoresis Analytical techniques: Colorimeter, Spectrophotometer, X-ray crystallography

BIOETHICS

Module IV

Bioethics : Introduction, Animal rights and animal laws in India, Prevention of cruelty to animals Act 1960, Biodiversity Act 2003.

Concept of 3 R – conservation (Refined- to minimize suffering, Reduced – to minimize animals, Replaced – modern tools and alternate means), Animal use in research and education.

Laboratory animal use, care and welfare, Animal protection initiatives- Animal Welfare Board of India, CPCSEA, ethical commitment. Working with human: Consent, harm, risk and benefits.

BIOSTATISTICS

Module V

Sample & Sampling techniques: Collection of data, classification of data, frequency distribution tables, graphical representation: - Bar diagrams, Histogram, Pie diagram and Frequency curves - Ogives.

Measures of Central Tendency: Mean, Median, Mode (Problem - Direct method only)

Measures of dispersion: Range, Quartile Deviation, Mean Deviation, Standard Deviation,

Standard error. (Merits & demerits and problems on SD).

Correlation: Definition, Types of correlation.(mention in brief)

Test of Hypothesis and Test of Significance: Basic concept, Levels of significance, test of significance, Procedure for testing hypothesis, types of hypothesis- Null hypothesis and Alternate hypothesis.

10 Hrs

References :

1. Gupta K.C, Bhamrah, H.S and G.S.Sandhu (2006) Research Techniques in Biological Sciences. Dominant Publishers and Distributors, New Delhi.

2. Khan and Khanum, (1990) Fundamentals of biostatistics. Press, Chicago,

3. Rastogi, V.B (2009) Fundamentals of Biostatistics, Ane Books Pvt. Ltd. New Delhi.

4. Ackoff, R.L. (1962) Scientific Method, New York : John Wiley Press.

5. Aggarwal. S.K.(2009) Foundation Course in Biology, 2nd Ed.. Ane's Student Edition. Ane Books Pvt. Ltd.

6. Anderson, J, Durston, B.H. and Poole, M. (1992). Thesis and assignment writing. Wiley Eastern Ltd.

 Best, J.W.and K.V. James, (1986) Research in Education.5th Edn. Prentice- Hall of India Pvt.Ltd.

8. Campell, R. (1990). Statistics for biologists. CBS Publishers and distributors.

9. Day, R.A. (1993). How to write and publish a scientific paper. Cambridge University Press.

Day, R.A. (2000) Scientific English: A guide for Scientists and other Professionals.
 Universities Press.

11. Fischer, R.A.(1960)The Design of Experiment. 7th rev.edn. New York: Hafner Publishing Co.,

12. Hawkins C. and Sorgi, M. (1987). Research: How to plan, speak and write about it. Narosa Publishing House.

13. Killick, H.J. (1971). Beginning ecology. Ibadan University Press.

14. Kleinbaum, D.G. and M.Klein (2009) Survival analysis-Statistics for Biology & Health2nd Ed. Springer International ed.

15. Knudsen J. W (1966) Biological Techniques: Collecting, Preserving, and Illustrating Plants and Animals.

16. Kothari, C.R. and G.Garg. (2014) Research Methodology. Methods and Techniques. 3rd edn.

17. Marie, M. (2005). Animal Bioethics: Principles and Teaching Methods Wageningen Academic Publishers.

18. Norman T.J. (2007) Bailey Statistical methods in biology, Cambridge University press.

19. Roberts, M. T. King and M. Reiss.(1994) Practical Biology for Advance Level. Thomas Nelson and Sons Ltd. Surrey, UK.

20. Ruxton, G.D. and Colegrave, N. (2006), Experimental design for the life sciences. Oxford University Press.

21. Sateesh, M.K. (2008) Bioethics and Biosafety; I.K. International Publishing House .

22. Taylor D.J. Green N.P.O and Stout G.W. (2008). Biological science (3rd edition- R.S. Oper Ed). Cambridge University press.

PRACTICAL

RESEARCH METHODOLOGY, BIOPHYSICS AND BIOSTATISTICS

2 credits

PART A. RESEARCH METHODOLOGY

Animal collection Tools, Techniques & Estimation

- 1. Quadrate study
- 2. Transect study
- 3. Sampling Methods
- 4. Species area curve
- 5. Simpson index

PART B - BIOPHYSICS

- 6. Study of simple and compound light microscopes
- 7. Micrometry -calibration and measurement of microscopic objects -low power
- 8. Camera Lucida (draw a few diagrams using Camera Lucida)
- 9. Paper chromatography (demonstration only)
- 10. Instrumentation demonstration (write notes on principle, equipment and its use)

pH Meter

Colorimeter/ Spectrophotometer

Centrifuge

PART C BIOSTATISTICS

- 1. MS Excel : To create mean and median, Construction of bar diagram, Pie diagram and Line graphs.
- 2. MS Access: To create grade of students
- 3. Internet: Access a web page on any biological topic.
- 4. Frequency distribution of the given samples to find out arithmetic mean, median, mode.
- 5. Range and standard deviation for a biological data
- 6. Correlation using any biological data.
- 7. Graphical representation of data. Construction of bar diagrams, Histograms, Pie diagram and Line graphs.

SEMESTER V.

CORE COURSE 13 ENVIRONMENTAL BIOLOGY AND HUMAN RIGHTS (54 Hrs)

(0.111

Objectives

To instill the basic concepts of Environmental Sciences, Ecosystems, Natural Resources, Population, Environment and Society

To make the students aware of natural resources, their protection, conservation, the factors polluting the environment, their impacts and control measures.

To teach the basic concepts of toxicology, their impact on human health and remedial measures

To create a consciousness regarding Biodiversity, environmental issues & conservation strategies

To develop the real sense of Human rights – its concepts & manifestations

MODULE 1 ECOSYSTEM

12 Hrs

Basic concepts of ecosystem Components of ecosystem: Abiotic (Sunlight, temperature, soil, water, atmosphere) and Biotic components (Producers, consumers, decomposers), Ecological pyramid- number, biomass, energy, **Functions of ecosystem**: Productivity-Food chain-Food web-Energy flow-Laws of Thermodynamics.Types of Ecosystem: Terrestrial-Forest-Grassland-Desert, Aquatic-Marine-Fresh water, Wetland &Biome Concept of limiting factors: Liebig's and Shelford's laws of limiting factors.

Biogeochemical cycles: Concept, gaseous and sedimentary cycles, Carbon cycle, Nitrogen cycle. **Renewable resources** (solar,wind, hydroelectric, biomass and geothermal) **and Non renewable resources** (mineral and metal ore, fossil fuels)

MODULE 2 CONCEPTS OF POPULATION AND COMMUNITY 8 Hrs

Concept of population: Population attributes- Population growth forms, Basic concepts of growth rates, density, natality, mortality, growth curves

Animal interactions: Positive- Commensalism- Mutualism-Protocooperation, Negative-Predation-Parasitism-Competition-Antibiosis

Characteristics of a community: Species diversity- richness, eveness, stratification, dominance, ecological indicators, Ecotone and Edge effect, Keystone species, Concepts of Ecological Niche and Guild, Ecological succession, community evolution- climax.

MODULE 3 BIODIVERSITY AND ENVIRONMENTAL ISSUES 16 Hrs

Introduction to Biodiversity: Types of biodiversity- Alpha, Beta and Gamma diversity. **Concept and importance of Biodiversity**: Levels of Biodiversity-Species diversity, Genetic diversity, Microbial, Ecosystem diversity, India as a mega-diversity nation, Biodiversity hotspots

Global Environmental Issues: Ozone depletion, Greenhouse effect, Global warming, Climate change, Carbon trading, carbon credit; Carbon sequestration, Acid rain, Oil spills, Nuclear accidents, IPCC/UNFCC.

National Environmental issues: Deforestation, forest fire, pollution(air, water, soil, noise thermal, nuclear- brief account only) solid waste management, sewage, drinking water crisis and water logging,

Toxic products and disaster: Types of toxic substances – degradable, non degradable, Impact on human – case studies: Endosulphan tragedy, Bhopal disaster

Flood, drought, cyclone, earthquake and landslide (Management and mitigation)

Local Environmental issues: Landscape alteration, sand mining, quarrying, changing crop pattern, conversion of paddy lands,

Threats to water resources of Kerala: Degrading Mangrove and wetland ecosystems of Kerala, RAMSAR sites, Marine ecosystem crisis- pollution, overfishing etc. Impact of tourism on Environment.

MODULE 4CONSERVATION OF BIODIVERSITY12 Hrs

Protected area concept – Sanctuary, National Park, Biosphere reserve, Core Zone, Buffer Zone, Corridor concept. Conservation reserves

Concept of threatened fauna – IUCN categories - extinct, extinct in the wild, critically endangered, endangered, vulnerable, near threatened, least concern and data deficient. Red and Green Data Books.

Man-animal conflict (Tiger, Elephant, Dog, Monkey) - causes and concern

Water conservation- rainwater harvestiong, watershed management

Environment education

Environmental laws (Brief account only): The Water (Prevention and Control of Pollution) Act, 1974, The Air (Prevention and Control of Pollution) Act, 1981, Indian Forests Act (Revised) 1982. The Environment (Protection) Act, 1986, Hazardous Wastes (Management and Handling) Rules, 1989, The Forest (Conservation) Act, 1980, The Wildlife Protection Act, 1972, Biodiversity Act, 2002.

MODULE 5HUMAN RIGHTS6 Hrs

Introduction, main concepts associated with Human Rights, Different types of human rights,

227

Manifestations & phenomena, Role of agencies in promoting human rights, Mechanisms for checking violations of human rights, National human right commission, Constitutional provisions related to Human rights.

References

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Preservation' - PHI Learning, Environmental Law in India: Issues and Responses

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Paul R.C., 2000.Situations of Human Rights in India. Efficient offset printers. ·

Arun kumar Palai(1999) National Human Rights Commission of India, Atlantic publishers

Sharma P.D. (2005) Environmental biology and Toxicology, Rastogi publication

MeeraAsthana and Astana D.K.1990 Environmental pollution and Toxicology Alka printers.

Odum, E.P. 1971. Fundamentals of Ecology. W.B. Saunders College Publishing, Philadelphia

Alan Beeby, 2006 Anne – Maria Brennan First Ecology, Ecological principles and Environmental issues . International students edition Sec. edition Oxford University Press.

Robert Ricklefs (2001). The Ecology of Nature. Fifth Edition. W.H. Freeman and Company.

Stiling Peter (2002). Ecology: Theories and applications. Prentice Hall of India pvt.Ltd. New Delhi.

Landis, Wayne and Hing-hoYu, Baca Raton, 1995. Introduction to Environmental Toxicology: Impacts of chemicals upon Ecological systems: Lewis Publishers.

SEMESTER 5 PRACTICAL

ENVIRONMENTAL BIOLOGY & TOXICOLOGY 36 HRS CREDIT 1

- 1. Estimation of dissolved Oxygen
- 2. Estimation of carbon di oxide
- 3. Estimation of soil organic carbon (Demonstration only)
- 4. Identification of marine/ fresh water planktons
- 5. Counting of plankton using plankton counting chamber
- 6. Study of equipments Secchi disc, Plankton net
- 7. Study of sandy shore fauna, rocky shore fauna.
- 8. Study of animal Association
- Visit to any two important areas of bio diversity: 1. Forest, 2.Sea shore, 3. Mangrove, 3. Wet lands, 4. Bird sanctuary, 5. Wild life sanctuary, 6. Sacred groves
- 10. Field study (compulsory)

SEMESTER V.

CORE COURSE 14 CELL BIOLOGYAND GENETICS

54 Hrs

Credits 3

Objectives

- 1. To understand the structure and function of the cell as the fundamentals for understanding the functioning of all living organisms.
- 2. To make aware of different cell organelles, their structure and role in living organisms.
- 3. To develop critical thinking, skill and research aptitudes in basic and applied biology
- 4. To emphasize the central role of genes and their inheritance in the life of all organisms.

CELL BIOLOGY

Module I

Introduction of cell and Diversity of cells: History, Cell theory, Prokaryotes, Eukaryotes, Mycoplasmas, Virus, Virions and Viroids, Prions.

Cell membrane & Permeability: Molecular models of cell membrane (Sandwich model, Unit membrane model, Fluid mosaic model). Cell properties - permeability, Transport [Diffusion, Osmosis, Passive transport, Active transport, bulk transport], Cell coat and Cell recognition.

Module II

Cell Organelles :Structure and functions of following cell organelles: Endoplasmic reticulum -Structure and functions. Ribosomes (Prokaryotic and Eukaryotic) Golgi complex - Structure and functions. Lysosomes - Polymorphism - GERL concept, functions. Mitochondria -Structure and functions. Nucleus: Structure and functions of interphase nucleus, Nuclear membrane, pore complex, structure and functions of nucleolus

Chromosomes – Structure & organization, Heterochromatin, Euchromatin, Nucleosomes, Polytene chromosomes-Balbiani rings, Endomitosis, Lamp brush chromosomes.

Module III

Cell Communication: Basic principles of cell communications, Cell signaling(in brief), Types of signaling, Mention signaling molecules (neurotransmitters, hormones, Growth Factors, Cytokines Vitamin A and D derivatives),

Cell Division: Cell cycle - G_1 , S, G_2 and M phases, Mitosis and Meiosis. The difference between Mitosis and Meiosis.

References:

1 Zoological Society of Kerala Study material. 2002. Cell Biology, Genetics and

Biotechnology

2. Karp, G. (2010). Cell and Molecular Biology: Concepts and Experiments. VI Edition.

22 HRS

10 Hrs

6 Hrs

John Wiley and Sons.Inc.

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4. Sarada K & Mathew Joseph (Editors) (1999) *Cell Biology, Genetics and Biotechnoloy*,

.5. Thomas A.P (Editor) (2011) *Cell & Molecular Biology The Fundamentals*. Green leaf publications. TIES. Kottaya

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8. Ali, S (2014) The Cell: Organization Function and Regulatory Mechanisms ,Pearson

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Cell.VII Edition. Pearson Benjamin Cummings Publishing, San Francisco. 4

 Bruce Albert, Bray Dennis, Levis Julian, Raff Martin, Roberts Keith and Watson James (2008). *Molecular Biology of the Cell*, V Edition, Garland publishing Inc., New York and London.

11. Cooper, G.M. and Hausman, R.E. (2009). The Cell: A Molecular Approach. V Edition.

ASM Press and Sunderland, Washington, D.C.; Sinauer Associates, MA.

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.13. Gupta, P. K (2002) Cell and Molecular Biology, (2ed), , Rastogi Publications., Meerut

14. James Darnell. (1998) Molecular Biology. Scientific American Books Inc

15. Ariel G Loewy Philip Sickevitz, John R. Menninger and Jonathan A.N. Gallants (1991) cell structure and function. Saunder's College Publication

16. James Darnell. (1998) Molecular Biology.Scientific American Books Inc.

GENETICS

32 Hrs

10 Hrs

Module I

MendelianGenetics: Mendel's experiments- Monohybrid Cross, Dihybrid Cross, Mendel's Laws, Test Cross, Back Cross and Reciprocal Cross. Chromosome Theory of Inheritance

Interaction of genes:Allelic: Incomplete Dominance (Four O Clock Plant).Co- Dominance (Skin colour in Cattle) Lethal Alleles: Dominant lethal gene[Creeper chicken] and recessive lethal gene

[cystic fibrosis].

Non Allelic: Complementary (Flower colour in Sweet Pea), Supplementary (Coat colour in mice), Epistasis - dominant (Plumage in poultry) and recessive (Coat colour in mice). Polygenes (Skin colour inheritance in man), Pleiotropism (Vestigial wing gene in Drosophila).

Multiple alleles – ABO Blood group system, Rh group and its inheritance. Erythroblastosis foetalis.

Module II

12 Hrs

Sex determination: Chromosome theory of sex determination (Autosome and Sex chromosomes), male heterogamy and female heterogamy, (xx-xy, xx-xo, ZZ-ZW, ZZ-ZO), Genic Balance theory of Bridges. Barr bodies, Lyon's hypothesis, Gynandromorphism, sex mosaics, intersex (Drosophila), Hormonal [free martin in calf] and Environmental (Bonelia) influence on Sex determination

Recombination and Linkage: Linkage and recombination of genes based on Morgan's work in Drosophila, Linked genes, Linkage groups, Chromosome theory of Linkage, Types of linkage- complete and incomplete. Recombination, cross over value, chromosome mapping. [Definition]

Sex Linked inheritance : Characteristics of Sex Linked inheritance, X Linked inheritance of man (Hemophilia), Y linked inheritance [Holandric genes], Incompletely Sex Linked genes or pseudoautosomal genes (Bobbed bristles in *Drosophila*), Sex limited genes (Beard in man) and Sex influenced genes (inheritance of baldness in man).

Module III

10 Hrs

Mutation: Types of mutations - Somatic, germinal, spontaneous, induced, autosomal and allosomal, chromosomal mutations, structural and numerical changes. Gene mutations. [Addition, Deletion and substitution].

HumanGenetics: Karyotyping, Normal Human chromosome Complement, Pedigree analysis, Aneuploidy and Non- disjunction. Autosomal abnormalities (Down syndrome, Cry du chat syndrome) Sex chromosomal abnormalities (Klinefelters syndrome, Turner's syndrome) Single gene disorder (Brief mention) Autosomal single gene disorder [sickle cell anaemia), Inborn errors of metabolism such as phenylketonuria, alkaptonuria, , Albinism. Multifactorial traits – polygenic disorder- cleft lip and cleft palate.

Genetic Counseling, Eugenics and Euthenics -Brief account only

References:

1. Gardner, J.E., Simmons, J.M and Snustad D.P..(2007). *Principles of Genetics* (8th edn.). John Wiley and Sons, India.

- 2. Klug, W.S and Cummings, M.R. (2011). *Concepts of Genetics* (7th edn). Pearson Education Inc.India.
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4. Shirly Annie Oommen, Sampath Kumar S., and Jinsu Varghese (Editors) (2012), *Gene toGenome*. Zoological Society of Kerala, Kottayam.

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11. Hartl, L.D. and E.W.Jones. (2009). *Genetics: Analysis of Genes and Genomes* (7th edn) Jones and Barlett Publishers Inc, USA.

12. Primrose, S.B., Twyman, R.M. and Old, R.W. (2001). *Principles of Gene Manipulation* (6th edn.) Blackwell Science Ltd., London.

13. Sobti, R.C. and Pachauri, S.S. (2009). *Essentials of Biotechnology*. Ane's Book Pvt. Ltd.New Delhi.

14. Sinnat Dunn & Dobzhansky 1959. Principles of Genetics (T.M.H. New Delhi)

SEMESTER V

PRACTICAL

CELL BIOLOGY AND GENETICS

36 Hrs 2 Credits

PART A: CELL BIOLOGY

1. Squash preparation of onion root tip for mitotic stages

- 2. Mounting of polytene chromosome (Drosophila/Chironomous.) Demonstration
- 3. Tissues (permanent slides of epithelial tissues, striated muscle, smooth muscle,

cartilage, bone)

- 4. Identification of cell organelles
- 5. Preparation of temporary whole mount.
- 6. Preparation of permanent whole mount (demonstration)
- 7. Preparation of human blood smear and identification of Leucocytes

PART B :GENETICS

- 1. Genetic problems on Monohybrid, Dihybrid Crosses and Blood group inheritance
- 2. Study of normal male and female human karyotype (use photographs or Xerox copies)
- 3. Abnormal human karyotypes Down, Edwards, Klinefelter and Turner syndromes (use photographs or Xerox copies)
- 4. Sexing of Drosophila.
- 5. Study of Barr body in human buccal epithelium

SEMESTER V.

CORE COURSE - 15: EVOLUTION, ETHOLOGY & ZOOGEOGRAPHY

54 Hrs

Credits 3

Objectives:

- To acquire knowledge about the evolutionary history of earth living and nonliving
- To acquire basic understanding about evolutionary concepts and theories
- To study the distribution of animals on earth, its pattern, evolution and causative factors
- To impart basic knowledge on animal behavioural patterns and their role

Prerequisite:

- Basic knowledge on principles of inheritance and variation
- Knowledge on molecular basis of inheritance
- Basic understanding on the mechanism and factors affecting evolution
- Knowledge on origin and evolution of man

PART I - EVOLUTION

Module I - Origin of life

Theories - Panspermia theory or Cosmozoic theory, Theory of spontaneous generation (Abiogenesis or Autogenesis), Special creation, Biogenesis, Endosymbiosis. Chemical evolution - Haldane and Oparin theory, Miller-Urey experiment; Direct evidences of evolution - Recapitulation Theory of Haeckel, Fossilization, Kinds of fossils, fossil dating, Homologous organs and analogous organs.

Module II - Theories of organic evolution

Lamarckism and its Criticism, Weismann's Germplasm theory, Darwinism and its Criticism, Neo-Darwinism, Theory of De Vries,

Population genetics and evolution: Hardy-Weinberg Equilibrium, gene pool, gene frequency. Factors that upset Hardy-Weinberg Equilibrium, Effects of genetic drift on population: Bottleneck effect and founder effect

Module III – Nature of evolution

Species and Speciation: Species concept, subdivisions of species (sub species, sibling species, cline and deme), Speciation: Types of speciation, Phyletic speciation (autogenous and allogenous transformations), True speciation, Instantaneous and gradual speciation, allopatric and sympatric speciation

Isolation: Types of isolating mechanisms-Geographic isolation (mention examples) and Reproductive isolation. Role of isolating mechanisms in evolution

Microevolution, Macroevolution (Adaptive radiation -Darwin finches) Mega evolution, Punctuated equilibrium, Geological time scale, and Mass extinction (brief account only). **Evolution of Horse**

PART II - ETHOLOGY	14 Hrs
Module IV- Introduction	1 Hr

Definition, History and scope of ethology

Module V – Learning, imprinting and behaviour

Types of learning with examples; patterns of behaviors – types of rhythms, navigation, homing instinct, hibernation, aestivation; pheromones- types and their effect on behavior, hormones and their action on behavior (aggressive and parental behavior)

Module VI – Social organization

Social organization in insects (ants) and mammals (monkey), Courtship behaviour and reproductive strategies

PART III - ZOOGEOGRAPHY

8 Hrs

9 Hrs

13 Hrs

9 Hrs

4 Hrs

Module VII – General Topics

Continental drift theory, Types and means of animal distribution, Factors affecting animal distribution; insular fauna – oceanic islands and continental islands,

Module VIII - Zoogeographical realms

Palaearctic region, Nearctic region, Neotropical region, Ethiopian region, Oriental region, Australian region (brief account with physical features and fauna, Wallace's line, Weber's line, Biogeography of India with special reference to Western Ghats

References:

EVOLUTION

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- 2. Barnes, C.W. (1988). Earth, Time and Life. John Wiley & Sons, NewYork
- 3. Bendall, D. S. (ed.) (1983). Evolution from Molecules to Man. Cambridge University Press, U.K.
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- 13. Strickberger, M.W.2000. Evolution. Jones and Bartlett, Boston.

ETHOLOGY

- 1. Agarwal. V. K. (2009). Animal Behaviour.S.Chand and Company Pvt. Ltd., New Delhi.
- Bonner, J.T. (1980). The Evolution of Culture in Animals. Princeton University Press.NJ, USA.

4 Hrs

- 3. David McFarland. (1999). Animal Behaviour. Pearson Education Ltd. Essex, England.
- 4. Dawkins, M.S. (1995). Unravelling Animal Behaviour. Harlow: Longman.
- 5. Dunbar, R. (1988). Primate Social Systems. Croom Helm, London.
- Gundevia J.S. and Singh H.G. (1996), A Text Book of Animal Behaviour. S. Chand and Company Pvt. Ltd., New Delhi.
- Aubrey M. and Dawkins M.S. (1998). An Introduction to Animal Behaviour. Cambridge University Press,UK.
- Sherman P.W and Alcock J., (2001) Exploring Animal Behaviour- Readings from American Scientist 3rd Edn. Sinauer Associates Inc. MA,USA. (Module 10 & 11).
- Wilson, E.O. (1975). Sociobiology.Harvard University Press, Cambridge, Mass. USA.(Module 9).

ZOOGEOGRAPHY

- 1. Briggs, J.C. (1996). Global Biogeography. Elsevier Publishers. (Module VI and VII).
- Chandran Subash M.D. (1997). On the ecological history of the Western Ghats.Current Science, Vol.73, No.2.146-155.
- Chundamannil Mammen.1993, History of Forest management in Kerala. Report No.89. Kerala Forest Research Institute, Peechi, India.
- Daniels, R.J.R and Vencatesan J. (2008), Western Ghats Biodiversity. People Conservation; Rupa& Co. New Delhi. India.
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- Tiwari, S. (1985), Readings in Indian Zoogeography (vol.1). Today & Tomorrow Printers & Publishers

PRACTICAL

EVOLUTION, ETHOLOGY & ZOOGEOGRAPHY

36 Hrs

Credit 1

- 1. Identification of Zoogeographical realms using map
- 2. Study on endemic species of each realm

- 3. Show the discontinuous distribution of (lung fishes, camel, elephant)
- 4. Providing a map trace the route of HMS Beagle
- Providing a map mark any two continental/oceanic islands.: Greenland, Madagascar, New Zealand, New Guinea, Maldives, Iceland, Hawaii – any two
- 6. Contributions of scientists (showing photos) Any four
- 7. Identification of different stages of horse evolution
- 8. Study on Homology and Analogy
- 9. Study on connecting links (Peripatus, Archaeopteryx, Protopterus, Echidna)
- 10. Pheromone traps
- 11. Skinner box & T Maze
- 12. Experiment to demonstrate phototaxis and chemotaxis using Drosophila/House fly
- 13. Identification of behaviour (Grooming/courtship dance of flamingos/stickle back fish/ Tail wagging dance/ Aggressive behaviour/ Auto/Allo grooming, Flehmen response) showing pictures (Any five)

SEMESTER V.

CORE COURSE VIII

HUMAN PHYSIOLOGY, BIOCHEMISTRY AND ENDOCRINOLOGY

54 Hrs

Credits 3

Objectives:

- 1. This course will provide students with a deep knowledge in biochemistry, physiology and endocrinology.
- 2. Defining and explaining the basic principles of biochemistry useful for biological studies for illustrating different kinds of food, their structure, function and metabolism.
- 3. Explaining various aspects of physiological activities of animals with special reference to humans.
- 4. Students will acquire a broad understanding of the hormonal regulation of physiological processes in invertebrates and vertebrates.
- 5. By the end of the course, students should be familiar with hormonal regulation of physiological systems in several invertebrate and vertebrate systems.

- 6. This also will provide a basic understanding of the experimental methods and designs that can be used for further study and research.
- 7. The achievement of above objectives along with periodic class discussions of current events in science, will benefit students in their further studies in the biological/physiological sciences and health-related fields, and will contribute to the critical societal goal of a scientifically literate citizenry.

HUMAN PHYSIOLOGY

Module I

Nutrition: Nutritional requirements – carbohydrates, proteins, lipids, minerals (Ca, P, Fe, I), vitamins (sources and deficiency disorders). Importance of dietary fibre and antioxidants.Balanced diet, Recommended Dietary Allowance (RDA). Nutrition during pregnancy and lactation, Infant nutrition, Malnutrition(PEM).

Digestion: Anatomy and histology of digestive glands (liver, pancreas, salivary, gastric and intestinal). Digestion and absorption of carbohydrates, proteins and fats.Nervous and hormonal control of digestion.

Module II

Respiration: Phases of respiration (external respiration, gas transport and internal respiration). Respiratory pigments: Haemoglobin, Myoglobin (Structure and Function). Transport of respiratory gases - transport of oxygen, oxyhaemoglobin curve, factors affecting oxyhaemoglobin curve, transport of carbon dioxide,(chloride shift). Control of respiration.Respiratory disturbances (Hypoxia, Hypercapnia, Asphyxia).Physiological effect of smoking, carbon monoxide poisoning, Oxygen therapy and artificial respiration.

Circulation: ESR, Haemopoiesis, blood pressure, ECG. Haemostasis (blood coagulation) clotting factors, intrinsic and extrinsic pathways, anticoagulants and its mechanism of action.Cardiovascular diseases (Jaundice, Atherosclerosis, Myocardial infarction, Thrombus, Stroke). Angiogram and angioplasty.

Module III

5 Hrs

Excretion: Histology of Bowman's capsule and tubular part. Urine formation – glomerular filtration, tubular reabsorption, tubular secretion.Urine concentration - counter current

31 Hrs

8 Hrs

mechanism. Acid – base balance, hormonal regulation of kidney function. Renal disorders (kidney stone, acute and chronic renal failure, and dialysis). Homeostasis: Definition, concept and importance in biological system. Thermal regulation and thermal adaptation in homeotherms.

Module IV

Nerve physiology: Ultra structure of neuron. Nerve impulse production (resting membrane potential, action potential), transmission of impulse along the nerve fiber, interneuron (synaptic) transmission, neuromuscular junction and transmission of impulses.Neurotransmitters (acetyl choline, adrenalin, dopamine).EEG. Memory, Neural disorders (brief account on Dyslexia, Parkinson's disease, Alzheimer's disease, Epilepsy).

Muscle physiology: Ultra structure of striated muscle, muscle proteins (myosin, actin, tropomyosin, troponin), Muscle contraction and relaxation-Sliding Filament Theory, cross bridge cycle, biochemical changes and ATP production in muscle, Cori cycle. Kymograph, Simple muscle twitch, muscle fatigue, tetanus, rigor mortis.

BIOCHEMISTRY

Module V

Carbohydrates: Basic structure, biological importance and classification of monosaccharides, oligosaccharides, polysaccharides with examples.

Proteins: Basic structure and classification of amino acids; structure, biological importance and classification of proteins with examples.

Lipids: Structure of fatty acid, saturated and unsaturated fatty acid, biological importance and classification of lipids with examples.

Vitamins and minerals: Major fat soluble and water soluble vitamins. Important minerals and trace elements required for living organisms. Biological importance of vitamins and minerals.

Enzymes: Chemical nature of enzymes, enzyme activation, enzyme inhibition, allosteric enzymes, isoenzymes, co-enzymes. Michaelis–Menten enzyme kinetics.

Module VI

10 Hrs

15 Hrs

5 Hrs

Carbohydrate metabolism: Glycogenesis, Glycogenolysis, Gluconeogenesis, Hexose monophosphate Shunt, Glycolysis, Citric Acid Cycle, Electron Transport Chain and ATP synthesis. Ethanol metabolism.

Protein metabolism: Deamination, Transamination, Transmethylation, Decarboxylation, Ornithine cycle.

Lipid metabolism: Biosynthesis of fatty acids, Beta oxidation, physiologically important compounds synthesized from cholesterol.

ENDOCRINOLOGY

Endocrinology and reproduction	8 Hrs
Module VII	8 Hrs

Endocrine physiology: Hormones – classification and mechanism of hormone action. Major endocrine glands(Histology is not included) their hormones, functions and disorders (hypothalamus, pituitary gland, pineal gland, thyroid gland, parathyroid gland, islets of Langerhans, adrenal gland),. Homeostasis and feedback mechanism.

References:

Albert L. Lehninger, Michael Cox and David L. Nelson; 2004; Biochemistry Lehninger.

Palgrave – Macmillan.

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Geetha N. 2014. Textbook of Medical Physiology:. Paras Medical Publishers, 3rd edition

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Melmed, Shlomo, Williams, Robert Hardin; 2011; Textbook of Endocrinology: Elsevier, 12th edition.

PRACTICAL

HUMAN PHYSIOLOGY, BIOCHEMISTRY AND ENDOCRINOLOGY

36 Hrs Credit1

HUMAN PHYSIOLOGY

- 1). Determination of haemoglobin content of blood
- 2). Total RBC count using Haemocytometer
- 3). Total WBC count using Haemocytometer
- 4). Estimation of microhaematocrit
- 5). Effect of hypertonic, hypotonic and isotonic solutions on the diameter of RBC.
- 6). Instruments: Kymograph, Sphygmomanometer and Stethoscope (principle and use)
- 7). Measurement of blood pressure using sphygmomanometer(demonstration only)

BIOCHEMISTRY

- 1. Qualitative analysis of protein, glucose, starch and lipids.
- Chromatography Determination of Rf value of amino acids and identification of amino acids (Identify the Amino Acids using different solvent front and solute front)

ENDOCRINOLOGY

- 1. Cockroach Corpora cardiaca & Corpora allata (Demonstration)
- 2. Effect of adrenalin on heart beat of Cockroach (Demonstration)

SEMESTER VI.

CORE COURSE 17 DEVELOPMENTAL BIOLOGY

54 Hrs

3 Credits

Objectives:

- 1. To achieve a basic understanding of the experimental methods and designs that can be used for future studies and research.
- To provide the students with the periodicclass discussions of current events in science which will benefitthem in their future studies in the biological/physiological sciences and health-related fields
- 3. To contribute tocritical societal goal of a scientifically literate citizenry.

Module1

10 Hrs

Introduction: Definition, Scope of developmental biology, sub-divisions (descriptive, comparative, experimental and chemical), historical perspectives, basic concepts and theories.

Reproductive Physiology: Gonads- anatomy of testis and ovary, spermatogenesis, oogenesis, gonadal hormones and their functions. Hormonal control of human reproduction - Female reproductive cycles (Estrous cycle, Menstrual cycle). Structure of mammalian sperm and egg, Pregnancy, parturition and lactation.Reproductive health and importance of sex education.

Egg types: Classification of eggs based on the amount, distribution and position of yolk. Mosaic and regulative, cleidoic and noncleidoic eggs.Polarity and symmetry of egg.

Fertilization: Mechanism of fertilization-(Encounter of spermatozoa and Ova, Approach of the Spermatozoon to the Egg, Acrosome Reaction and Contact of Sperm and Ovum, Activation of Ovum, Migration of Pronuclei and Amphimixis,), Significance of fertilization, Polyspermy, Parthenogenesis- Different types and significance.

Module II

14 Hrs

Cleavage: Types, planes and patterns of cleavage, Cell lineage of Planaria. Influence of yolk on cleavage.

Blastulation: Morula, blastula formation, types of blastula with examples.

Fate maps: Concept of fate maps, construction of fate maps (artificial and natural), structure of a typical chordate fate map. Significance of fate map.

Gastrulation: Major events in gastrulation. Morphogenetic cell movements. Influence of yolk on gastrulation. Exogastrulation.Concept of germ layers and derivatives.

Cell differentiation and gene action: Potency of embryonic cells (Totipotency,

Pleuripotency, Unipotency of embryonic cells). Determination and differentiation in embryonic development, Gene action during development with reference to Drosophila (maternal effect genes), Zygotic genes.

Module III

20 Hrs

Embryology of Frog: Gametes, fertilization, cleavage, blastulation, fatemap, gastrulation, neurulation, notogenesis. Differentiation of Mesoderm and Endoderm, Development of eye.Metamorphosis of frog, Hormonal and environmental onrol.

Embryology of chick: Structure of egg, fertilization, cleavage, blastulation, fate map, gastrulation. Development and role of Primitive streak, Salient features of 18hour, 24 hour, 33 hour & 48 hour chick embryo.Extra embryonic membranes in chick.

Human development: Fertilisation, cleavage, blastocyst, implantation, placenta. Gestation, parturition and lactation.Human intervention in reproduction, contraception and birth control. Infertility, Invitro fertilization (test tube baby)

Module IV

Experimental embryology: Spemann's constriction experiments, Organizers and embryonic induction. Embryo transfer technology, cloning, stem cell research. Ethical issues.

Teratology / Dysmorphology, Developmental defects: Teratogenesis, important teratogenic agents.(Radiations, chemicals and drugs, infectious diseases) genetic teratogenesis in human beings,

Developmental defects: Prenatal death (miscarriage and still birth). Intrauterine Growth Retardation (IUGR).

Module V

General topics: Classification and functions of placenta in mammals. Prenatal diagnosis (Amniocentesis, Chorionic villi sampling, Ultra sound scanning, Foetoscopy, Maternal serum alpha-fetoprotein, Maternal serum beta-HCG).Regeneration in animals.

5 Hrs

References:

Anthony S. Fauci, Eugene Braunwald, Dennis L. Kasper, Stephen L. Hauser, Dan L. Longo, J. Larry Jameson and Joseph Loscalzo; 2008; Harriosns Principles of Internal Medicine; Chruch Livingston 17th Ed.

Balnisky B.I.; 1981 An Introduction to Embryology, W.B. Saunders and Co.

Berril, N..J.; and Kars, G.; 1986. Developmental biology, Mc Graw Hills

Dutta 2007 Obstrestics, Church Livingston 17 Ed

Majumdar N. N -1985 Vetebrate embryology; Tata McGraw-Hill, New Delhi

Melissa A & Gibbs, 2006; A practical Guide to Developmental Biology, Oxford university press (Int. student edition)

Scott F. Gilbert; 2003; Developmental biology; Sinauer Associates Inc.,U.S.; 7th Revised edition.

Vijayakumarn Nair, K. & George, P. V. 2002. A manual of developmental biology, Continental publications, Trivandrum

Taylor D J, Green NPO & G W Stout. (2008) Biological Science third edition. Cambridge university press. Ref pp 748 biology 755

PRACTICAL

DEVELOPMENTAL BIOLOGY

36 Hrs

Credit 1

Model/Chart/ Slide may be used

- 12. Embryological studies- Blastula (frog, chick)
- 13. Embryo transfer, cloning, gastrula (frog, chick)
- 14. Amniocentesis
- 15. Embryotransfer technology, cloning
- 16. Study of placenta- pig and man
- 17. 18 hour, 24 hour, 33 hour and 48 hour chick embryo.

- 18. Candling method.
- 19. Vital staining- demonstration.
- 20. Male and female reproductive organs in cockroach
- 21. Calculate the fecundity of fish.
- 22. Calculate the gonado-somatic index of given fish.

SEMESTER VI.

CORE COURSE 18. MICROBIOLOGY & IMMUNOLOGY

54 Hrs

MICROBIOLOGY

Module I

Introduction: History and scope of microbiology. Outline classification of Microbes. (bacteria, fungus & virus)

Methods in Microbiology: Sterilization and disinfection - physical and chemical methods.

Culture media – selective media, enrichment media, differential media.Plating techniques and isolation of pure colony. Culture preservation techniques: refrigeration, deep freezing, freezing under liquid nitrogen, lyophilization.

Module II

15 Hrs

10 Hrs

Morphology and fine structure of bacteria: Size, shape, cilia, pili, flagella, capsule, cell

wall and its composition. Cytoplasmic membrane, protoplast, spheroplast, intracellular

membrane systems, cytoplasm, vacuoles, genetic material, cell inclusions, bacterial spores.

Bacterial growth Curve, Staining techniques - gram staining.

Bacterial Reproduction Sexual – (conjugation, transduction) and Asexual (budding,, fragmentation). Virology: Structure of virus; Human, animal, and bacterial virus. Viral replication, cultivation of animal viruses.

Module III

8 Hrs

Infections & Diseases: Types of infections – primary, secondary and nosocomial infections.

(Brief Account only)Contagious diseases – epidemic, endemic and pandemic, mode of Transmission – food, water, air, vectors and carriers.

Diseases: Epidemiology, symptomology, diagnosis and treatment. Bacterial - Clostridium tetany (tetanus), Viral – HIV virus (AIDS), fungal –*Candida albicans* (candidiasis).

IMMUNOLOGY

Module IV

9Hrs

Introduction to Immunology: Innate and acquired immunity, passive (natural and artificial) and active immunity (Natural and Artificial).Mechanisms of innate immunity - barriers, inflammation, phagocytosis.

Lymphoid organs: Primary (Thymus, Bone marrow) and secondary lymphoid organs (lymph nodes, spleen).

Lymphocytes: T and B cells, Natural killer cells, memory cells, macrophages.

Module V

Antigens, Types of antigens, haptens, adjuvants, immunoglobulin structure, classes and functions of immunoglobulins.

Types of Immunity-, humoral & cell mediated immunity Monoclonal & polyclonal antibodies

Antigen – antibody reactions, Precipitation test, Agglutination test, VDRL WIDAL, ELISA. Auto immune diseases: Pernicious Anemia, Rheumatoid Arthritis. Immunodeficiency -

AIDS. Hyper sensitivity- Type I, (E.g. Anaphylaxix) II(Transfusion reaction), III (Arthus reaction) and IV (Mantaux Test) (in brief).

Vaccines

Introduction Types of vaccines, Current Vaccines, Recent trends in vaccine preparation

References

1. Ananthanarayan R & Jayaram Paniker C K. (2009) Text Book of Microbiology Orient

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9Hrs

Longman Private Ltd.

2. Gladys Francis & Mini K.D., (Editors) (2012), Microbiology, Zoological Society of Kerala, Kottayam.

3. Kuby J, Kindt T., Goldsby R. and Osborne B. (2007). Kuby immunology

4. Sharma K. (2005) Manual of Microbiology: Tools and Techniques, Ane books

5. Susan Panicker & George Abraham (Editors) (2008), Micro Biology and Immunology,

Zoological Society of Kerala, Kottayam.

6. Colemen: (2002). Fundamentals of Immunology

7. Darla J. Wise & Gordon R. Carter: (2004): Immunology A Comprehensive Review Iowa state University Press. A Blackwell science company,

8. Hans G. Sch, Legal General Microbiology, Seventh Ed. Cambridge Low Price Ed.

9. Helen Hapel, Maused Harney Siraj Misbah and Next Snowden: (2006) Essentials of

Clinical Immunology Fifth Ed. Blackwell Publishing Company,

10. Heritage, J, E.G.V. Evaus and R.A.Killungten (2007): Introductory Microbiology

Cambridge University Press 6. Ivan Roitt I (2002) Essentials of Immunology ELBS.

MICROBIOLOGY AND IMMUNOLOGY PRACTICAL

72 Hrs

2 Credits

1. Instruments – Autoclave, Hot air oven, Bacteriological incubator – Laminar air flow

2. Preparation of solid and liquid media for microbial cultures.

(Ingradients, pH and method of preparation) (Demonstration)

(a) Solid media (1) Nutrient agar (2) Mac Conkey's agar

(b) Liquid Media(1) Nutrient broth (2) Peptone water.

3. Culture methods (Demonstration)

- (a) Streak plate technique and isolation of pure colonies.
- (b) Lawn culture (c) Pour plate culture (d) Liquid culture
- 4. Examination of microbes in living condition

Hanging drop method for demonstrating motility of bacteria.

- 5. Gram staining preparation, procedure, identification of Gram + ve and Gram –ve bacteria.
- 6. Antibiotic sensitivity test (demonstration only)
- 7. Streak plating (individual performance)
- 8. Preparation of a fungal smear Lactophenol cotton blue staining and mounting
- 9. Determination of ABO blood groups and Rh factor (Antigen antibody Reaction)

10. Study through photographs/ illustration, the primary immune (Bone marrow and thymus) and secondary immune (spleen and lymph nodes) organs in Rat/Man

SEMESTER VI. CORE COURSE 19

BIOTECHNOLOGY, BIOINFORMATICS & MOLECULAR BIOLOGY

BIOTECHNOLOGY

Module I

Introduction: Scope, Brief History, Scope and Importance

Tools and Techniques in Biotechnology: Enzymes (restriction endonucleases, ligases, linkers & adapters), Vectors-[Plasmids, Phage vectors, Cosmids, Artificial Chromosomes] Host cells. Basic steps & techniques in rDNA technology

Gene Libraries, Construction of genomic library and cDNA Library. PCR technique and DNA amplification, Brief description of screening methods – Probes, Nucleic Acid hydridization, In situ Hybridization, Fluorescence in situ Hybridization (FISH), Colony hybridization. Methods of transfer of desired gene into target cell.Blotting Techniques- Southern, Northern, Western blotting.DNA Finger printing (DNA Profiling) and its application. Molecular markers - RFLP

Module II

9 Hrs

20 HRS

Animal Cell Culture: Brief account on methods, substrates, media and procedure of animal cell culture, Stem Cells, types and potential use, Organismal Cloning- reproductive & therapeutic-brief account only.

Applications of Biotechnology: Applications in Medicine(insulin, growth hormone, gene therapy), Agriculture(GM plants and biopesticides),Environment(bioremediation), Industry (Single Cell Protein) and applications of Fermentation Technology- lactic acid, vitamins, food and beverages.

Potential Hazards of Biotechnological Inventions: Risks related to genetically modified organisms (GMO) and biologically active products, Biological warfare & Biopiracy. Protection of biotechnological inventions.Intellectual Property Rights, Patenting and patent protection.

References

1. Singh B.D Biotechnology 2002. Kalyan Publishers New Delhi.

2. Brown C.H., Campbell I & Priest F, G. 1987. Introduction of Biotechnology (Blackwell scientific publishers Oxford).

3. Colin Ratledge Bijorn Kristiansesn, 2008. Basic Biotechnology 3 rd ed. Cambridge University.

4. Janarathanan S & Vincent S. 2007. Practical Biotechnology, Method of Protocols. University Press.

5. John E. Smith. Biotechnology Cambridge Low priced ed. (Third Ed) 2005 Madingan, Martinko and Parker 2002, Biology of Microorganisms, Brock Eighth Ed. Prentice Hall.

6. Singh B.D. Biotechnolgy 2002, Kalyan Publishers New Delhi.

7. Sudha Gangal 2007. Biotechnology Principles and & practice of Animal Tissue culture, Universities Press.

BIOINFORMATICS

Module III

8 Hrs

Introduction: Definition, importance and role of bioinformatics in life sciences. Computational Biology.

Biological databases: Nucleotide sequence databases (NCBI- GENBANK, DDBJ and EMBL). Protein databases - structure and sequence databases (PDB, SWISSPROT and UNIPROT). Introduction to Sequences alignments: Local alignment and Global alignment, Pair wise alignment (BLAST and FASTA] and multiple sequence alignment. Phylogenetic Tree construction and Analysis

Module IV

Molecular visualization software - RASMOL. Basic concepts of Drug discovery pipe line, computer aided drug discovery and its applications. Human Genome Project.

MOLECULAR BIOLOGY

Module V

Nature of Genetic Materials: Discovery of DNA as genetic material – Griffith's transformation experiments. Avery Macarty and Macleod, Hershey Chase Experiment of Bacteriophage infection, Prokaryotic genome; Eukaryotic genome.Structure and.types of DNA & RNA.DNA replication. Modern concept of gene (Cistron, muton, recon, viral genes)., Brief account of the following-- Split genes (introns and exons), Junk genes, Pseudogenes, Overlapping genes, Transposons.

Module VI

Gene Expressions: Central Dogma of molecular biology and central dogma reverse, one geneone enzyme hypothesis, One gene-one polypeptide hypothesis Characteristics of genetic code, Contributions of Hargobind Khorana.

Protein synthesis [prokaryotic]: Transcription of mRNA, Reverse transcription, post transcriptional modifications, Translation, Post translational modifications.

Gene regulations: Prokaryotic(inducible & repressible systems) Operon concept -Lac operon and Tryptophan operon, Brief account of Eukaryotic gene regulation.

251

6 Hrs

12 Hrs

20 Hrs

References

1. Bruce Albert, Bray Dennis, Levis Julian, Raff Martin, Roberts Keith and Watson James (2008). Molecular Biology of the Cell, V Edition, Garland publishing Inc., New York and London.

2. De Robertis, E.D.P. and De Robertis, E.M.F. (2006).Cell and Molecular Biology.VIII Edition. Lippincott Williams and Wilkins, Philadelphia.

3. Gupta, P. K (2002) Cell and Molecular Biology, (2ed), , Rastogi Publications., Meerut

4. James Darnell. (1998) Molecular Biology. Scientific American Books Inc

5. Thomas AP(Editor). 2011 Cell & Molecular Biology The Fundamentals. Green leaf publications .TIES Kottayam

6. Zoological Society of Kerala Study material. (2011) Cell and Molecular Biology

PRACTICAL SYLLABUS.

(BIOTECHNOLOGY, BIOINFORMATICS & MOLECULAR BIOLOGY)

BIOTECHNOLOGY

- 1. Identify and comment on the item provided: (Western blotting / Southern blotting / Northern blotting / PCR)
- 2. Write down the procedure involved in DNA isolation

BIOINFORMATICS

- 1. Download/use print out/pictures of genome sequences of any 2 organisms. Identify and mention the characteristic features of both.
- 2. Download/ use print out/pictures of a protein sequence , identify it & comment on its amino acid composition
- 3. Download / use print out/pictures of a macromolecule. Write a brief note on the

bioinformatics tool used to visualize its structure.

MOLECULAR BIOLOGY

1. Identify and comment on its molecular composition / structural orientation / functional significance (Any tissue / Cell organelles/ DNA, DNA replication, RNA different types using models or diagrams)

V1 SEMESTER. CORE COURSE 20. OCCUPATIONAL ZOOLOGY . (APICULTURE, VERMICULTURE, QUAIL FARMING & AQUACULTURE)

54 Hrs Credits 3

Objectives:

- 1. To equip the students with self employment capabilities.
- 2. To provide scientific knowledge of profitablefarming.
- 3. To make the students aware of cottage industries.

Module 1. APICULTURE

Definition, Different species of honey bees, Organization of honey bee colony, Social life and adaptation of honey bees. Communication among honey bees. Bee keeping methods and equipments, Management and maintenance of an apiary, Growth period, honey flow period and dearth period Division of the colony, uniting two colonies, , replacing old queen with new queen, swarming management, monsoon management. Enemies of bees. Diseases of bees, Bee pasturage. Uses of honey bees, By-products of honey bees, Honey and wax composition. Testing the quality of honey.Extraction of wax, Uses of honey and wax.Royal jelly, Propolis. Apitherapy, Agencies supporting apiculture.

Activity :Visitto an apiculture unit. Field visit and report submission - 10 Hrs Field visit and report submission on any two items are taken for internal evaluation.

MODULE: 2. VERMICULTURE

8 Hrs

18 Hrs

Introduction, Ecological classification of earth worms. Species of earth worms used for vermicultre, Reproduction & life cycle, Role of earth worm in solid waste management, in agriculture, in medicine etc. Preparation of vermibed, Maintenance & monitoring, Preparation of vermicompost, Preparation of vermiwash.

Activity : Submission of a report after preparing a vermiculture unit or visiting a vermicomposting unit.

MODULE: 3.QUAIL FARMING (Coturnix coturnix) 4 hrs

Introduction, care of quail chicks, care of adult quails, care of breeding quails, ration for quail, care of hatching eggs, health care, use of quail egg and meat.Sources of quality chicks.

24 Hrs

MODULE: 4. AQUACULTURE.

Advantages and salient features of aquaculture, Types of Aquaculture, Biotic and abiotic features of water, Importance of algae in aquaculture, Common cultivable fishes of Kerala, Fish diseases, Composite fish culture, Integrated fish culture, Carp culture, Prawn culture Mussel culture Pearl culture. Processing & Preservation.

Aquarium management - Setting up of an aquarium, Biological filter & Aeration, Breeding of gold fish, gourami (Osphronemus), fighter and Guppy (live bearer). Nutrition and types of feed for aquarium fishes, Establishment of commercial ornamental fish culture unit. Fish Transportation - Live fish packing and transport Common diseases of aquarium fishes and their management. Aquaponics (a brief introduction only).

Activity - Setting up of an Aquarium

Field visit – Visiting an Aquaculture farm

References:

NPCS Board, The complete book on Bee keeping and honey processing, NIIR Project consultancy services, 106E, Kamala nagar, Delhi- 110007. Shukla G.S, & Updhyay V.B, Economic zoology ,Rastogi Publ. Meerut. Pradip.V.Jabde , Text book of applied zoology, 2005 Applied Zoology, Study Material Zoological Society of Kerala , CMS college Campus Clive. A Edwards, Norman. Q. & Rhonda. 2011. Vermitechnology: earthworms, organic waste & environmental management. Chauhan, H.V.S. Poultry, Disease, diagnosis and treatment, Wiley eastern Ltd Delhi. Otieno.F.O 2014. Quail farming: markets & market strategies Pillai T.V.R., Aquaculture, principles and practices. Ronald j. Roberts (1978) Fish pathology, Cassel Ltd London. Cowey C. B. et. al. (1985) Nutrition and feeding in fishes, academy press. Farm made aquafeeds. FAO fisheries Technical paper, 343. Harisankar J. Alappat& A. Bijukumar, Aquarium Fishes. B. R. Publ. Corporation, Delhi. MPEDA, A hand Book on AquafarmingOrnamentalfishes, MPEDA, Kochi. Amber Richards. 2014. Aquaponics at home. Pradip.V.Jabde. 1993. Text book of applied zoology Venkitaraman, P.R, 1983, Text book of Economic zoology(SudharsanaPuubl. Kochi) Addison Webb, Bee Keepingfor profit and pleasure, Agrobios Ltd. Edwards.C.A.&Lafty, J.R.1972 Biology of earthworms(Chapman & Hall Led.London) Applied Zoology, Study Material Zoological Society of Kerala, CMS college Campus George cust& Peter Bird, Tropical Fresh water Aquaria, Hamlyn London. Verreth J. Fish larval nutrition, Chapman & Hall Publ. Bone Packer. 2014. Aquaponic system

PRACTICAL

Occupational Zoology

36 Hrs

Credit 1

- 1. General Identification, Economic importance, Morphology, scientific names and common names of the following
 - f) Economic important and morphology of culturable fishes (Catla, Rohu, Grass carp, Common carp, Silver carp, *Etroplus suratensis*, *Oreochromis /Tilapia, Mugil cephalus* and *Anabas Testudineus*)
 - g) Identification and morphology of ornamental fishes (gold fish, fighter, Gourami, Angel fish, Guppy
 - h) Two species of earthworms used in Vermiculture

- i) Four species of honey bees
- j) Economic importance and morphology of shell fishes (Any three species of prawn, two marine mussels, two oysters one rock oyster - *Crasostria* and pearl oyster -*Pinctada fucata* and freshwater mussel - *Lamellidens marginalis*).
- 2. Castes of bees
- 3. Principle & uses of Aquarium filters, Aquarium aerator, Aquarium plants, Oven, Pelletiser, Screw Press, die plate
- 4. Identification and study of fish parasites and diseases (five numbers each) using slides/pictures
- 5. Bee keeping equipments, Beehive, Smoker, honey extractor, Queen Cage,
- 6. Bees wax, Honey, Vermicompost (Identification-Uses)
- 7. Formulation of artificial feed for aquarium fishes demonstration
- 8. Tests for determining the adulteration in honey.
- 9. Mounting of pollen basket
- 10. Mounting of mouth parts of honey bee
- 11. Separation of cocoon from worm castings.

SEMESTER V. OPEN COURSES (FOR OTHER STREAMS)

1. VOCATIONAL ZOOLOGY 72 Hrs

4hrs/Week, Credits 3

Objectives of the Course

- To develop critical thinking skill and research aptitude among students, by introducing the frontier areas of the biological science.
- To emphasize the central role that biological sciences plays in the life of all organisms.
- To introduce the student to some of the present and future applications of bio-sciences

Ornamental fish culture. Methods and techniques involved in the formulation of fish feed. Fish Transportation: Live fish packing and transport, Common diseases of aquarium fishes and their management. Establishment of commercial ornamental fish culture unit,

Activity: field visit to an ornamental fish breeding Centre to understand breeding practices of

Module 3Quail farming (*Coturnix coturnix*)

Introduction, care of quail chicks, care of adult quails, care of breeding quails ,ration for quail, care of hatching eggs, health care, use of quail egg and meat, Sources of quality chicks.

Activity: Visit to a quail farm or viewing a quail documentary to familiarize the quail farming practices

Module 5Vermiculture and composting

Introduction, ecological classification of earth worms, Life history, Species of earth worms used for vermicultre, Preparation of vermibed; Preparation of vermicompost, Preparation of vermiwash, Maintenance and management of vermicomposting unit, Role of vermiculture in solid waste management.

Activity: - Preparation of a vermiculture unit or visit to a vermicomposting unit.

Module 6 Apiculture

 \geq To acquire basic knowledge and skills in aquarium management, Quail farming, vermicomposting and apiculture for self-employment

- \triangleright To learn the different resources available and to develop an attitude towards sustainability
- \geq Give awareness to society about need for waste management and organic farming

Module 1Aquarium management

General introduction to Aquarium, Aims and types of aquarium (material, size and shape), Requirements of an aquarium - filtration of waste, physical, chemical and biological; Setting an aquarium (self-sustainable with biological filters), Major indigenous aquarium fishes of Kerala. Activity: Setting up of a freshwater aquarium and rearing of aquarium fishes

Introduction to ornamental fishes: Present status of ornamental fish culture in India with special

reference to Kerala, Breeding of Gold fish, Fighter, Gourami (Osphroneus), and Guppy (live

bearer). Nutrition and types feed for aquarium fishes, Use of live fish feed organisms in

Module 2Ornamental Fish Culture

various aquarium fishes.

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10 Hrs

12 Hrs

20 Hrs

12 Hrs

18 Hrs

Definition, Uses of bees, species of bees cultured, organization of honey bee colony, bee keeping methods (modern method only) and equipments, management and maintenance of an apiary-growth period, dividing the colony, uniting two colonies, replacing old queen with new queen, honey flow period, Bee pasturage, Death period, Enemies of bees, Bee diseases, uses of honey and wax, Apitherapy, Propolis, Royal jelly, Agencies supporting apiculture. **Activity:** Identify different types of honey bees and rearing equipments

Field visit and report Submission

Field visit and report writing on any two items are taken for internal evaluation, instead of assignment and seminar. Conduct a workshop on various cultural practices and the preparation of byproducts.

References:

Applied Zoology, Study Material Zoological Society of Kerala, CMS College Campus, Kottayam.

Addison Webb (1947), Bee Keeping- for profit and pleasure, Museum Press, agro bios India Ltd.

Alka Prakash (2011), Laboratory Manual of Entomology, New age International, New Delhi. Arumugan N. (2008) Aquaculture, Saras publication.

Biju Kumar A and Harishanker J Alappat (1995) A Complete Guide To Aquarium Keeping. Published by Books For All, New Delhi.

Chauhan, H.V.S. and S. Roy, (2008). Fungal Diseases. In: Poultry Diseases, Diagnosis and Treatment, Chauhan, H.V.S. and S. Roy (Eds.). 3rd Ed., New Age International (P) Ltd., New Delhi

Cowey C. B. Mackie, A.M. and Bell, J. G (1985) Nutrition and feeding in fishes. Academy press.

David Alderton (2008). Encyclopedia of Aquarium and Pond fish. Published by Dorling Kindersley, DK Books.

Dey, V.K. (1997). A Hand Book on Aquafarming- Ornamental fishes. Manual. MPEDA Cochin.

George Cust and & Peter Bird. (1978). Tropical Fresh water Aquaria, Published by Hamlyn London. illustrated by George Thompson.

Harisankar J. Alappat and Bijukumar. A. (2011) Aquarium Fishes. B. R. Publ. Corporation, Delhi.

Herbert R. and Leonard P. Schultz Axelrod (1955) Handbook of Tropical Aquarium Fishes, McGraw-Hill, 1955.
Joy P.J., George Abraham K., Aloysius M. Sebastian and Susan Panicker (Eds) (1998) Animal Diversity, Zoological Society of Kerala, Kottayam
Michael B. New; Alber G.J. Tacon (1994) Farm made aquafeeds FAO fisheries technical paper No.343, Rome, FAO. 1994
Nalina Sundari, M.S and Santhi, R (2006) Entomology. MJP Publishers
NPCS Board of Consultants & Engineers, Chennai.(2015) The complete book on Bee keeping and honey processing, 2nd Edition, NIIR Project consultancy services, 106- E kamala Nagar Delhi – 110007.
Ronald j. Roberts (1978) Fish pathology , Cassel Ltd London .
Vijayakumaran Nair, K, Manju, K.G. and Minimol, K. C.(2015) Applied Zoology, Academia

press, Thiruvananthapuram

OPEN COURSE(FOR OTHER STREAMS)

2. PUBLIC HEALTH AND NUTRITION

72 Hrs

4hrs/Week

Credits 3

Objectives:

- To inculcate a general awareness among the students regarding the real sense of health.
- To understand the role of balanced diet in maintaining health.
- To motivate them to practice yoga and meditation in day-to-day life.

PART I HEALTH, EXERCISE & NUTRITION

Module 1 Definition and Meaning of Health

Dimensions and Determination of Health

Physical Activity and Health benefits

Effect of exercise on body systems – Circulatory, Respiratory, Endocrine, Skeletal and Muscular

Programmes on Community health promotion (Individual, Family and Society) Dangers of alcoholic and drug abuse, medico-legal implications

10 Hrs

Module 2	Nutrition and Health	10 Hrs
	Concept of Food and Nutrition, Balanced diet	
	Vitamins, Malnutrition, Deficiency Disease	
	Determining Caloric intake and expenditure	
	Obesity, causes and preventing measures	
	Role of Diet and Exercise, BMI	
Module 3	Safety Education in Health promotion	8 Hrs
	Principles of Accident prevention	
	Health and Safety in daily life.	
	Health and Safety at work.	
	First aid and emergency care.	
	Common injuries and their management.	
	Modern life style and hypokinetic diseases.	
	Diabetese, Cardiovasculard disorders-Prevention and	
	Management.	
Module 4	Life Skill Education	8 Hrs
	Life skills, emotional adjustment and well being,. Yoga,	Meditation and
	Relaxation, Psychoneuroimmunology	
	PART II PUBLIC HEALTH AND SANITATION	
Module 5	Public health and water quality.	11 Hrs
	Potable water, Health and Water quality	
	Faecal bacteriae and pathogenic microorganisms transmi	tted by water.
	Determination of sanitary quality of drinking water, water	er purification
	techniques	
Module 6	Public health and diseases	15 Hrs
	Water borne dseases-Cholera and Typhoid.Prevention	of Water borne
	diseases.	
	Food borne diseases and Prevention	
	Botulinum, Salmenellosis, Hepatitis A	
	Vector borne diseases & Control measures	
	Chikungunya, Filariasis and Dengu fever	
	Zoonotic disease-Leptospirosis & its control	
	Emerging diseases - Swine flue (H1N1), bird flue (H5N	[1),

SARS, Anthrax

Re-emerging diseases -TB, Malaria

Health Centre visit & Report Presentation

10 Hrs

References:

- Gladys Francis & Mini K.D., (Editors) (2012), Microbiology, Zoological Society of Kerala, Kottayam.
- Greenberg, Jerol S and Dintiman George B (1997) Wellness Creating a life of Health and Fitness, London Allyn and Bacon Inc.
- K Park, (2008) Park's Text Book of Preventive and Social Mediine 18th Edition. Banarasidass Bhenot Publication
- Norman Bezzaant HELP First Aid for everyday emergencies. Jaico Publishing House, Bombay, Delhi
- 11. Tom Sanders and Peter Emery. (2004) Molecular basis of human nutrition: Taylor & Francis Publishers Ane Book
- 12. Pelczar M.J. Jr. E.C.S. Chane & N.R. Krieg, Microbiology (Concept & Applications). 5th edition. Tata McGraw Publishing Company Ltd.

SEMESTER V.

OPEN COURSE (FOR OTHER STREAMS)

3. MAN, NATURE AND SUSTAINABLE DEVELOPMENT

72 Hrs 4Hrs/Week Credits 3

Objectives:

- 8. To understand how Man originated and attained present status
- 9. To learn the basic concepts of Ecosystems and its functioning
- 10. To study the use and abuse of nature by Man
- 11. To learn the different resources available on earth
- 12. To study global environmental problems and its impact on human well being

10 Hrs

10 Hrs

7 Hrs

7 Hrs

14. To familiarize with sustainable development and develop an attitude for sustainability

Module I.Man in Nature

- Introduction Evolution of Man
- Out of Africa and Candelabra Model
- The Fossils and the Molecular Evidences
- Hunter-Gatherer and the Agriculturist
- Speech and Languages
- Cultural Evolution
- Altruism and Morality

Module II. The Biosphere

- Earth-Continents and Continental drift
- Concept of Landscapes and Habitats
- Lithosphere- Forest (Tropical and Temperate)
- Grasslands, Deserts and Montane
 - The Biomes of the World
- Hydrosphere- Oceans, Estuaries
 - Freshwater
 - Water the Elixir of Life
- Atmosphere- Structure and stratification

Module III.Dominance of Man on Earth

- Industrial Revolution Human Population Growth Resource Utilization Environmental Consequences Modern Agriculture and Green Revolution Environmental Impacts
- Imperialism and its Ecological Root

Renewable and Non- renewable Biodiversity

Importance of Biodiversity -the Six E^S Hotspots of Biodiversity

Biotic Richness of India

Monoculture and loss of Genetic Diversity

Extinction Crisis, IUCN and Red Data Book

Module V.Global Environmental Issues Threatening Natural

Resources and Human Life

10 Hrs

Deforestation, Landscape alterations, Soil erosion, Flood and Drought, Desertification, Overexploitation, Pollution (Air, Water and Soil- Pollutants and Consequences only), Acid rain, Ozone depletion, Greenhouse effect and Global Warming (use case studies to illustrate the points) Waste disposal (Biodegradable and Non-degradable eg. Plastic and E- waste), Oil spill Energy - Production, Consumption and its Impact on Environment Quality of the Environment and Human Health

Module VI.Man's Perspective on Nature

10 Hrs

8 Hrs

Eco Spirituality, Eco-theology and Eco-feminism

Community initiatives

Indigenous People's Perspective (tribal and traditional communities)

Native American, Amazonian, Australian Aborigines, Bishnoi Contributions of -John Muir, Aldo Leopold, Thoreau, Rachel Carson Edward Abbey, Arne Ness, Carolyn Merchant, Vandana Shiva

Module VII. Global Strategies for Conservation

UN conference on Man and Environment-1972 UNEP and its Contributions

The World Conservation Strategy-1980

World Commission on Environment and Development

The Earth Summit -1992

The UNFCC and IPCC

Conservation Strategies in India-MoEF

Legal System- Mention Major Conservation Acts

People's Participation in Conservation:

Chipko Movement and Narmada Bachao Andolan,

Silent Valley

Module VIII Sustainable Development

Definition and Concept Principles and Goals Environment versus Development Debate Johannesburg Conference -2002 Strategies for Sustainable development Sustainable Development in the era of Globalization Gandhian Environmentalism Education for Sustainable Development (UNESCO-ESD) Building a Sustainable society Sustainable life styles

References:

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SEMESTER VI.

ZOOLOGY CORE CHOICE BASED COURSES FOR B.Sc. ZOOLOGY PROGRAMME ELECTIVE COURSE I ECOTOURISM & SUSTAINABLE DEVELOPMENT

72 Hrs

4hrs/week Credits 3

Objectives:

- 1. To introduce the concepts, principles and applications of tourism and its sustainability
- 2. To critically analyse the cost and benefits of ecotourism, including related laws and policies, community involvement and future trends
- 3. To develop an appreciation among students with respect to tourism development from the sustainability perspective
- 4. To equip the students with basic knowledge for the emerging ecotourism industry

Module I. Fundamentals of Tourism

Introduction- Tourism, concepts and definitions

History, types, Characteristics

The facilitating sectors

Attractions

Geography, heritage

Wildlife, nature

Quality Control

Module II. Major areas of eco-tourism

Concepts, practices and case studies for each:

Marine tourism

Wildlife tourism

Adventure tourism

Module III. Emerging trends in eco-tourism

Cultural tourism

Pilgrimage tourism

12 Hrs

10 Hrs

10 Hrs

Farm tourism	
Backwater tourism	
Health tourism	
Module IV. Problems and prospects of eco-tourism	10 Hrs
Economics and benefits of ecotourism	
Cultural issues and negative aspects of ecotourism	
Environmental Impacts of Tourism	
Module V. Sustainable tourism	12 Hrs
Quality, Standards	
Systems of sustainable tourism: environmental, sociocultural, Economical	
Environment and conservation: basic principles	
Current practices of eco-conservation in tourism industry	
Sustainable tourism and society	
Community based ecotourism	
Eco-development committee (EDC) of Periyar Tiger Rerserve	
People initiatives	
Module VI. Eco-tourism guides	8 Hrs
Ecotourism guiding and case studies	
Activity	
Field visit to Ecologically relevant places & Report writing	10 Hrs
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Ghimire, K.B. and M. Pimbert. 1997. Social change and conservation: environn	nental
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ELECTIVE COURSE

2. AGRICULTURAL PEST MANAGEMENT 72 Hrs

4 Hrs/week - 3 Credits

Objectives

- 1. To acquire basic skills in the observation and study of nature.
- 2. To impart basic awareness regarding pest problem and crop loss due to their dominance.
- 3. To inculcate interest in adopting biological control strategies for pest control.
- 4. To understand various pests affecting our local crops and select the best method for their control
- 5. To acquire basic knowledge and skills in agriculture management to enable the learner for self-employment.

Module I

Pest and crop loss: Introduction, historical perspective-origin of pest, Evolution of pest. Causes of pest outbreak- biotic, abiotic and genetic factors.Modern agricultural practices and pest problem - high yielding varieties, monoculture, fertilizers, pesticides, irrigation, and cultural practices.

Module II

Pest categories: Types of pests- insect pest and non-insect pest.

Insect pest: insect structure and function-external features (body parts), mouth parts of phytophagous insects, internal anatomy, growth, development, reproduction, life cycle and metamorphosis (one example each from ametabolous, hemimetabolous and holometabolous insect), diapause. types of insect pests-key pests, occasional pests, potential pests.

10 Hrs

I0 Hrs

Non insect pests: General features, different types-Rodents(mention the nature of crop loss by them),Mites-Main types of mites; plant injury caused by mite, millipedes and centipedes, slugs and snails (mention the damage of invasive Giant African Snail).

Activity: Identify a minimum of 5 invasive species (plant / animal) in your locality and make a report on their ecological impact.

Module III

Pest and plants: Plant feeding insects-plant host range, types of injury, relationship of pest injury and yield.

Host plant resistance: Characterization of resistance, mechanism of resistance (antixenosis, antibiosis, tolerance), biophysical, biochemical and genetic bases of resistance.

Module IV

20 Hrs

7 Hrs

Pest control-principles and practices: Types of control-cultural control, biological control, chemical control, integrated pest management, miscellaneous control.

Cultural control: Water management, tillage, sanitation, plant diversity, crop rotation, planting time, harvesting practices etc

Biological control: Parasitoids and predators, control by insect pathogens. Techniques in biological control-conservation, introduction and augmentation. Biopesticides

Chemical control: Origin of chemical control, chemistry, mode of action and nomenclature (organochlorines, organophosphates, carbamates, synthetic pyrethroids, miscellaneous group) of pesticides, pesticide formulations and pesticide appliances (sprayers and dusters). Brief mention of attractants, repellents, chemosterilants and pheromones

Activity 1: Conduct a workshop on preparation of biopesticides of various types suitable for kitchen garden and agricultural fields.

Integrated Pest Management (IPM)

Miscellaneous control: Mechanical (hand picking, exclusion by screens and barriers, trapping, clipping, pruning etc), physical (hot and cold treatment, moisture, light traps etc), sterility principle

Module V

Bionomics and control of major pests of crops and stored grains: Biology, life cycle and nature of damage by different pests of following crops and their control

Pests of paddy: *Leptocorisa acuta, Scirpophaga incertulas*, Spodoptera mauritia, *Orseolia oryzae, Nilaparvata lugens*

Pests of coconut: Oryctes rhinoceros, Rhyncophorus ferrugineus, Opisina arenosella, Aceria guerreronis

Pests of Banana: Cosmopolites sordidus, Pentalonianigronervosa

Pests of vegetables - Brinjal: Leucinodesorbonalis, Euzopheraperticella, Henosepilachnavigintioctopunctata, Urentiushystricellus

Gourds -Bactoceracucurbitae, Anadevidiapeponis, Epilachna spp. Raphidopalpafoveicollis, Baristrichosanthis

Pest of stored grains: Sitophilusoryzae, Corcyra cephalonica Triboliumcastraneum, Trogodermagranarium, Callasobruchuschinensis

Activity 2: Conduct a poster exhibition on various types of pests of paddy, coconut, banana and vegetable varieties of Kerala.

Activity 3: Collect different types of pest of stored grains from the local provision shops or houses and make a taxonomic study and prepare a powerpoint presentation on them.

Activity 4: Visit a minimum of 5 kitchen gardens in the neighborhood and enlist the common traditional pest control measures used in them.

Activity 5: Organise awareness classes on the ill effects of chemical pesticides and manure on human health with the support of local examples.

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ELECTIVE COURSE

3. VECTOR AND VECTOR BORNE DISEASES

72 Hrs

3 Credits

10 Hrs

Objectives:

Introduction: Vector : mechanical and biological vector, Reservoirs, Host-vector relationship, Vectorial capacity, Host Specificity.

Insect vectors: Mosquitoes, flies, fleas, lice, ticks and bugs- General account of ecology morphology and mouth parts

Module I1

Salient features and distribution of mosquito species: Anopheles, Aedes, Culex, and Mansonia.

Module II1

Study of Vector Borne disease[Life cycle and pathology]: Mosquito-borne diseases -Filariasis. Sand fly-borne diseases - Leishmaniasis, Malaria, Dengue, Chikungunya, Phlebotomus fever. Tse- tse fly - sleeping sickness. House fly borne diseases :typhoid fever, cholera, dysentery, anthrax, Myiasis, . Flea-borne diseases - Plague, Typhus fever. Louseborne diseases -Relapsing fever, Trench fever, Vagabond's disease, Phthiriasis.

Module IV

Introduction to Vector control: Aims, objectives and advantages. History and background, recent trends, alternatives to the use of insecticides (chemical & microbial), types of vector control - selective, integrated and comprehensive vector control.

6 Hrs

25 Hrs

13 Hrs

Module I

Control measures of mosquitoes, sand fly, tsetse fly and domestic flies

Module V

Introduction to epidemiology: History, Definition, scope and uses of epidemiology. Epidemiology and public health. Achievements in epidemiology: Smallpox Methyl mercury poisoning Rheumatic fever and rheumatic heart disease Iodine deficiency diseases Tobacco use, asbestos and lung cancer, Hip fractures. HIV/AIDS, SARS.

Field report on two case studies of epidemiology in India. 10 Hrs

References:

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8Hrs

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ELECTIVE COURSE

4. NUTRITION, HEALTH AND LIFESTYLE MANAGEMENT

72 Hrs.

3 Credi

Objectives:

- 1. To provide students with a general concept of health and the parameters that define health and wellness.
- 2. To understand principles of nutrition and its role in health.
- 3. To familiarize the students regarding food safety, food laws & regulations.
- 4. To provide knowledge and understanding regarding life style diseases.

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5. To promote an understanding of the value of good life style practices, physical fitness and healthy food habits for life style disease management.

Module I

Nutrition and health: Nutritional requirements of man, classification of major nutrients

including protein, vitamins and minerals, water, role of fibre, biological value of food components, food groups and sources, balanced diet, RDA, BMI, BMR, Calorie intake and expenditure, Healthy eating pyramid, Nutrition in infancy, preschool, school, adolescent, pregnancy, lactation and old age. Nutrition in diseases and special conditions. Food safety: Nutrition education, food sanitation and hygiene, food adulteration and consumer protection.

Module II

Understanding of health: Define health, basic concepts, dimensions of health, basic
parameters of health care. (Health Parameters: Individual normal standards, devices.1. Blood
pressure, 2. Brain activities and sleep, 3.Focus or attention, 4.Pulse, 5. Body temperature,
6. Daily physical activities, 7. Electrocardiogram (ECG), 8.Cardiac fitness 9. Stress,

10. Haematological parameters, 11. BMI

Module III

Introduction to Life style diseases

Common life style diseases: Alzheimer's disease and other neural disorders, asthma, cancer, cardio vascular diseases - including hypertension, Atherosclerosis and stroke, chronic obstructive pulmonary disease, Diabetes Mellitus or Type 2 Diabetes, kidney disorders and chronic renal failure, constipation, depression, gastro-intestinal disturbances including diarrhoea and peptic ulcer, liver cirrhosis and other liver diseases, obesity, osteoporosis, occupational lifestyle diseases.

Modern lifestyle disorders: sleeping habits, junk food, poor eating habits, anxiety, food poisoning

15 Hrs

18 Hrs

15 Hrs

Module IV

Causes of lifestyle diseases: Defects of modern food habits and unbalanced diet options, food adulteration, environmental pollution, poor life style choices, drug abuse, tobacco smoking, alcohol and drug consumption, lack of adequate exercise, wrong body posture, disturbed biological clock, stressful environmental conditions

Module V

14 Hrs

Prevention and control of life style diseases:

Healthy life style habits and practices, healthy eating habits, exercise and fitness, good sleep patterns, a strict no to alcohol, drugs, and other illegal drugs. Uncontrollable factors like age, gender, heredity and race.

Healthy diet: disease prevention through appropriate diet and nutrition, avoiding foods that are high in fats, salt and refined products. Avoid junk food and replace by natural food/ organic food.

Physical exercise: Moderate exercise for fitness of body, walking, stretching, right postures of sitting & standing, relaxation and cutting down of stress, sports, aerobic exercise and yoga. Health literacy as a public health goal: Awareness programs in schools, colleges and through mass media.

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MODEL III B.Sc INDUSTRIAL MICROBIOLOGY &ZOOLOGY (DOUBLE MAIN)

B.Sc. (INDUSTRIAL MICROBIOLOGY & ZOOLOGY) (DOUBLE MAIN)

MODEL –III DOUBLE MAIN – INDUSTRIAL MICROBIOLOGY.

B.Sc. INDUSTRIAL MICROBIOLOGY & ZOOLOGY

Total Credits 120 Total Instructional Hours 150

a. SCHEME OF DISTRIBUTION OF HOURS, CREDITS & EXAMINATIONS

i. THEORY

Theory Examinations will be conducted by the University at the end of the respective semester in which the course is conducted. Duration 3 Hrs (Internal External ratio =1:4)

	Course			Hrs per	a 114	Marks	ratio
No	Code	Course Title	Course Category	week	Credits	Intl	Extl
1		English- (From Board of studies - English)	Common Course	5	4	1	4
2	ZIICRT01	Fundamentals of microbiology	Core Course 1 : Theory	2	2	1	4
3		Fundamentals of microbiology	Core Course 1 : Practical	2	0	0	0
4	ZI1CRT02	Microbial Diversity	Core Course 2 : Theory	2	2	1	4
5		Microbial Diversity	Core Course 2 : Practical	2	0	0	0
6	ZI1CRT03	Microbial physiology	Core Course 3 : Theory	2	2	1	4
7		Microbial physiology	Core Course 3 : Practical	2	0	0	0
8		Biochemistry-1 (From Board of Studies Biochemistry)	Complementary I : Theory	2	2	1	4
9		Practical	Complementary I : Practical	2	0	0	0
		'Computer Science -I (From Board of Studies	Complementary II : Theory	2	2	1	4

- Computer Science)					
Practical	Complementary II : Practical	2	0	0	0
Total			14		

	Course	Course	Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1		English (From Board of studies – English	Common Course	5	4	1	4
3	ZI2CRT04	Microbial waste management	Core Course 4 : Theory	2	2	1	4
4		Microbial waste management	Core Course 4 : Practical	2	2	1	4
5	ZI2CRT05	Medical microbiology	Core Course 5 : Theory	2	2	1	4
6		Medical microbiology	Core Course 5 : Practical	2	2	1	4
7	ZI2CRT06	Agricultural microbiology	Core Course 6 : Theory	2	2	1	4
8		Agricultural microbiology	Core Course 6 : Practical	2	2	1	4
8		Biochemistry-2 (From Board of Studies - Biochemistry)	Complementary I : Theory	2	2	1	4
9		Practical	Complementary I : Practical	2	2	1	4
		Computer Science – 2 (From Board of Studies – Computer Science)	Complementary II : Theory	2	2	1	4
		Practical	Complementary II : Practical	2	2	1	4
	Total				24		

	Course	¹ OUTSE	Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1	ZI3CRT07	Microbial Genetics and Recombinant DNA Technology	Core Course 7 : Theory	3	3	1	4
2		Microbial Genetics and Recombinant DNA Technology	Core Course 7 : Practical	2	0	0	0
3	ZI3CRT08	Industrial Microbiology	Core Course 8 : Theory	3	3	1	4
4		Industrial Microbiology	Core Course 8 : Practical	2	0	0	0
5	ZI3CRT09	Fermentation Technology	Core Course 9 : Theory	3	3	1	4
6		Fermentation Technology	Core Course 9 : Practical	2	0	0	0
7		Biochemistry-3(From Board Of Studies" Biochemistry)	Complementary I : Theory	3	3	1	4
8		Practical	Complementary I : Practical	2	0	0	0
9		Computer science-3 (From Board Of studies, Computer Science)	Complementary II : Theory	3	3	1	4
10		Practical	Complementary II : Practical	2	0	0	0
	Total				15		

	Course	Course	Course	Hrs per		Mark	s ratio
No	Code	Course Title	Category	week	Credits	Intl	Extl
1	ZI4CRT10	Food microbiology	Core Course 10 : Theory	3	3	1	4
2		Food microbiology	Core Course 10 : Practical	2	2	1	4
3	ZI4CRT11	Diary microbiology	Core Course 11 : Theory	3	3	1	4
4		Diary microbiology	Core Course 11 : Practical	2	2	1	4
5	ZY4CRT04	Research methodology,Biophysics & Biostatistics (Core Course IVof <i>Board of</i> <i>Studies - Zoology</i>)	Core Course 12 : Theory	3	3	1	4
6		Research methodology, Biophysics &Biostatistics	Core Course 12 : Practical	2	2	1	4
7		Biochemistry - 4 (From Board of Studies - Biochemistry)	Complementary I : Theory	3	3	1	4
8		Practical	Complementary I : Practical	2	2	1	4
9		Computer Science - 4 (From Board of Studies – Computer Science)	Complementary II : Theory	3	3	1	4
10		Practical	Complementary II : Practical	2	2	1	4
	Total				25		

	Course		Course	Hrs per week	Credits	Marks ratio	
No	Code	Course Title	Category			Intl	Extl
1	ZY5CRT05	Environmental Biology & Human rights	Core Course 13 : Theory	3	3	1	4
2	ZY5CRPO5	Environmental Biology & Human rights	Core Course 13 : Practical	2	0	0	0
3	ZY5CRT06	Cell Biology & Genetics	Core Course 14 : Theory	3	3	1	4
4	ZY5CRPO6	Cell Biology & Genetics	Core Course 14 : Practical	2	0	0	0
5	ZY5CRT07	Evolution, Ethology & Zoogeography	Core Course 15 : Theory	3	3	1	4
6	ZY5CRPO7	Evolution, Ethology & Zoogeography	Core Course 15 : Practical	2	0	0	0
7	ZY5CRT08	Human Physiology, Biochemistry & Endocrinology	Core Course 16 : Theory	3	3	1	4
8	ZY5CRPO8	Human Physiology, Biochemistry & Endocrinology	Core Course 16 : Practical	2	0	0	0
	ZY5OPT01	1 – Vocational Zoology (Apiculture, Vermiculture, Ornamental fish culture)	Open Courses for other		3	1	
	ZY5OPT02	2 – Public health and Nutrition	streams (Select any one out of	4			4
9	ZY5OPT03	3 – Man, nature & Sustainable Development	three)				
		Project work & Field Visit/Study Tour, Visit to research institutes, Group activity	Project work	1	0		
			25 hrs	15			

NT	Course		Course	Hrs per		Marks ratio	
No	Code	Course Title	Category	week	Credits	Intl	Extl
1	ZY6CRT09	Developmental Biology	Core Course 17 : Theory	3	3	1	4
2	ZY6CRP09	Developmental Biology	Core Course 17 : Practical	2	2	1	4
3	ZY6CRT10	Immunology	Core Course 18 : Theory	3	3	1	4
4	ZY6CRP10	Immunology	Core Course 18 : Practical	2	2	1	4
5	ZY6CRT11	Biotechnology, Bioinformatics and Molecular Biology	Core Course 19 : Theory	3	3	1	4
6	ZY6CRP11	Biotechnology, Bioinformatics and Molecular Biology	Core Course 19 : Practical	4	2	1	4
7	ZY6CRT12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course 20 : Theory	3	3	1	4
8	ZY6CRP12	Occupational Zoology (Aquaculture, Apiculture, Vermiculture & Quail farming)	Core Course 20 : Practical	2	2	1	4
	ZY6CBT01	Elective 1: Ecotourism & Sustainable Development					
	ZY6CBT02	Elective 2: Agricultural pest management	Choice Based Core Elective				
9	ZY6CBT03	Elective 3: Vector & Vector borne Diseases	Courses (Select any one out of	3	2	1	4
	ZY6CBT04	Elective 4: Nutrition, Health & life style management	four)				
10	ZI6PRP01	Project work & Field Visit/Study Tour, Visit to research institutes, Group activity	Project		2		
11	ZI6OJP01	OJ (64 Hrs + 36 Hrs)	OJ		3		
			25 hrs	27			

ii. SCHEME FOR PRACTICAL EXAMINATION FOR BSc (Industrial Microbiology & Zoology) Double Main

University Practical Examinations will be conducted at the end of even semesters (Semester II, IV and VI).

			E	Mark	s-ratio	
Semester	Code	Course	Exam duration	Internal (I)	External (E)	Credits
	Z12CRP01	Practical 1 (Core 1 & 4)	3 hrs	1	4	2
	Z12CRP02	Practical 2 (Core 2 & 5)	3 hrs	1	4	2
II	Z12CRP03	Practical 3 (Core 3 & 6)	3 hrs	1	4	2
		Complimentary 1 Biochemistry	3 hrs	1	4	2
		Complimentary 2 Computer Science	3hrs	1	4	2
	Z14CRP04	Practical 4 (Core 7 & 10)	3 hrs	1	4	2
	Z14CRP05	Practical 5 (Core 8 & 11)	3 hrs	1	4	2
IV	Z14CRP06	Practical 6 (Core 9 & 12)	3 hrs	1	4	2
		Complimentary 1 Biochemistry	3 hrs	1	4	2
		Complimentary 2 Computer Science	3hrs	1	4	2
	ZY6CRP03	Practical 7 (Core 13 & 17)	3 hrs	1	4	2
X7T	ZY6CRP04	Practical 8 (Core 14 & 18)	3 hrs	1	4	2
VI	ZY6CRP05	Practical 9 (Core 15 & 19)	3 hrs	1	4	2
	ZY6CRP06	Practical 10 (Core 16 & 20)	3 hrs	1	4	2

b. SYLLABUS -

THEORY & PRACTICALS

SEMESTER I Z11CRT01 - CORE COURSE 1 FUNDAMENTALS OF MICROBIOLOGY

Objectives

1. To introduce students to the fascinating world of microbes

2. To define the science of microbiology and describe some of the general methods used in the

study and culture of microorganisms

3. To impart awareness on how microbes can be controlled

Module 1

History and Scope of Microbiology

Spontaneous generation theory, Contributions of Leuwenhoek, Louis Pasteur, Robert Koch, Edward Jenner, Joseph Lister, Alexander Fleming, John Tyndall.

Module 2

Morphology and Structure of bacteria.

Size, shape and arrangements of bacteria. Structure and arrangement of bacterial flagella, pili, capsule, structure and composition of Gram positive and Gram negative cell wall. Cytoplasmic membrane, protoplasts, spheroplasts, intracellular membrane systems, mesosomes, cytoplasm, vacuoles, nuclear material, endospores and cysts, cell inclusions

Module 3

Sterilization and disinfection

Sterilization-Principles and methods, physical and chemical methods., Antibiotics classification and mechanism of action. Drug resistance, Antibiotic sensitivity tests..

Module 4

Culture media and methods.

Culture media-Definition Media components : Peptone, yeast extract, beef extract, agar, blood/serum - Types : Selective media, Enriched media, Enrichment media, Indicator media, and Differential media, Transport media, Anaerobic media. Aerobic and Anaerobic culture methods.Culture preservation techniques Stains - Acidic, Basic and neutral stains - Staining techniques- Simple staining, differential staining (Gram stain and acid fast stain), Structural staining (spore, flagella, capsule and granule)

4 Hrs

2 Credits

36 Hrs

8 Hrs

8Hrs

Module 5

Microscopy - principles and application – Bright field, Dark field, Phase contrast, Fluorescence, SEM and TEM, ,

References

Lim, D. 1998. *Microbiology*. 2nd Edition; McGraw-Hill Publication.

Ingraham, J. L. and Ingraham, C. A. 2004. *Introduction to Microbiology: A case history approach*. 3rd Edition. Thomson Brooks/Cole, Pacific Grove, Ca.

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Pelczar, M. J. Jr., Chan, E. C. S. and Krieg, N. R. 1993. *Microbiology*, 5th Edition, Tata MacGraw Hill Press.

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Willey, J. M., Sherwood, L. M. and Woolverton, C. J. 2013. *Prescott's Microbiology*. 8th Edition, McGraw-Hill Higher Education.

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Tortora G. J., Funke B. R. and Case C. L. 2006. *Microbiology: An Introduction*. 8th Edition. Pearson Education Inc.

Russell, A. D., Hugo, W. B., and Ayliffe, G. A. J. 1999. *Principles and practice of disinfection, preservation and sterilization*, 3rdEdition. Blackwell Science, Oxford.

Black, J. G. 2013. *Microbiology: Principles and Explorations*. 6th Edition, John Wiley and Sons, Inc.

SEMESTER I

CORE COURSE 1 FUNDAMENTALS OF MICROBIOLOGY (PRACTICAL)

36 Hrs 0 Credit

- 1. General rules in microbiology laboratory
- 2. Instrumentation
 - a. Microscopy
 - b. Incubator
 - c. Hot air oven
 - d. Autoclave
 - e. Quebec colony counter
 - f. water bath
- 3. Sterilization of glass wares
- 4. Preparation of cotton plug
- 5. Preparation of media

Solid media- Nutrient agar, Mac Conkey agar, Blood agar, Chocolate agar, SDA, PDA (for fungi)

Liquid media- Nutrient broth and Glucose broth

- 6. Isolation methods.
 - a. Serial dilution
 - b. Pour plate
 - c. Spread plate
 - d. Streak plate
 - e. Stab culture
- 7. Antibiotic sensitivity tests.

References

Lammert, J. M. 2006. *Techniques for Microbiology: A Student Handbook*. Benjamin Cummings.

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

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Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2ndEdition, New Age International (P) Limited, New Delhi.

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Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Goldman, E. and Green, L. H. 2008. *Practical Handbook of Microbiology*.2nd Edition. CRP Press.

SEMESTER I

Z11CRT02 - CORE COURSE 2 MICROBIAL DIVERSITY

36 Hrs 2 Credits

Objectives

1. To impart knowledge on major categories of microorganisms and analyze their classification and diversity

2. To describe the differences in prokaryotic and eukaryotic cell morphology and structure

Module 1

Principles of Classification, classification based on morphological characteristics, biochemical characteristics, staining reactions, genetic and molecular characteristics, principles of bacterial taxonomy, Outline classification of Bacteria according to Bergey's manual.

Module II

Archaebacteria and extremophiles, brief account on characteristics.Mycoplasma- general properties, structure, cultural characteristics and classification. Actinomycetes - General characteristics and classification, Rickettsiae -Classification, Morphology and distinguishing characteristics

Module III

Viruses - general properties, Structure and Replication of Bacteriophages, TMV, HIV. Cultivation of Viruses, Virions, Viroids and Prions

Module IV

Fungi -morphological features, classification, reproduction and economic importance, ascomycota, basidiomycota, zygomycota, deuteromycota. Cultivation of fungi, Distinguishing characteristics of Rhizopus, Mucor, Aspergillus, Penicillium and Fusarium. Yeasts - a brief account on Candida and Saccharomyces.

Module V

6 Hrs

8 Hrs

8 Hrs

6 Hrs

Algae- characteristics, morphology and structure. Algal pigments, motility, reproduction, classification and economic importance of algae.Cyanobacteria- distribution, characteristics and classification, Ultra structure of Cyanobacterial cell.

References

Topley, W. W. C., Wilson, G. S., Parker, T. and Collier, L. H. 1990. *Topley and Wilson's Principles of Bacteriology, Virology and Immunology*. 8th Edition. Edward Arnold, London. Black, J. G. 2005. *Microbiology, Principles and exploration*. 6th Edition. John Wiley & Sons. Tortora, G. J., Funke, B. R. and Case, C. L. 2012. *Microbiology: An Introduction*. 11th Edition. Pearson education Pvt. Ltd. Singapore.

Lim, D. V. 2002. Microbiology. Dubuque, IA: Kendall/Hunt.

Willey, J., Sherwood, L. M. and Woolverton, C. J. 2011. Microbial growth. In *Prescott's Microbiology*, 8th Edition. McGraw-Hill Companies Inc.: New York, NY, USA.

Salle, A. J. 1971. *Fundamental Principles of Bacteriology*,7th Edition, Tata MacGraw Hill Publishing Company Ltd.

Pelczar, M. J. Jr., Chan, E. C. S., Krieg, N. R. 1986. *Microbiology. McGraw Hill Book Company*, London.

Stanier, R. Y., Ingraham, J. L., Wheelis, M. L. and Painter, P. R. 2005. *General Microbiology*. 5th Edition. McMillan.

Russell A. D., Hugo W. B. and Ayliffe G. A. J. 1999. *Principles and practice of disinfection, preservation, and sterilization*, 3rd Edition. Blackwell Science, Oxford.

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Madigan, M., Martinko, J., Buckley, D. and Stahl, D. 2014. *Brock Biology of Microorganisms*, 14th Edition. Benjamin Cummings, New York.

CORE COURSE 2 MICROBIAL DIVERSITY (PRACTICALS)

36 Hrs 0 Credit

I. Staining techniques

1. Simple staining

2. Differential staining

- a. Gram's staining,
- b. Acid fast staining

3. Structural staining

- a. Endospore staining
- b.. Capsule staining
- c. Negative staining

4. Fungal staining

Lacto phenol cotton blue mounting

II Bacterial motility analysis

.Hanging drop technique

References

Lammert , J. M. 2006. *Techniques for Microbiology: A Student Handbook*. Benjamin Cummings.

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.

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Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2ndEdition, New Age International (P) Limited, New Delhi.

Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*.MJP Publishers, Chennai.

Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Goldman, E. and Green, L. H. 2008. *Practical Handbook of Microbiology*.2nd Edition. CRP Press.

Cappuccino, J. and Sherman, N. 2013. Microbiology: A Laboratory Manual. 10th Edition.

Benjamin-Cummings Publishing Company, Subs Of Addison Wesley Longman, Inc.

Z11CRT03 - CORE COURSE 3 MICROBIAL PHYSIOLOGY

36 Hrs 2 Credits

Objectives

- To gain a fundamental understanding of cellular composition, membrane transport, energy generation, diversity of metabolic processes, growth and cell death, and techniques used to elucidatephysiological processes.
- 2. To get a better understanding of how microbes grow, how they respond and in turn influence their environments.

Module 1

Microbial Nutrition - Nutritional requirements - C, N, P, S, and minerals, Nutritional classification of bacteria. Uptake of nutrients - passive diffusion, facilitated diffusion, Active transport, Group translocation .

Module 2

Bacterial Growth, Bacterial growth curve and generation time.Continuous culturing of bacteria- chemostat, turbidostat,. Effect of Temperature, pH, Oxygen concentration and Radiation on bacterial growth.Enumeration methods of bacteria- SPC, Direct microscopic count, turbidometric estimation. Reproduction and Growth: Modes of cell division- binary fission, budding, and Spore formation.

Module 3

Photosynthesis -Photosynthetic microorganisms, Photosynthetic apparatus in prokaryotes and eukaryotes, photosynthetic pigments, Mechanism of cyclic and non-cyclic photophosphorylation, Calvin cycle.

8 Hrs

8 Hrs

4 Hrs

Microbial Metabolism- Glycolysis, Krebs's cycle, Pentose Phosphate Pathway, gluconeogenesis, ED pathway. Fermentation -Alcoholic fermentation, Homo and hetro-lacticacid fermentation.

Module 5

6 Hrs

Nitrogen metabolism -Nitrogen Cycle, nitrification, denitrification and ammonification. Transamination and deamination reactions, Nitrogen fixation in symbiotic, associative and free living system, oxygen regulation of nitrogen fixation.

References

Doelle, H. W. 1975. Bacterial Metabolism. 2nd Edition. Academic Press.

Moat, A. G. and Foster, J. W. 1988. *Microbial physiology*. 2nd Edition. Springer Verlag.

White, D. 2000. *Physiology and Biochemistry of Prokaryotes*. 2nd Edition. Oxford University Press, New York.

Caldwell, D. R. 1995. *Microbial physiology and Metabolism*. Wm. C Brown Publishers, England.

Madigan, M. T., Martinko, J. M., Stahl, D. A. and Clark, D. P. 2012. *Brock Biology of Microorganisms*, 13th Edition, Benjamin Cummings, San Francisco.

Lim, D. 1998. *Microbiology*. 2nd Edition; McGraw-Hill Publication.

Ingraham, J. L. and Ingraham, C. A. 2004. *Introduction to Microbiology: A case history approach*. 3rd Edition. Thomson Brooks/Cole, Pacific Grove, Ca.

Madigan, M. T. and Martinko, J. M. 2006. *Brock's Biology of Microorganisms*. 11th Edition. Pearson Education Inc.

Pelczar, M. J. Jr., Chan, E. C. S. and Krieg, N. R. 1993. *Microbiology*, 5th Edition, Tata MacGraw Hill Press.

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Prescott, L. M., Harley, J. P. and Klein, D. A. 2006. *Microbiology*. 6th Edition. Edition, McGraw Hill Higher Education.

Willey, J. M., Sherwood, L. M. and Woolverton, C. J. 2013. *Prescott's Microbiology*. 8th Edition, McGraw-Hill Higher Education.

SEMESTER I

CORE COURSE 3 MICROBIAL PHYSIOLOGY

(PRACTICALS)

36 Hrs

0 Credit

- 1. Effect of pH on the growth of bacteria on solid media
- 2. Effect of salts on the growth of microorganisms.
- 3. Effect of temperature on growth of microorganisms.
- 4. Effects of antibiotics on bacterial growth.
- 5. Measurement of size Micrometer
- 6. Measurement of cell number- Haemocytometer

7. Biochemical test:

- a. IMVIC Test
- b. Triple sugar iron agar test
- c. Urease test
- d. Catalase test
- e.Oxidase
- 8. Determination of growth curve of E.coli

References

- 1. Lammert, J. M. 2006. *Techniques for Microbiology: A Student Handbook*. Benjamin Cummings.
- Aneja, K. R. 2003. Experiments in Microbiology, Plant Pathology and Biotechnology. 4th Edition. New Age International (P) Limited, New Delhi.
- 3. Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.
- Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.
- Kannan, N. 2002. Laboratory manual in general microbiology. 2nd Edition, Panima Publishing Co., New Delhi.

- 6. Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2nd Edition, New Age International (P) Limited, New Delhi.
- 7. Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*. MJP Publishers, Chennai.
- 8. Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.
- 9. Goldman, E. and Green, L. H. 2008. *Practical Handbook of Microbiology*. 2nd Edition. CRP Press.
- Cappuccino, J. and Sherman, N. 2013. *Microbiology: A Laboratory Manual*. 10th Edition. Benjamin-Cummings Publishing Company, Subs Of Addison Wesley Longman, Inc.
- 11. Chakraborty, P. and Chakraborty, G. 2005. *Practical pathology*. Vol. 33. Kolkata: New Central Book Agency (P) Ltd.

SEMESTER II

Z12CRT04 -MICROBIAL WASTE MANAGEMENT

36 Hrs

2 Credits

Module 1

Solid waste- types and sources of solid waste,

Solid waste disposal; sanitary landfills, composting; static pile, aerated pile, and feed reactor, role of microorganisms in composting.

Vermicomposting -biomethanation.

Module 2

Liquid waste- sources of liquid waste, components of industrial waste water, treatment of liquid waste; microbiology of municipal sewage;

Sewage treatment - primary treatment- screening, sedimentation, floatation, coagulation and flocculation, secondary treatment - trickling filter, activated sludge, oxidation pond and tertiary treatments;

Anaerobic sludge digestion process

Module 3

Disinfections- chlorination – methods of chlorination- break point chlorination, super chlorination, chloramines, and chlorine dioxide.UV and Ozone treatment. Disposal of treated sewage

Module 4

Microbiology of water pollution

Microbial indicators of water pollution, BOD, COD, eutrophication.Microbiological water quality standards.Aspects of water pollution- biofilm, bio corrosion, bio augmentation.Bacteriological techniques for the examination of water – total count, most probable number, membrane filter technique.Water borne diseases

4 Hrs.

6 Hrs.

8 Hrs.

8 Hrs.

Module 5

Biodegradation - biodegradation of xenobiotic compounds; stimulating biodegradation, hospital waste management, bioremediation, bio sorption.Bioremediation of hazardous waste, dyes, oil, pesticides;Biodegradation of lignin, cellulose and plastics.

References

Odum, E. P. and Barrett, G. W. 2005. *Fundamentals of Ecology*. 5th Edition. Thomson Brooks/Cole, Belmont, CA.

iel, 1999. *Environment Aspects of Microbiology*. 1st Edition, Bright Sun Publications, Chennai. Rajendran, P. and Gunasekaran, P. 2006. *Microbial Bioremediation*. MJP Publishers, Chennai.

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Eriksson, K-E. L., Blanchettee, R. A. and Ander, P. 1990. *Microbial and Enzymatic Degradation of Wood and Wood components*. Springer.

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SEMESTER 1I Z12CRT04- MICROBIAL WASTE MANAGEMENT PRACTICAL -11

36 Hrs

1 Credit

- 1. Bacterial examination of water by MPN technique and IMVIC test.
- 2. Bacterial examination of water by membrane filter technique
- 3. Analysis of water by standard plate count
- 4. Determination of dissolved oxygen
- 5. Estimation of BOD water, raw / treated sewage
- 6. Estimation of COD from water, raw / sewage
- 7. Determination of total alkalinity of water

References

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

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Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Z12CRT05 - CORE COURSE 5 MEDICIAL MICROBIOLOGY

36 Hrs 2 Credits

Objectives

- 1. To develop a knowledge of medically important microorganisms and their relevance of infectious diseases
- 2.To promote understanding of pathogenesis, diagnosis, prevention and control of medically important microorganisms

Module 1.

Normal microbial flora of Human body, A systematic study of *Staphylococcus aureus*, *Streptococcus pyogenes*, *Escherichia coli*, *Klebsiella pneumonia*, *Pseudomonas aeruginosa*, *Vibriocholerae*.

Module 2

Structure and clinical importance of hepatitis B virus, HIV, influenza virus, polio virus. Etiology, Epidemiology, Symptomology, Pathogenesis, Diagnosis and treatment of Tuberculosis, Syphilis, Actinomycosis.

Module 3

A brief account on air borne diseases, Respiratory tract infections -Infections of the upper and lower respiratory tract

Module 4

Urinary tract infections, Genital tract infections, sexually transmitted diseases, and nosocomial infections

Module 5

Fungal diseases -Superficial and deep mycoses

Protozoan Diseases : Malaria., amoebic dysentery, Sleeping sickness.

9 Hrs

8 Hrs

8 Hrs

6 Hrs

References

Cann, A. J. 2005. Principles of Molecular Virology. 4th Edition. Elsevier Academic Press.

Pichare, A. P. and Nagoba, B. S. 2013. *Medical Microbiology: Prep Manual for Undergraduates*. Elsevier India Pvt. Ltd.

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Greenwood, D., Slack, R. C. B., Peutherer, J. F. and Barer, M. R. 2007. *Medical Microbiology: A Guide to Microbial Infections: Pathogenesis, Immunity, Laboratory Diagnosis and Control.* 17th Edition. Elsevier Health Sciences UK.

Topley, W. W. C., Wilson, G. S., Parker, M. T. and Collier, L. H. 1990. *Topley and Wilson's Principles of Bacteriology, Virology and Immunology*. 8th Edition. London: Edward Arnold.

Zinsser, H. and Joklik, W. K. 1992. *Zinsser microbiology*. 20th Edition. Norwalk, CT: Appleton & Lange.

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Emmons, C. W., Binford, C. H., Utz, J. P., Kwon-Chung, K. J. 1977. *Medical Mycology*. 3rd Edition. Philadelphia, Lea & Febiger.

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Z12CRP02 - CORE COURSE 5 MEDICAL MICROBIOLOGY

(PRACTICALS)

36 Hrs

2 Credits

- 1. Microbiology of laboratory safety, General concept for specimen collection, handling.
- 2. Study of the morphology, staining characters, cultural characters and identification of *Staphylococci, Streptococci, E.coli, Klebsiella, Pseudomonas* and *Vibrio*,
- 3. Test for hemolytic property of bacteria.
- 4. Isolation and identification of bacteria from clinical samples
- 5. Culture methods for isolation and identification of fungi- KOH mount preparation, Lacto phenol cotton blue staining, Slide culture technique etc.
- 6. Study of normal microbial flora of human being

References

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Rowland, S. S., Walsh, S. R., Teel, L. D. and Carnahan, A. M. 1994Pathogenic and Clinical Microbiology: A Laboratory Manual. Lippincott Williams and Wilkins.

Gradwohl, R. B. H., Sonnenwirth, A. C. and Jarett, L. 1980. *Gradwohl's clinical laboratory methods and diagnosis*. 8th Edition. Mosby, London.

Cappuccino, J. G. and Sherman, N. 2008. *Microbiology: A Laboratory Manual*. 9th Edition. Pearson/Benjamin Cummings.

Prince, C. P. 2009. *Practical Manual of Medical Microbiology*. Jaypee Brothers Medical Publishers (p) Ltd., New Delhi.

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Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2nd Edition, New Age International (P) Limited, New Delhi.

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Chakraborty, P. and Chakraborty, G. 2005. *Practical pathology*. Vol. 33. Kolkata: New Central Book Agency (P) Ltd.

Z12CRT06 - CORE COURSE 6 AGRICULTURAL MICROBIOLOGY

Objectives

1. To study the various interactions that exists among microbes and also between microbes and plants

2. To describe the roles and importance of microbes in various areas of agriculture

Module 1

Distribution of microorganisms in soil. Soil microorganisms and microbial interactions mutualism, synergism (protocooperation), commensalisms.Amensalism,competition, parasitism, predation, neutralism. Role of microorganisms in cycling of elements- nitrogen, carbon, sulfur, phosphorus cycles

Module 2

Plant – microbe Interaction -Microorganisms of rhizosphere, rhizoplane, phylloplane and myccorhizae- ectomycorrhizae, endomycorrhizae and vesicular arbuscular myccorrhizae Biological nitrogen fixation- Biochemistry and physiology of nitrogen fixation, *nif* genes.

Module 3

Microbial diseases of plants

Bacterial diseases: Bacterial leaf blight of rice, Citrus canker Fungal diseases: Root rot of pepper, Downy mildew of grapes, and Tikka disease of groundnut. Mycoplasmal diseases - Sandal spike, Grassy shoot disease of sugar cane Viral Disease – TMV

Module 4

Bio fertilizers: Types and importance. Production and quality control: *Rhizobium*, *Azotobacter*, *Azospirillum*, Cyanobacteria, mycorrhizae: vesicular arbuscular mycorrhizae. phosphate solubilizing bacteria.

12 Hrs

36 Hrs

Credits 2

5 Hrs

8 Hrs

Module 5

BioPesticides: bacterial, viral and fungal pesticides.Biological control of plant diseases.

References

Agrios, G. 2005. Plant Pathology. 5th Edition, Academic Press.

Hull, R. 2002. Matthew's Plant Virology. 4th Edition, Academic Press.

Atlas, R. N. and Bartha, R. 1998. *Microbial Ecology: Fundamentals & Applications*.4thEdition. Benjamin & Cummings Science Publishing, California.

Subba Rao, M. S. 1995. *Soil microorganisms and plant growth*. Oxford & IBH Publishing Co. Pvt. Ltd. New Delhi.

Bagyaraj, D. J. and Rangaswami, G. 2005. *Agricultural microbiology*. 2ndEdition, Prentice Hall of India.

Mitchell, R. 1974. Introduction to environmental microbiology. Prentice-Hall, Englewood Cliffs, N. J.

Campbell, R. E. 1983. *Microbial ecology*. 2nd Edition, Blackwell Scientific Publications, Oxford; Boston.

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Ahmad, I., Ahmad, F. and Pichtel, J. 2011. *Microbes and Microbial Technology: Agricultural and Environmental Applications*. Springer, New York.

Dart, R. K. 1980 *Microbiological aspects of pollution control*. 2nd Edition. Elsevier Scientific, Amsterdam.

Jan Dirk van Elsas. 1997. Modern Soil Microbiology. Taylor and Francis.

Rajvaidya, N. and Markandey, D. K. 2006. *Agricultural Applications of Microbiology*. APH Publishers.

Z12CRP03 - CORE COURSE 6 AGRICULTURAL MICROBIOLOGY (PRACTICALS)

36 Hrs

2 Credits

- 1. Isolation & enumeration of microorganism from soil.
- 2. Isolation & Cultivation of rhizobium
- 3. Morphological observation of rhizobium from root nodules
- 4. Isolation of microorganism from rhizosphere, and calculation of R: S Ratio
- 5. Study of antagonism between soil microorganisms.
- 6. Study of symptoms of various plant diseases
 - a. Downy mildew of grapes.
 - b. Citrus canker
 - c.Bacterial leaf blight of rice

References

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.

Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2ndEdition, New Age International (P) Limited, New Delhi.

Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*.MJP Publishers, Chennai.

Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Schmidt, E. L.1967. *A Practical Manual of Soil Microbiology Laboratory Methods*. Food and Agriculture Organization of the United Nations.

Rozar, A. 2002. *Practical Methods for Environmental Microbiology and Biotechnology*.Krishna Prakashan Media Ltd., Meerut.

Z13CRT07 - CORE COURSE 7 **MICROBIAL GENETICS AND RECOMBINANT DNA TECHNOLOGY**

54 Hrs **3** Credits

Objectives

1. To learn about bacterial genetics, recombinant DNA, and molecular genetics

2. To understand about the biological processes, using mechanisms of gene regulation as the model.

Module 1

Bacterial chromosome, DNA replication in prokaryote. Meselson and Stahl experiment, modes of replication- rolling circle model.

Module 2

Module 3

Module 5

Mutagenesis, Spontaneous and induced mutatagenesis, transition, transversion, silent, missense, non-sense, neutral, frame shift and conditional mutations. Forward and reverse mutations.

Recombination methods in bacteria: transformation, transduction and conjugation

Module 4.

Plasmids, plasmid as cloning vector, brief account on pBR 322, pUC 8, phage vectors- M13, lamda and cosmids

Basic steps involved in Recombinant DNA Technology, Isolation of DNA, isolation of vector, enzymes in recombinant DNA technology- type II restriction endonucleases, ligases, S1

12 Hrs

8 Hrs

8Hrs

8 Hrs

nuclease, alkaline phosphatase, terminal transferase, DNA polymerase I, reverse transcriptase. Production of Recombinant DNA, Transformation,

Module 6

10 Hrs

Molecular biological techniques, polymerase chain reaction and types, DNA Sequencing- Maxam and Gilbert method, Sangers chain termination method Blotting techniques- southern, northern, and western blotting

References:

Primrose, S., Twyman, R. and Old, B. 2001. *Principles of Gene Manipulation*, 6thEdition, Blackwell Science Ltd.

Chakravarty, A. K. 2013. Introduction to Biotechnology. OUP India.

Chaudhuri, K. 2012. Microbial Genetics. The Energy and Resources Institute, TERI.

Sridhar, S. 2005. Genetics and Microbial Biotechnology.Dominant Publishers and Distributors.

Nicholl, D. S. T. 1994. An Introduction to Genetic Engineering. Cambridge University Press.

Old, R. W. and Primrose, S. B. 2008. *Principles of Gene manipulation*, 4th Edition, Blackwell scientific publications, London.

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Watson, J. D., Gilman, M., Witkowski, J. and Zoller, M. 1992. *Recombinant DNA*. 2nd Edition, Scientific American Books.

Lewin, B. 2000. Genes VIII. Oxford University Press. Oxford.

Balasubramanian, D., Bryce, C., Dharmalingam, K., Green, J. and Jayaraman, K. 1996. *Concepts in Biotechnology*. University Press, India.

Trevan, M. D., Boffey, S., Coulding K. H. and Standury, P. 1990. *Biotechnology. The basic principles*. Tata MC Graw Hill Edition.

Freifelder, S. 1987. Microbial Genetics. Jones and Bartlett, Boston.

Klug, W. S. and Cummings, M. R. 1996. *Essentials of Genetics*. Mentics Hail. New Jersey. Gardner, E. J., Simmons, M. J. and Snustard, D. P. 1991. *Principles of Genetics*. 8th Edition. John Wiley and Sons, NY.

Glazer, A. N. and Nikaido, H. 2007. *Microbial Biotechnology: Fundamentals of Applied Microbiology*. 2nd Edition. Cambridge University Press.

CORE COURSE7 MICROBIAL GENETICS AND RECOMBINANT DNA TECHNOLOGY (PRACTICALS)

36 Hrs 0 Credit

- 1. Isolation of chromosomal DNA from E.coli
- 2. Bacterial transformation
- 3. Immobilization of yeast cells by sodium alginate method
- 4. Polymerase chain reaction
- 5. Blotting techniques
- 6. Western blotting(Only demonstration)

References

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.

Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2nd Edition, New Age International (P) Limited, New Delhi.

Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*. MJP Publishers, Chennai.

Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Janarthanan, S.and Vincent, S. 2007. Practical Biotechnology: Methods and Protocols. Orient BlackSwan/Universities Press.

Jane, M., Valan Arasu, M. and Duraipandiyan, V. 2012. Practical Handbook in Microbial Biotechnology. Lambert Academic Publishing.

Z13CRT08 - CORE COURSE 8 **INDUSTRIAL MICROBIOLOGY**

54 Hrs **3** Credits

Objectives

- 1. To study the sources of microorganisms for use in industrial microbiology and biotechnology
- 2. Discuss the genetic manipulation of microorganism to construct strains that better meet the needs of an industrial or biotechnological process
- 3. Discuss the preservation of microorganisms
- 4. Describe the design or manipulation of environments in which desired processes will be carried out
- 5. Discuss the management of growth characteristics to produce the desired product

Module 1

History, development and scope of industrial microbiology, Industrially important microbes, Strain improvement and selection, Screening of industrially important microorganisms-Primary and Secondary Screening. Strain development.

Module 2

Preservation and storage of microorganisms.Development of inoculum for industrial fermentation. Fermentation media, formulation of media, saccharine materials, starchy materials, cellulosic materials, nitrogenous materials, enhancers and precursors, antifoams.

Industrial sterilization- batch and continuous sterilization.

Module 3

Fermenter, Structure of a typical Batch fermenter, Parts, function of each part, probes, valves, agitators, aerators, baffles, Types of bioreactors:, Types of fermenters: Batch Fermenter, Continuous Stirred Tank Fermenter, Fluidized Bed Fermenter, Solid State Fermenter, Air Lift Fermenter, Tubular Fermenter.

Module 4

10 Hrs

14 Hrs

Fermentation process: Surface, Submerged and Continuous fermentation .Computer control of fermentation process.

Module 5

12 Hrs

Downstream Processing, Intra cellular and extra cellular product recovery.Physical and chemical methods.Cell disruption- methods, solvent extraction, and purification. Product recovery. Drying, Packing and labeling. Market Potential, Good Manufacturing Practices.

References

- 1. Whitaker, A., Stanbury, P. F. and Hall, S. J. 2009. *Principles of Fermentation Techniques*. Elsevier.
- Demain, A. L. and Solomon, N. A. 1986. *Manual of Industrial Microbiology*. Oxford University Press, Oxford.
- Waites, M. J., Morgan, N. L., Rockey, J. S. and Higton, G. 2001. Industrial Microbiology: An Introduction. Blackwell Science Ltd.
- Prescott, S. C., Dunn, C. G., and Reed, G. 1982. Prescott and Dunn's Industrial Microbiology, 4th Edition. AVI Pub. Co., Westport, Conn.
- 5. Waites, M. J. 2001. Industrial Microbiology. Blackwell Science, Oxford.
- 6. McNeil, B. and Harvey, L. M. 1990. *Fermentation: A Practical Approach*.Oxford; New York: IRL Press.
- 7. Enfors, S. O. and Häggström, L. 2000. *Bioprocess Technology: Fundamentals and Applications*. Royal Institute of technology, Stockholm, Sweden.
- 8. Crueger, W., Crueger, A., and Brock, T. D. 1990. *Biotechnology: A Textbook of Industrial Microbiology*. Sinauer Associates.
- 9. Casida, L. E. 1968. Industrial Microbiology. Wiley, New York; London.
- 10. Okafor, N. 2007. Modern Industrial Microbiology and Biotechnology. CRC Press.

SEMESTER III

CORE COURSE 8 INDUSTRIAL MICROBIOLOGY (PRACTICALS)

36 Hrs 0 Credit

- 1. Crowded plate technique for screening microbial production of enzymes and antibiotics
- 2. Solid state fermentation
- 3. Submerged fermentation

4. Preservation techniques:

- a. Serial sub culturing
- b. Over laying with mineral oil
- c. Lyophilization
- d. Liquid nitrogen storage.
- e. Methods for the storage of Fungi
- 5. Estimation of lactic acid from milk.

References

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.

Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2nd Edition, New Age International (P) Limited, New Delhi.

Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*. MJP Publishers, Chennai.

Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Goldman, E. and Green, L. H. 2008. *Practical Handbook of Microbiology*. 2nd Edition. CRP Press.

Baltz, R. H., Davies, J. E. and Demain, A. L. 2012. *Manual of Industrial Microbiology and Biotechnology*. 3rd Edition. Washington DC: American Society of Microbiology.

Z13CRT09 - CORE COURSE 9 FERMENTATION TECHNOLOGY

54 Hrs 3 Credits

Objectives

- 1. To study the major products or uses of industrial microbiology and biotechnology
- 2. To understand aout fermentation and subsequent processing approaches available for themanufacture of biological products and the design and operation of these systems.

Module 1

Fermentative production of Pharmaceuticals, production of penicillin and streptomycin. Production of vitamins and growth stimulants, riboflavin, cyanocobalamines and gibberellins

Module 2

Fermentative production of Alcohol, Acetic acid, citric acid, lactic acid, Lysine and glutamic acid, Protease and amylase, Ethanol and Glycerol

Module 3

Production of biofuels – Hydrogen and Methane Production of Beverages- Beer and Wine Production of Single cell Protein- Baker's Yeast and Spirullina

Module 4

Industrial applications of enzymes, Enzyme immobilization- methods and applications .

Module 5

Microbial recovery of metals- bioleaching of copper, gold and uranium Microbially enhanced oil recovery (MEOR)

10 Hrs

ons.

9 Hrs

10Hrs

10 Hrs

References

Whitaker, A., Stanbury, P. F. and Hall, S. J. 2009. *Principles of Fermentation Techniques*. Elsevier.

Demain, A. L. and Solomon, N. A. 1986. *Manual of Industrial Microbiology*. Oxford University Press, Oxford

Prescott, S. C., Dunn, C. G., and Reed, G. 1982. *Prescott and Dunn's Industrial Microbiology*, 4th Edition. AVI Pub. Co., Westport, Conn.

Hui, Y. H., Meunier-Goddik, L., Hansen, A. L., Josephsen, J., Nip, W.-K., Stanfield, P. S. and Toldra, F. 2004. *Handbook of Food and Beverage Fermentation Technology*. New York : Marcel Dekker Incorporated.

Waites, M. J. 2001. Industrial Microbiology. Blackwell Science, Oxford.

McNeil, B. and Harvey, L. M. 1990. *Fermentation: A Practical Approach*.Oxford; New York: IRL Press.

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Srivastava , M. 2008. Fermentation Technology. Alpha Science International.

Enfors, S. O. and Häggström, L. 2000. *Bioprocess Technology: Fundamentals and Applications*. Royal Institute of technology, Stockholm, Sweden.

Crueger, W., Crueger, A., and Brock, T. D. 1990. *Biotechnology: A Textbook of Industrial Microbiology*. Sinauer Associates.

Casida, L. E. 1968. Industrial Microbiology. Wiley, New York; London.

CORE COURSE 9 FERMENTATION TECHNOLOGY (PRACTICALS)

36 Hrs 0 Credit

- 1. Study of alcoholic fermentation of fruit juice by yeast.
- 2. Production of citric acid by Aspergillus niger
- 3. Estimation of citric acid.

References

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.

Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2nd Edition, New Age International (P) Limited, New Delhi.

Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*. MJP Publishers, Chennai.

Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

McNeil, B. and Harvey, L. M. 2008. *Practical Fermentation Technology*. John Wiley & Sons, Ltd, Chichester.

Kulandaivelu, S. and Janarthanan, S. 2012.Practical Manual on FermentationTechnology.International Publishing House Pvt. Limited.

Z14CRT10 - CORE COURSE10 FOOD MICROBIOLOGY

54 Hrs **3** Credits

Objectives

- 1. To study interaction between micro-organisms and food .
- 2. To discusses the factors affecting the presence of micro-organisms in foods and their capacity to survive and grow.
- 3. To study recent developments in procedures used to assay and control the microbiological quality of food.

Module1

A brief account of micro-organisms important in food industry- molds, yeasts and bacteria, Factors affecting microbial growth in food, extrinsic, intrinsic, implicit and processing factors.

Module 2

Principles of food preservation - asepsis, removal of microorganisms, anaerobic conditions, high and low temperatures, drying, radiation. Chemical preservatives -food additives.Principles of canning.

Module 3

A brief account of microbiological basis of spoilage of vegetables, fruits, milk, egg, meat and fish. Microbial role in production of Bread, vinegar, sauerkraut, beer, and wine.Cultivation of mushrooms.

Module 4

Bacteriological examination of milk. Preservation of milk, pasteurization - different methods. Fermented dairy products- cheese, yoghurt, kefir, butter and butter milk.

Module 5

12 Hrs.

12 Hrs.

8 Hrs.

10 Hrs

Food borne Diseases- Food poisoning and food born infections and intoxication. Mycotoxins -Aflatoxins.Hazard Analysis Critical Control Points (HACCP)

References

Casida, L. E. 1968. Industrial Microbiology. Wiley, New York; London.

Doyle, M. P., Beuchat, L. R. and Montville, T. J. 2001. *Food Microbiology: Fundamentals and Frontiers*. 2nd Edition. ASM Press, Washington, D.C.

Frazier, W. C. and Westhoff, D. C. 2004. *Food Microbiology*. Tata McGraw Hills Publishing Company Limited.

Rose, A. H. 1983. Food microbiology. Academic Press, London.

Garbutt, J. H. 1997. Essentials of food microbiology. Arnold, London.

Wood, B. J. B. 1998. *Microbiology of fermented foods*. 2nd Edition. Blackie Academic and Professional, London.

Ayres, J. C., Mundt, J. O. and Sandine, W. E. 1980. *Microbiology of foods*. Freeman, SanFrancisco.

Robinson, R. K. 1990. *Dairy Microbiology*. 2nd Edition. Elsevier Science Pub. Co., London; New York.

Adams, M. R. and Moss, M. O. 2008. Food Microbiology, 3rd Edition. RSC Publishers.

Ray, B. 2003. Fundamentals of Food Microbiology. Boca Raton, FL: CRC Press.

Prescott, S. C., Dunn, C. G. and Reed, G. 1982. *Prescott and Dunn's Industrial Microbiology*. 4th Edition. AVI Pub. Co., Westport, Conn.

Waites, M. J. 2001. Industrial Microbiology. Blackwell Science, Oxford.

McNeil, B. and Harvey, L. M. 1990. *Fermentation: A Practical Approach*.Oxford; New York: IRL Press.

Jay, J. M., Loessner, M. J. and Golden, D. A. 2005. Modern Food Microbiology. Springer Science & Business Media.

SEMESTER 4

Z14CRP04 - CORE COURSE10 FOOD MICROBIOLOGY

(PRACTICALS)

36 Hrs 2 Credits

- 1. Microbiological examination of food
- 2. Isolation of bacteria from idli batter &curd.
- 3. Microbiological examination of soft drinks.
- 4. Standard plate count of milk.
- 5. Determination of quality of milk sample by methylene blue reduction test.
- 6. Detection of number of bacteria in milk by breed count.
- 7. Quality testing of milk by resazurin test.
- 8. Determination of phosphatase activity of milk.
- 9. Detection of mastitis through milk test.
- 10. Cultivation of Edible mushroom (Demo)

References

Aneja, K. R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. 4th Edition. New Age International (P) Limited, New Delhi.

Aneja, K. R. 2001. *Experiments in microbiology, plant pathology, tissue culture and mushroom production technology*. 3rd Edition. New Age International (P) Limited.

Dubey, R. C. and Maheswari, D. K. 2002. *Practical Microbiology*. 2nd Edition, S. Chand & Co., New Delhi.

Kannan, N. 2002. *Laboratory manual in general microbiology*. 2nd Edition, Panima Publishing Co., New Delhi.

Gunasekaran, P. 2002. *Laboratory manual in microbiology*. 2nd Edition, New Age International (P) Limited, New Delhi.

Kalaichelvan, P. T. 2005. *Microbiology and Biotechnology Laboratory manual*. MJP Publishers, Chennai.

Murugalatha, N. et al. 2012. Microbiological techniques. MJP Publishers, Chennai.

Harrigan, W. F. 1998. Laboratory Methods in Food Microbiology. 3rd Edition.Gulf Professional Publishing.

Yousef, A. E. and Carlstrom, C. 2003. Food Microbiology: A Laboratory Manual, John Wiley and Sons.

Roberts, D. and Greenwood, M. 2003. Practical Food Microbiology. 3rd Edition, Blackwell publishers.

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SEMESTER 4

Z14CRT11 - CORE COURSE 11 DAIRY MICROBIOLOGY

1. To provide a scientific background to dairy microbiology by re-examining the basic concepts of general food microbiology and the microbiology of raw milk.

2. The role of dairy starter cultures in manufacturing fermented dairy products, developing novel functional dairy products through the incorporation of probiotic strains

3. Offer a practical approach to the following aspects: well-known and newfound pathogens that are of major concern to the dairy industry.

Module I

Milk -introduction, composition, microorganisms in milk- bacteria, yeast, mold. Nutritive value of milk.Starter cultures and their biochemical activities- Streptococcus thermophiles and Lactobacillus bulgaricus. Starter culture preparation. Dairy processing unit operationsclarification, separation, standardization, toning of milk, homogenization.

Module II

Bacteriological examination of milk.Preservation of milk, pasteurization - different methods and advantages, sterilization, dehydration, bacteriological standards and grading of milk.

Module III

General principles underlying spoilage of milk and milk products, sources for contamination of milk, milk borne diseases, antimicrobial systems in milk.

Module IV

Fermented dairy products- cheese ,cultured buttermilk, bulgarian butter milk, ice cream, lassie, condensed and dry milk products, yoghurt; low lactose milk, kefir and kumiss.

Module V

12 Hrs

10 Hrs

14 Hrs

10 Hrs

8 Hrs

54 Hrs **3** Credits Hygiene in manufacturing milk products, cleaning of dairy equipment's, dairy processing plant sanitation, probiotic role of lactic acid bacteria and fermented milk products, utilization and disposal of dairy byproduct- whey

References

Prajapati, J. B.1995. *Fundamentals of Dairy Microbiology*. Akta Prakashal Nadiad, Gujarat.Robinson, R. K. 1990. *Dairy Microbiology*. Volume II and I. Elsevier AppliedScience, London.

Marth, E. H. and Steele, J. Applied dairy microbiology. 2nd Edition. CRC Press.

Milk and Milk Products - Fourth edition - clarence henry eckles, Tata McGraw Hillpublishing company Limited, New Delhi, 1957

Dey, S. 1994. Outlines of Dairy Technology. Oxford Univ. Press, New Delhi.

Robinson, R. K. 1986. Modern Dairy Technology. (2 vol. set). Elsevier AppliedScience, UK.

Rosenthal, I. 1991. Milk and Milk Products. VCH, New York.

Warner, J. M. 1976. Principles of Dairy Processing. Wiley Eastern Ltd. New Delhi.

Yarpar, W. J. and Hall, C. W. 1975. Dairy Technology and Engineering. AVI, Westport.

Rheinhermer, G. 1986. Aquatic Microbiology. John Wiley and Sons, NY.

Robinson, R. K. 1981. *Dairy Microbiology: The microbiology of milk products*. Applied Science Publishers.

Law,B. A. 1997. Microbiology and Biochemistry of Cheese and Fermented Milk. 2nd edition. Blackie Academic & Professional, London.

SEMESTER 4

Z14CRP05 CORE COURSE 11 DAIRY MICROBIOLOGY (PRACTICALS)

36 Hrs 2 Credits

- 1. Qualitative analysis of milk by standard plate count method .
- 2. Isolation of coliforms from milk.
- 3. Detection of number of bacteria in milk by breed count.
- 4. Quality testing of milk by rezazurin test
- 5. Methylene blue reduction test for microbial contamination of milk.
- 6. Determination of phosphatase activity of milk
- 7. Detection of mastitis through milk test.
- 8. Isolation of Lactobacilli and Staphylococcus from curd

SEMESTER IV

CORE COURSE 12

RESEARCH METHODOLOGY, BIOPHYSICS AND BIOSTATISTICS

54 Hrs

3 Credits

Objectives

- 1. To familiarise the learner the basic concept of scientific method in research process.
- 2. To have a knowledge on various research designs.
- 3. To develop skill in research communication and scientific documentation.
- 4. To create awareness about the laws and ethical values in biology.
- 5. To equip the students with the basic techniques of animal rearing collection and preservation
- 6. To help the student to apply statistical methods in biological studies.

RESEARCH METHODOLOGY

Module I

Research Process: Scientific method in research (eight steps).

Importance of literature reviewing in defining a problem,

Identifying gap areas from literature review.

Research Communication and scientific documentation: Project proposal writing,

Research report writing, (Structure of a scientific paper), Thesis, dissertation, research article.

Presentation techniques: Oral presentation, Assignment, Seminar, Debate, Workshop,

Colloquium, Conference

Sources of Information: Primary and secondary sources. Library- Books, Journals,

Periodicals, Reviews, Internet.

Search engines Online libraries, e-Books, e-Encyclopedia, Institutional Websites. Plagiarism

Module II

Animal Collection – Tools &techniques

Sampling techniques Quadrate

Line transect

Measurements

Density

Abundance

Frequency

Biodiversity indices - concepts

Simpson index

Collection methods, techniques and equipments

Plankton Insects Fish 12 Hrs

Bird

Preservation techniques - Taxidermy

Rearing techniques

Laboratory and field.

Units of measurements- units, SI system, Equivalent weight, normality, molarity

BIOPHYSICS

Module III

14 Hrs

Basic understanding on principle and uses of the following:

Microscopy

(a) Light microscopy, Bright field (Compound Microscope), Phase contrast, Dark field microscopy, Fluorescence, Polorization microscopy, Video microscopy.

(b) Electron - Scanning (SEM), Transmission (TEM) and STEM

Micrometry – Stage and Eyepiece micrometers Camera Lucida Instrumentation pH Meter Separation Techniques: Centrifuge, Chromatography, Electrophoresis

Analytical techniques: Colorimeter, Spectrophotometer, X-ray crystallography

BIOETHICS

Module IV

5 Hrs

Bioethics : Introduction, Animal rights and animal laws in India, Prevention of cruelty to animals Act 1960, Biodiversity Act 2003.

Concept of 3 R – conservation (Refined- to minimize suffering, Reduced – to minimize animals, Replaced – modern tools and alternate means), Animal use in research and education.

Laboratory animal use, care and welfare, Animal protection initiatives- Animal Welfare Board of India, CPCSEA, ethical commitment. Working with human: Consent, harm, risk and benefits.

BIOSTATISTICS

Module V

Sample & Sampling techniques: Collection of data, classification of data, frequency distribution tables, graphical representation: - Bar diagrams, Histogram, Pie diagram and Frequency curves - Ogives.

Measures of Central Tendency: Mean, Median, Mode (Problem - Direct method only) Measures of dispersion: Range, Quartile Deviation, Mean Deviation, Standard Deviation, Standard error. (Merits & demerits and problems on SD).

Correlation: Definition, Types of correlation.(mention in brief)

Test of Hypothesis and Test of Significance: Basic concept, Levels of significance, test of significance, Procedure for testing hypothesis, types of hypothesis- Null hypothesis and Alternate hypothesis.

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21. Sateesh, M.K. (2008) Bioethics and Biosafety; I.K. International Publishing House .

22. Taylor D.J. Green N.P.O and Stout G.W. (2008). Biological science (3rd edition- R.S. Oper Ed). Cambridge University press.

RESEARCH METHODOLOGY, BIOPHYSICS AND BIOSTATISTICS

(PRACTICAL) 2 credits

PART A. RESEARCH METHODOLOGY

Animal collection Tools, Techniques & Estimation

- 1. Quadrate study
- 2. Transect study
- 3. Sampling Methods
- 4. Species area curve
- 5. Simpson index

PART B - BIOPHYSICS

- 1. Study of simple and compound light microscopes
- 2. Micrometry -calibration and measurement of microscopic objects -low power
- 3. Camera Lucida (draw a few diagrams using Camera Lucida)
- 4. Paper chromatography (demonstration only)
- Instrumentation demonstration (write notes on principle, equipment and its use)
 pH Meter, Colorimeter/ Spectrophotometer, Centrifuge

PART C BIOSTATISTICS

- 1. MS Excel : To create mean and median, Construction of bar diagram, Pie diagram and Line graphs.
- 2. MS Access: To create grade of students
- 3. Internet: Access a web page on any biological topic.
- 4. Frequency distribution of the given samples to find out arithmetic mean, median, mode.
- 5. Range and standard deviation for a biological data
- 6. Correlation using any biological data.
- 7. Graphical representation of data. Construction of bar diagrams, Histograms, Pie diagram and Line graphs.

CORE COURSE 13 ENVIRONMENTAL BIOLOGY AND HUMAN RIGHTS 54 Hrs

Objectives

To instill the basic concepts of Environmental Sciences, Ecosystems, Natural Resources, Population, Environment and Society

To make the students aware of natural resources, their protection, conservation, the factors polluting the environment, their impacts and control measures.

To teach the basic concepts of toxicology, their impact on human health and remedial measures

To create a consciousness regarding Biodiversity, environmental issues & conservation strategies

To develop the real sense of Human rights - its concepts & manifestations

MODULE 1 ECOSYSTEM

12 Hrs

Basic concepts of ecosystem Components of ecosystem: Abiotic (Sunlight, temperature, soil, water, atmosphere) and Biotic components (Producers, consumers, decomposers), Ecological pyramid- number, biomass, energy, **Functions of ecosystem**: Productivity-Food chain-Food web-Energy flow-Laws of Thermodynamics.Types of Ecosystem: Terrestrial-Forest-Grassland-Desert, Aquatic-Marine-Fresh water, Wetland &Biome Concept of limiting factors: Liebig's and Shelford's laws of limiting factors.

Biogeochemical cycles: Concept, gaseous and sedimentary cycles, Carbon cycle, Nitrogen cycle. **Renewable resources** (solar,wind, hydroelectric, biomass and geothermal) **and Non renewable resources** (mineral and metal ore, fossil fuels)

MODULE 2 CONCEPTS OF POPULATION AND COMMUNITY 8 Hrs

Concept of population: Population attributes- Population growth forms, Basic concepts of growth rates, density, natality, mortality, growth curves

Animal interactions: Positive- Commensalism- Mutualism-Protocooperation, Negative-Predation-Parasitism-Competition-Antibiosis

Characteristics of a community: Species diversity- richness, eveness, stratification, dominance, ecological indicators, Ecotone and Edge effect, Keystone species, Concepts of Ecological Niche and Guild, Ecological succession, community evolution- climax.

MODULE 3 BIODIVERSITY AND ENVIRONMENTAL ISSUES 16 Hrs

Introduction to Biodiversity: Types of biodiversity- Alpha, Beta and Gamma diversity. **Concept and importance of Biodiversity**: Levels of Biodiversity-Species diversity, Genetic diversity, Microbial, Ecosystem diversity, India as a mega-diversity nation, Biodiversity hotspots

Global Environmental Issues: Ozone depletion, Greenhouse effect, Global warming, Climate change, Carbon trading, carbon credit; Carbon sequestration, Acid rain, Oil spills, Nuclear accidents, IPCC/UNFCC.

National Environmental issues: Deforestation, forest fire, pollution(air, water, soil, noise thermal, nuclear- brief account only) solid waste management, sewage, drinking water crisis and water logging,

Toxic products and disaster: Types of toxic substances – degradable, non degradable, Impact on human – case studies: Endosulphan tragedy, Bhopal disaster

Flood, drought, cyclone, earthquake and landslide (Management and mitigation)

Local Environmental issues: Landscape alteration, sand mining, quarrying, changing crop pattern, conversion of paddy lands,

Threats to water resources of Kerala: Degrading Mangrove and wetland ecosystems of Kerala, RAMSAR sites, Marine ecosystem crisis- pollution, overfishing etc. Impact of tourism on Environment.

Protected area concept – Sanctuary, National Park, Biosphere reserve, Core Zone, Buffer Zone, Corridor concept. Conservation reserves

Concept of threatened fauna – IUCN categories - extinct, extinct in the wild, critically endangered, endangered, vulnerable, near threatened, least concern and data deficient. Red and Green Data Books.

Man-animal conflict (Tiger, Elephant, Dog, Monkey) - causes and concern

Water conservation- rainwater harvestiong, watershed management

Environment education

Environmental laws (Brief account only): The Water (Prevention and Control of Pollution) Act, 1974, The Air (Prevention and Control of Pollution) Act, 1981, Indian Forests Act (Revised) 1982. The Environment (Protection) Act, 1986, Hazardous Wastes (Management and Handling) Rules, 1989, The Forest (Conservation) Act, 1980, The Wildlife Protection Act, 1972, Biodiversity Act, 2002.

MODULE 5 HUMAN RIGHTS

6 Hrs

Introduction, main concepts associated with Human Rights, Different types of human rights, Manifestations & phenomena, Role of agencies in promoting human rights, Mechanisms for checking violations of human rights, National human right commission, Constitutional provisions related to Human rights.

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Preservation' - PHI Learning, Environmental Law in India: Issues and Responses

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Arun kumar Palai(1999) National Human Rights Commission of India, Atlantic publishers
Sharma P.D. (2005)Environmental biology and Toxicology, Rastogi publication
MeeraAsthana and Astana D.K.1990 Environmental pollution and Toxicology Alka printers.
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Alan Beeby, 2006 Anne – Maria Brennan First Ecology, Ecological principles and
Environmental issues . International students edition Sec. edition Oxford University Press.
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Delhi.

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SEMESTER 5 PRACTICAL

ENVIRONMENTAL BIOLOGY & TOXICOLOGY 36HRS CREDIT 1

1. Estimation of dissolved Oxygen

2. Estimation of carbon di oxide

3. Estimation of soil organic carbon (Demonstration only)

4. Identification of marine/ fresh water planktons

5. Counting of plankton using plankton counting chamber

6. Study of equipments - Sechi disc, Plankton net

7. Study of sandy shore fauna, rocky shore fauna.

8. Study of animal Association

9. Visit to any two important areas of bio diversity: 1. Forest, 2.Sea shore, 3. Mangrove, 4. Wet lands, 5. Bird sanctuary, 6. Wild life sanctuary, 7. Sacred groves

10. Field study (compulsory)

SEMESTER V.

CORE COURSE 14 CELL BIOLOGY AND GENETICS

338

Objectives

1. To understand the structure and function of the cell as the fundamentals for understanding the functioning of all living organisms.

2. To make aware of different cell organelles, their structure and role in living organisms.

3. To develop critical thinking, skill and research aptitudes in basic and applied biology4.To emphasize the central role of genes and their inheritance in the life of all organisms.

CELL BIOLOGY

Module I

Introduction of cell and Diversity of cells: History, Cell theory, Prokaryotes, Eukaryotes, Mycoplasmas, Virus, Virions and Viroids, Prions.

Cell membrane & Permeability: Molecular models of cell membrane (Sandwich model, Unit membrane model, Fluid mosaic model). Cell properties - permeability, Transport [Diffusion, Osmosis, Passive transport, Active transport, bulk transport], Cell coat and Cell recognition.

Module II

Cell Organelles :Structure and functions of following cell organelles: Endoplasmic reticulum -Structure and functions. Ribosomes (Prokaryotic and Eukaryotic) Golgi complex - Structure and functions. Lysosomes - Polymorphism - GERL concept, functions. Mitochondria -Structure and functions. Nucleus: Structure and functions of interphase nucleus, Nuclear membrane, pore complex, structure and functions of nucleolus

Chromosomes – Structure & organization, Heterochromatin, Euchromatin, Nucleosomes, Polytene chromosomes-Balbiani rings, Endomitosis, Lamp brush chromosomes.

Module III

6 Hrs

22 HRS

10 Hrs

Cell Communication: Basic principles of cell communications, Cell signaling(in brief), Types of signaling, Mention signaling molecules (neurotransmitters, hormones, Growth Factors, Cytokines Vitamin A and D derivatives),

Cell Division: Cell cycle - G_1 , S, G_2 and M phases, Mitosis and Meiosis. The difference between Mitosis and Meiosis.

References:

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Biotechnology

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cell structure and function. Saunder's College Publication

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GENETICS

32 Hrs

Module I

10 Hrs

MendelianGenetics: Mendel's experiments- Monohybrid Cross, Dihybrid Cross, Mendel's Laws, Test Cross, Back Cross and Reciprocal Cross. Chromosome Theory of Inheritance

Interaction of genes:Allelic: Incomplete Dominance (Four O Clock Plant).Co- Dominance (Skin colour in Cattle) Lethal Alleles: Dominant lethal gene[Creeper chicken] and recessive lethal gene

[cystic fibrosis].

Non Allelic: Complementary (Flower colour in Sweet Pea), Supplementary (Coat colour in mice), Epistasis - dominant (Plumage in poultry) and recessive (Coat colour in mice). Polygenes (Skin colour inheritance in man), Pleiotropism (Vestigial wing gene in Drosophila).

Multiple alleles – ABO Blood group system, Rh group and its inheritance. Erythroblastosis foetalis.

Module II

12 Hrs

Sex determination: Chromosome theory of sex determination (Autosome and Sex chromosomes), male heterogamy and female heterogamy, (xx-xy, xx-xo, ZZ-ZW, ZZ-ZO), Genic Balance theory of Bridges. Barr bodies, Lyon's hypothesis, Gynandromorphism, sex mosaics, intersex (Drosophila), Hormonal [free martin in calf] and Environmental (Bonelia) influence on Sex determination

Recombination and Linkage: Linkage and recombination of genes based on Morgan's work in Drosophila, Linked genes, Linkage groups, Chromosome theory of Linkage, Types of linkage- complete and incomplete. Recombination, cross over value, chromosome mapping. [Definition]

Sex Linked inheritance : Characteristics of Sex Linked inheritance, X Linked inheritance of man (Hemophilia), Y linked inheritance [Holandric genes], Incompletely Sex Linked genes or pseudoautosomal genes (Bobbed bristles in *Drosophila*), Sex limited genes (Beard in man) and Sex influenced genes (inheritance of baldness in man).

Module III

Mutation: Types of mutations - Somatic, germinal, spontaneous, induced, autosomal and allosomal, chromosomal mutations, structural and numerical changes. Gene mutations. [Addition, Deletion and substitution].

HumanGenetics: Karyotyping, Normal Human chromosome Complement, Pedigree analysis, Aneuploidy and Non- disjunction. Autosomal abnormalities (Down syndrome, Cry du chat syndrome) Sex chromosomal abnormalities (Klinefelters syndrome, Turner's syndrome) Single gene disorder (Brief mention) Autosomal single gene disorder [sickle cell anaemia), Inborn errors of metabolism such as phenylketonuria, alkaptonuria, , Albinism. Multifactorial traits – polygenic disorder- cleft lip and cleft palate.

Genetic Counseling, Eugenics and Euthenics -Brief account only

References:

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5. Singh, B.D. (2006). *Biotechnology*. Kalyani Publishers, New Delhi.

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SEMESTER V

PRACTICAL

CELL BIOLOGY AND GENETICS

36 Hrs 2 Credits

PART A: CELL BIOLOGY

- 1. Squash preparation of onion root tip for mitotic stages
- 2. Mounting of polytene chromosome (Drosophila/Chironomous.) Demonstration
- 3. Tissues (permanent slides of epithelial tissues, striated muscle, smooth muscle, cartilage, bone)
- 4. Identification of cell organelles
- 5. Preparation of temporary whole mount.
- 6. Preparation of permanent whole mount (demonstration)
- 7. Preparation of human blood smear and identification of Leucocytes

PART B : GENETICS

1. Genetic problems on Monohybrid, Dihybrid Crosses and Blood group inheritance

2. Study of normal male and female human karyotype (use photographs or Xerox copies)

- 3. Abnormal human karyotypes Down, Edwards, Klinefelter and Turner syndromes (use photographs or Xerox copies)
- 4. Sexing of Drosophila.
- 5. Study of Barr body in human buccal epithelium

SEMESTER V CORE COURSE - 15: EVOLUTION, ETHOLOGY & ZOOGEOGRAPHY

54 Hrs

Credits 3

Objectives:

- To acquire knowledge about the evolutionary history of earth living and nonliving
- To acquire basic understanding about evolutionary concepts and theories
- To study the distribution of animals on earth, its pattern, evolution and causative factors •
- To impart basic knowledge on animal behavioural patterns and their role •

Prerequisite:

- Basic knowledge on principles of inheritance and variation
- Knowledge on molecular basis of inheritance
- Basic understanding on the mechanism and factors affecting evolution
- Knowledge on origin and evolution of man •

PART I - EVOLUTION

Module I - Origin of life

Theories - Panspermia theory or Cosmozoic theory, Theory of spontaneous generation (Abiogenesis or Autogenesis), Special creation, Biogenesis, Endosymbiosis.

Chemical evolution - Haldane and Oparin theory, Miller-Urey experiment;

Direct evidences of evolution - Recapitulation Theory of Haeckel, Fossilization, Kinds of fossils, fossil dating, Homologous organs and analogous organs.

Module II - Theories of organic evolution

Lamarckism and its Criticism, Weismann's Germplasm theory, Darwinism and its Criticism, Neo-Darwinism, Theory of De Vries,

Population genetics and evolution: Hardy-Weinberg Equilibrium, gene pool, gene frequency. Factors that upset Hardy-Weinberg Equilibrium, Effects of genetic drift on population: Bottleneck effect and founder effect

Module III – Nature of evolution

Species and Speciation: Species concept, subdivisions of species (sub species, sibling species, cline and deme), Speciation: Types of speciation, Phyletic speciation (autogenous and allogenous transformations), True speciation, Instantaneous and gradual speciation, allopatric and sympatric speciation

Isolation: Types of isolating mechanisms-Geographic isolation (mention examples) and Reproductive isolation. Role of isolating mechanisms in evolution

13 Hrs

9 Hrs

8 Hrs

Microevolution, Macroevolution (Adaptive radiation -Darwin finches) Mega evolution, Punctuated equilibrium, Geological time scale, and Mass extinction (brief account only). **Evolution of Horse**

PART II - ETHOLOGY

Module 1V – Introduction 1 Hr

Definition, History and scope of ethology

Module V – Learning, imprinting and behaviour

Types of learning with examples; patterns of behaviors – types of rhythms, navigation, homing instinct, hibernation, aestivation; pheromones- types and their effect on behavior, hormones and their action on behavior (aggressive and parental behavior)

Module VI – Social organization

Social organization in insects (ants) and mammals (monkey), Courtship behaviour and reproductive strategies

PART III - ZOOGEOGRAPHY

Module VII – General Topics

Continental drift theory, Types and means of animal distribution, Factors affecting animal distribution; insular fauna - oceanic islands and continental islands,

Module VIII - Zoogeographical realms

Palaearctic region, Nearctic region, Neotropical region, Ethiopian region, Oriental region, Australian region (brief account with physical features and fauna, Wallace's line, Weber's line, Biogeography of India with special reference to Western Ghats

References:

EVOLUTION

- 1. Barton, N. H., Briggs, D. E. G., Eisen, J. A., Goldstein, D. B. and Patel, N. H. (2007). Evolution.Cold Spring, Harbour Laboratory Press.
- 2. Barnes, C.W. (1988). Earth, Time and Life. John Wiley & Sons, NewYork
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- 5. Campbell, N. A. and Reece J. B. (2011). Biology. IX Edition, Pearson, Benjamin, Cummings.

14 Hrs

9 Hrs

10 Hrs

4 Hrs

4 Hrs

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- 7. Douglas, J. F (1997). Evolutionary Biology. Sinauer Associates.
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- 12. Rob Desalle and Ian Tattersall (2008). Human Origins: What Bones and Genomes Tell Us about Ourselves. Texas A&M University Press, USA.
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ETHOLOGY

- 1. Agarwal. V. K. (2009). Animal Behaviour.S.Chand and Company Pvt. Ltd., New Delhi.
- Bonner, J.T. (1980). The Evolution of Culture in Animals. Princeton University Press.NJ, USA.
- 3. David McFarland. (1999). Animal Behaviour. Pearson Education Ltd. Essex, England.
- 4. Dawkins, M.S. (1995). Unravelling Animal Behaviour. Harlow: Longman.
- 5. Dunbar, R. (1988). Primate Social Systems. Croom Helm, London.
- Gundevia J.S. and Singh H.G. (1996), A Text Book of Animal Behaviour. S. Chand and Company Pvt. Ltd., New Delhi.
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ZOOGEOGRAPHY

- 1. Briggs, J.C. (1996). Global Biogeography. Elsevier Publishers. (Module VI and VII).
- Chandran Subash M.D. (1997). On the ecological history of the Western Ghats.Current Science, Vol.73, No.2.146-155.
- Chundamannil Mammen.1993, History of Forest management in Kerala. Report No.89. Kerala Forest Research Institute, Peechi, India.

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- Mani, M.S. (1974). Ecology and Biogeography of India; The Hague: .Dr. W. Junk b.v. Publishers,
- Nair, C.S. (1991). The Southern Western Ghats: A Biodiversity Conservation Plan. INTACH, New Delhi.
- Ramesh, B.R and R Gurukkal (2007), Forest Landscapes of the Southern Western Ghats, India- Biodiversity, Human Ecology and management Strategies. (French Institute of Pondicherry) India.
- Tiwari, S. (1985), Readings in Indian Zoogeography (vol.1). Today & Tomorrow Printers & Publishers

PRACTICAL

EVOLUTION, ETHOLOGY & ZOOGEOGRAPHY

36 Hrs

Credit 1

- 1. Identification of Zoogeographical realms using map
- 2. Study on endemic species of each realm
- 3. Show the discontinuous distribution of (lung fishes, camel, elephant)
- 4. Providing a map trace the route of HMS Beagle
- Providing a map mark any two continental/oceanic islands.: Greenland, Madagascar, New Zealand, New Guinea, Maldives, Iceland, Hawaii – any two
- 6. Contributions of scientists (showing photos) Any four
- 7. Identification of different stages of horse evolution
- 8. Study on Homology and Analogy
- 9. Study on connecting links (Peripatus, Archaeopteryx, Protopterus, Echidna)
- 10. Pheromone traps
- 11. Skinner box & T Maze
- 12. Experiment to demonstrate phototaxis and chemotaxis using Drosophila/House fly
- 13. Identification of behaviour (Grooming/courtship dance of flamingos/stickle back fish/ Tail wagging dance/ Aggressive behaviour/ Auto/Allo grooming, Flehmen response) showing pictures (Any five)

CORE COURSE 16.

HUMAN PHYSIOLOGY BIOCHEMISTRY, AND ENDOCRINOLOGY

54 Hrs

Credits 3

Objectives:

- 1. This course will provide students with a deep knowledge in biochemistry, physiology and endocrinology.
- 2. Defining and explaining the basic principles of biochemistry useful for biological studies for illustrating different kinds of food, their structure, function and metabolism.
- 3. Explaining various aspects of physiological activities of animals with special reference to humans.
- 4. Students will acquire a broad understanding of the hormonal regulation of physiological processes in invertebrates and vertebrates.
- 5. By the end of the course, students should be familiar with hormonal regulation of physiological systems in several invertebrate and vertebrate systems.
- 6. This also will provide a basic understanding of the experimental methods and designs that can be used for further study and research.
- 7. The achievement of above objectives along with periodic class discussions of current events in science, will benefit students in their further studies in the biological/physiological sciences and health-related fields, and will contribute to the critical societal goal of a scientifically literate citizenry.

HUMAN PHYSIOLOGY 31 Hrs

Module I

Nutrition: Nutritional requirements – carbohydrates, proteins, lipids, minerals (Ca, P, Fe, I), vitamins (sources and deficiency disorders). Importance of dietary fibre and antioxidants.Balanced diet, Recommended Dietary Allowance (RDA). Nutrition during pregnancy and lactation, Infant nutrition, Malnutrition(PEM).

Digestion: Anatomy and histology of digestive glands (liver, pancreas, salivary, gastric and intestinal). Digestion and absorption of carbohydrates, proteins and fats.Nervous and hormonal control of digestion.

Module II

8 Hrs

Respiration: Phases of respiration (external respiration, gas transport and internal respiration). Respiratory pigments: Haemoglobin, Myoglobin (Structure and Function). Transport of respiratory gases - transport of oxygen, oxyhaemoglobin curve, factors affecting oxyhaemoglobin curve, transport of carbon dioxide,(chloride shift). Control of respiration.Respiratory disturbances (Hypoxia, Hypercapnia, Asphyxia).Physiological effect of smoking, carbon monoxide poisoning, Oxygen therapy and artificial respiration.

Circulation: ESR, Haemopoiesis, blood pressure, ECG. Haemostasis (blood coagulation) – clotting factors, intrinsic and extrinsic pathways, anticoagulants and its mechanism of action.Cardiovascular diseases (Jaundice, Atherosclerosis, Myocardial infarction, Thrombus, Stroke).Angiogram and angioplasty.

Module III

Excretion: Histology of Bowman's capsule and tubular part. Urine formation – glomerular filtration, tubular reabsorption, tubular secretion.Urine concentration – counter current mechanism. Acid – base balance, hormonal regulation of kidney function. Renal disorders (kidney stone, acute and chronic renal failure, and dialysis). Homeostasis: Definition, concept and importance in biological system. Thermal regulation and thermal adaptation in homeotherms.

Module IV

Nerve physiology: Ultra structure of neuron. Nerve impulse production (resting membrane potential, action potential), transmission of impulse along the nerve fiber, interneuron (synaptic) transmission, neuromuscular junction and transmission of impulses.Neurotransmitters (acetyl choline, adrenalin, dopamine).EEG. Memory, Neural disorders (brief account on Dyslexia, Parkinson's disease, Alzheimer's disease, Epilepsy).

Muscle physiology: Ultra structure of striated muscle, muscle proteins (myosin, actin, tropomyosin, troponin), Muscle contraction and relaxation-Sliding Filament Theory, cross bridge cycle, biochemical changes and ATP production in muscle, Cori cycle. Kymograph, Simple muscle twitch, muscle fatigue, tetanus, rigor mortis.

5 Hrs

BIOCHEMISTRY

Module V

5 Hrs

15 Hrs

Carbohydrates: Basic structure, biological importance and classification of monosaccharides, oligosaccharides, polysaccharides with examples.

Proteins: Basic structure and classification of amino acids; structure, biological importance and classification of proteins with examples.

Lipids: Structure of fatty acid, saturated and unsaturated fatty acid, biological importance and classification of lipids with examples.

Vitamins and minerals: Major fat soluble and water soluble vitamins. Important minerals and trace elements required for living organisms. Biological importance of vitamins and minerals.

Enzymes: Chemical nature of enzymes, enzyme activation, enzyme inhibition, allosteric enzymes, isoenzymes, co-enzymes. Michaelis–Menten enzyme kinetics.

Module VI

Carbohydrate metabolism: Glycogenesis, Glycogenolysis, Gluconeogenesis, Hexose monophosphate Shunt, Glycolysis, Citric Acid Cycle, Electron Transport Chain and ATP synthesis. Ethanol metabolism.

Protein metabolism: Deamination, Transamination, Transmethylation, Decarboxylation, Ornithine cycle.

Lipid metabolism: Biosynthesis of fatty acids, Beta oxidation, physiologically important compounds synthesized from cholesterol.

ENDOCRINOLOGY

Endocrinology and reproduction Module VII

Endocrine physiology: Hormones – classification and mechanism of hormone action. Major endocrine glands(Histology is not included) their hormones, functions and disorders (hypothalamus, pituitary gland, pineal gland, thyroid gland, parathyroid gland, islets of Langerhans, adrenal gland),. Homeostasis and feedback mechanism.

10 Hrs

8 Hrs 8 Hrs

References:

Biochemistry by U.Satyanarayana and U Chakrapani.Elsevier; 4 edition (2013) **Outlines of Biochemistry by** S.C.Rastogi. CBSPublishers (4)

Medical biochemistry by N.V. Bhagavan, fourth edition Academic Press, 2002

Introduction to Biological chemistryby Awapara J. Prentice Hall.1968 Harper's Illustrated Biochemistry by Harper. 29th edition (Lange basic science.) Biochemistry byLehninger.Palgrave - Macmillan (2007) Text Book of Medical Physiology :Guyton, 13th edition; Elsevier Comparative Animal Physiology :Prosser and Brown, Saunders (1962) Textbook of Physiology : Prof. A K Jain, Publisher: Avichal Publishing Company Textbook of Medical Physiology : Geetha N. Paras Medical Publishers, 3rd edition. Textbook of human physiology :Sarada Subramanyam and K. Madhavankutty, S. Chand & Company Ltd, 2014 Textbook of Endocrinology :Williams, R.H.Elsevier, 12th edition. General and Comparative Endocrinology: Barrington, E.J.W. Oxford University Pres Endocrine Physiology :Martin, C.R.Oxford University Press.

PRACTICAL

HUMAN PHYSIOLOGY, BIOCHEMISTRY AND ENDOCRINOLOGY

36 Hrs Credit1

HUMAN PHYSIOLOGY

- 1). Determination of haemoglobin content of blood
- 2). Total RBC count using Haemocytometer
- 3). Total WBC count using Haemocytometer
- 4). Estimation of microhaematocrit
- 5). Effect of hypertonic, hypotonic and isotonic solutions on the diameter of RBC.
- 6). Instruments: Kymograph, Sphygmomanometer and Stethoscope (principle and use)

7). Measurement of blood pressure using sphygmomanometer(demonstration only)

BIOCHEMISTRY

- 1. Qualitative analysis of protein, glucose, starch and lipids.
- Chromatography Determination of Rf value of amino acids and identification of amino acids (Identify the Amino Acids using different solvent front and solute front)

ENDOCRINOLOGY

- 1. Cockroach Corpora cardiaca & Corpora allata (Demonstration)
- 2. Effect of adrenalin on heart beat of Cockroach (Demonstration)

SEMESTER VI.

CORE COURSE 17 DEVELOPMENTAL BIOLOGY

54 Hrs 3 Credits

Objectives:

- 1. To achieve a basic understanding of the experimental methods and designs that can be used for future studies and research.
- To provide the students with the periodicclass discussions of current events in science which will benefitthem in their future studies in the biological/physiological sciences and health-related fields
- 3. To contribute tocritical societal goal of a scientifically literate citizenry.

Module1

Introduction: Definition, Scope of developmental biology, sub-divisions (descriptive, comparative, experimental and chemical), historical perspectives, basic concepts and theories.

Reproductive Physiology: Gonads- anatomy of testis and ovary, spermatogenesis, oogenesis, gonadal hormones and their functions. Hormonal control of human reproduction - Female reproductive cycles (Estrous cycle, Menstrual cycle). Structure of mammalian sperm and egg, Pregnancy, parturition and lactation.Reproductive health and importance of sex education.

Egg types: Classification of eggs based on the amount, distribution and position of yolk. Mosaic and regulative, cleidoic and noncleidoic eggs.Polarity and symmetry of egg.

Fertilization: Mechanism of fertilization-(Encounter of spermatozoa and Ova, Approach of the Spermatozoon to the Egg, Acrosome Reaction and Contact of Sperm and Ovum, Activation of Ovum, Migration of Pronuclei and Amphimixis,), Significance of fertilization, Polyspermy, Parthenogenesis- Different types and significance.

Module II

14 Hrs

Cleavage: Types, planes and patterns of cleavage, Cell lineage of Planaria. Influence of yolk on cleavage.

Blastulation: Morula, blastula formation, types of blastula with examples.

Fate maps: Concept of fate maps, construction of fate maps (artificial and natural), structure of a typical chordate fate map. Significance of fate map.

Gastrulation: Major events in gastrulation. Morphogenetic cell movements. Influence of yolk on gastrulation. Exogastrulation.Concept of germ layers and derivatives.

Cell differentiation and gene action: Potency of embryonic cells (Totipotency, Pleuripotency, Unipotency of embryonic cells). Determination and differentiation in embryonic development, Gene action during development with reference to Drosophila (maternal effect genes), Zygotic genes.

Module III

20 Hrs

5 Hrs

Embryology of Frog: Gametes, fertilization, cleavage, blastulation, fatemap, gastrulation, neurulation, notogenesis. Differentiation of Mesoderm and Endoderm, Development of eye.Metamorphosis of frog, Hormonal and environmental onrol.

Embryology of chick: Structure of egg, fertilization, cleavage, blastulation, fate map, gastrulation. Development and role of Primitive streak, Salient features of 18hour, 24 hour, 33 hour & 48 hour chick embryo.Extra embryonic membranes in chick.

Human development: Fertilisation, cleavage, blastocyst, implantation, placenta. Gestation, parturition and lactation.Human intervention in reproduction, contraception and birth control. Infertility, Invitro fertilization (test tube baby)

Module IV

Experimental embryology: Spemann's constriction experiments, Organizers and embryonic induction. Embryo transfer technology, cloning, stem cell research. Ethical issues.

Teratology / Dysmorphology, Developmental defects: Teratogenesis, important teratogenic agents.(Radiations, chemicals and drugs, infectious diseases) genetic teratogenesis in human beings,

Developmental defects: Prenatal death (miscarriage and still birth). Intrauterine Growth Retardation (IUGR).

Module V

5 Hrs

General topics: Classification and functions of placenta in mammals. Prenatal diagnosis (Amniocentesis, Chorionic villi sampling, Ultra sound scanning, Foetoscopy, Maternal serum alpha-fetoprotein, Maternal serum beta-HCG).Regeneration in animals. **References:**

Balnisky B.I 1981An Introduction to Embryology, W.B. Saunders and Co.

Majumdar N. N - Vetebrate embryology

Vijayakumarn Nair K.and P. V George. A manual of developmental biology, Continental publications, Trivandrum

Taylor D J, Green NPO & G W Stout.Biological Science, third edition.Cambridge university press.

Dutta, Obstrestics, Church Livingston 17 Ed

Harrison ,Harriosns Book of Internal Medicine Chruch Livingston 17th Ed.

Berril, N.J and Kars G.Developmental biology, Mc Graw Hills

Gibbs. Practical guide to developmental biology.

Gilbert S. F - Developmental biology

PRACTICAL

DEVELOPMENTAL BIOLOGY

36 Hrs

Credit 1

Model/Chart/ Slide may be used

- 1. Embryological studies- Blastula (frog, chick)
- 2. Embryo transfer, cloning, gastrula (frog, chick)
- 3. Amniocentesis

- 4. Embryotransfer technology, cloning
- 5. Study of placenta- pig and man
- 6. 18 hour, 24 hour, 33 hour and 48 hour chick embryo.
- 7. Candling method.
- 8. Vital staining- demonstration.
- 9. Male and female reproductive organs in cockroach
- 10. Calculate the fecundity of fish.
- 11. Calculate the gonado-somatic index of given fish.

SEMESTER VI.

3 Credits

ZY6CRT11 - CORE COURSE 18 IMMUNOLOGY

54 hrs 3 Credits

Objectives

- 1. Microbiology is designed to introduce graduate students to the basics of bacteria, fungus and virus their outline classification.
- 2. To provide pathogens that are the etiological agents of the significant infectious diseases worldwide.
- 3. The course will focus on the basic mechanisms of microbial pathogenesis with emphasis on the host-microbe interactions and the treatments to combat these diseases.
- 4. Immunology is designed to provide a basis of terminology relevant to the basic concepts of immunology. It commences with the important components (cell, tissues; antibodies; immunoglobulins) involved in host defense against infectious agents.

IMMUNOLOGY

Module I

Introduction to Immunology: Innate and acquired immunity, passive (natural and artificial)

and active immunity (natural and artificial). Mechanisms of innate immunity - barriers,

inflammation, phagocytosis.

Lymphoid organs: Primary (thymus, bone marrow) and secondary lymphoid organs (lymph nodes, spleen).

Lymphocytes: T and B cells, Natural killer cells, memory cells, macrophages.

Module 2

Types of antigen, immunoglobulin structure, classes and functions of immunoglobulins. Monoclonal & polyclonal Antibodies Antibody mediated immune response (humoral immunity), Cell mediated immune response.

Antigen – antibody reactions, precipitation test, agglutination test, VDRL WIDAL, ELISA., Complement Fixation Test

Module 4

Module 3

Auto immune diseases: Pernicious Anemia, Rheumatoid Arthritis. Immunodeficiency AIDS. Hyper sensitivity- Type I, (E.g. Anaphylaxix) II(Transfusion reaction), III (Arthus reaction) and IV (Mantaux Test) (in brief).

Module 5

Vaccines

Brief history of vaccination, Types of vaccines (BCG, DPT, Polio vaccine and TAB vaccine, DNA vaccines, Toxoids ,adjuvants

- 1. Ananthanarayan R & Jayaram Paniker C K. (2009) Text Book of Microbiology Orient Longman Private Ltd.
- 2. Kuby J, Kindt T., Goldsby R. and Osborne B. (2007). Kuby immunology
- 3. Microbiology and Immunology, Study Material Series published by Zoological Society of Kerala
- 4. Sharma K. (2005) Manual of Microbiology: Tools and Techniques, Ane books

References

- 1. Colemen: (2002). Fundamentals of Immunology
- 2. Darla J. Wise & Gordon R. Carter: (2004): Immunology A Comprehensive Review Iowa state University Press. A Blackwell science company,
- 3. Hans G. Sch, Legal General Microbiology, Seventh Ed. Cambridge Low Price Ed.
- 4. Helen Hapel, Maused Harney Siraj Misbah and Next Snowden: (2006) Essentials of Clinical Immunology Fifth Ed. Blackwell Publishing Company,
- 5. Heritage, J, E.G.V. Evaus and R.A.Killungten (2007): Introductory Microbiology **Cambridge University Press**

12 Hrs

12 Hrs

8 Hrs

6. Ivan Roitt I (2002) Essentials of Immunology ELBS.

SEMESTER VI

IMMUNOLOGY

(PRACTICAL)

72 hrs 2 Credits

- 1. Determination of ABO blood groups and Rh factor (Antigen antibody Reaction)
- 2. Widal Test(Slide)
- 3.VDRL (Slide)

SEMESTER VI.

CORE COURSE 19

BIOTECHNOLOGY, BIOINFORMATICS & MOLECULAR BIOLOGY,

BIOTECHNOLOGY

Module I

20 Hrs

11 Hrs

9 Hrs

Introduction: Scope, Brief History, Scope and Importance

Tools and Techniques in Biotechnology: Enzymes (restriction endonucleases, ligases, linkers & adapters), Vectors-[Plasmids, Phage vectors, Cosmids, Artificial Chromosomes] Host cells. Basic steps & techniques in rDNA technology

Gene Libraries, Construction of genomic library and cDNA Library. PCR technique and DNA amplification, Brief description of screening methods – Probes, Nucleic Acid hydridization, In situ Hybridization, Fluorescence in situ Hybridization (FISH), Colony hybridization. Methods of transfer of desired gene into target cell.Blotting Techniques- Southern, Northern, Western blotting.DNA Finger printing (DNA Profiling) and its application. Molecular markers - RFLP

Module II

Animal Cell Culture: Brief account on methods, substrates, media and procedure of animal cell culture, Stem Cells, types and potential use, Organismal Cloning- reproductive & therapeutic-brief account only.

Applications of Biotechnology: Applications in Medicine(insulin, growth hormone, gene therapy), Agriculture(GM plants and biopesticides),Environment(bioremediation), Industry (Single Cell Protein) and applications of Fermentation Technology- lactic acid, vitamins, food and beverages.

Potential Hazards of Biotechnological Inventions: Risks related to genetically modified organisms (GMO) and biologically active products, Biological warfare & Biopiracy. Protection of biotechnological inventions.Intellectual Property Rights, Patenting and patent protection.

References

1. Singh B.D Biotechnology 2002. Kalyan Publishers New Delhi.

2. Brown C.H., Campbell I & Priest F, G. 1987. Introduction of Biotechnology (Blackwell scientific publishers Oxford).

3. Colin Ratledge Bijorn Kristiansesn, 2008. Basic Biotechnology 3 rd ed. Cambridge University.

4. Janarathanan S & Vincent S. 2007. Practical Biotechnology, Method of Protocols. University Press.

5. John E. Smith. Biotechnology Cambridge Low priced ed. (Third Ed) 2005 Madingan, Martinko and Parker 2002, Biology of Microorganisms, Brock Eighth Ed. Prentice Hall.

6. Singh B.D. Biotechnolgy 2002, Kalyan Publishers New Delhi.

7. Sudha Gangal 2007. Biotechnology Principles and & practice of Animal Tissue culture, Universities Press.

BIOINFORMATICS

Module III

Introduction: Definition, importance and role of bioinformatics in life sciences. Computational Biology.

Biological databases: Nucleotide sequence databases (NCBI- GENBANK, DDBJ and EMBL). Protein databases - structure and sequence databases (PDB, SWISSPROT and UNIPROT). Introduction to Sequences alignments: Local alignment and Global alignment, Pair wise alignment (BLAST and FASTA] and multiple sequence alignment. Phylogenetic Tree construction and Analysis

Module IV

Molecular visualization software - RASMOL. Basic concepts of Drug discovery pipe line, computer aided drug discovery and its applications. Human Genome Project.

6 Hrs

14 Hrs

MOLECULAR BIOLOGY

Module V

Nature of Genetic Materials: Discovery of DNA as genetic material – Griffith's transformation experiments. Avery Macarty and Macleod, Hershey Chase Experiment of Bacteriophage infection, Prokaryotic genome; Eukaryotic genome.Structure and.types of DNA & RNA.DNA replication. Modern concept of gene (Cistron, muton, recon, viral genes)., Brief account of the following-- Split genes (introns and exons), Junk genes, Pseudogenes, Overlapping genes, Transposons.

Module VI

Gene Expressions: Central Dogma of molecular biology and central dogma reverse, one geneone enzyme hypothesis, One gene-one polypeptide hypothesis Characteristics of genetic code, Contributions of Hargobind Khorana.

Protein synthesis [prokaryotic]: Transcription of mRNA, Reverse transcription, post transcriptional modifications, Translation, Post translational modifications.

Gene regulations: Prokaryotic(inducible & repressible systems) Operon concept -Lac operon and Tryptophan operon, Brief account of Eukaryotic gene regulation.

References

1. Bruce Albert, Bray Dennis, Levis Julian, Raff Martin, Roberts Keith and Watson James (2008). Molecular Biology of the Cell, V Edition, Garland publishing Inc., New York and London.

- De Robertis, E.D.P. and De Robertis, E.M.F. (2006).Cell and Molecular Biology.VIII Edition. Lippincott Williams and Wilkins, Philadelphia.
- 3. Gupta, P. K (2002) Cell and Molecular Biology, (2ed), , Rastogi Publications., Meerut
- 4. James Darnell. (1998) Molecular Biology. Scientific American Books Inc
- Thomas AP(Editor). 2011 Cell & Molecular Biology The Fundamentals. Green leaf publications .TIES Kottayam
- 6. Zoological Society of Kerala Study material. (2011) Cell and Molecular Biology

20 Hrs

8 Hrs

PRACTICAL

BIOTECHNOLOGY, BIOINFORMATICS & MOLECULAR BIOLOGY

BIOTECHNOLOGY

1. Identify and comment on the item provided: (Western blotting / Southern blotting / Northern blotting / PCR)

BIOINFORMATICS

- 1. Download/use print out/pictures of genome sequences of any 2 organisms. Identify and mention the characteristic features of both.
- 2. Download/ use print out/pictures of a protein sequence , identify it & comment on its amino acid composition
- 3. Download / use print out/pictures of a macromolecule. Write a brief note on the

bioinformatics tool used to visualize its structure.

MOLECULAR BIOLOGY

 Identify and comment on its molecular composition / structural orientation / functional significance (Any tissue / Cell organelles/ DNA, DNA replication, RNA different types using models or diagrams) 2. Write down the procedure involved in DNA isolation

V1 SEMESTER. CORE COURSE 20 OCCUPATIONAL ZOOLOGY . (APICULTURE, VERMICULTURE, QUAIL FARMING & AQUACULTURE)

Objectives:

- 1. To equip the students with self employment capabilities.
- 2. To provide scientific knowledge of profitablefarming.
- 3. To make the students aware of cottage industries.

Module 1. APICULTURE

Definition, Different species of honey bees, Organization of honey bee colony, Social life and adaptation of honey bees. Communication among honey bees. Bee keeping methods and equipments, Management and maintenance of an apiary, Growth period, honey flow period and dearth period Division of the colony, uniting two colonies, , replacing old queen with new queen, swarming management, monsoon management. Enemies of bees. Diseases of bees, Bee pasturage. Uses of honey bees, By-products of honey bees, Honey and wax composition. Testing the quality of honey.Extraction of wax, Uses of honey and wax.Royal jelly, Propolis. Apitherapy, Agencies supporting apiculture.

Activity :Visitto an apiculture unit. Field visit and report submission - 10 Hrs Field visit and report submission on any two items are taken for internal evaluation.

MODULE: 2. VERMICULTURE

Introduction, Ecological classification of earth worms. Species of earth worms used for vermicultre, Reproduction & life cycle, Role of earth worm in solid waste management, in agriculture, in medicine etc. Preparation of vermibed, Maintenance & monitoring, Preparation of vermicompost, Preparation of vermiwash.

Activity : Submission of a report after preparing a vermiculture unit or visiting a vermicomposting unit.

MODULE: 3.QUAIL FARMING (Coturnix coturnix)

4 hrs

8 Hrs

Introduction, care of quail chicks, care of adult quails, care of breeding quails, ration for quail, care of hatching eggs, health care, use of quail egg and meat.Sources of quality chicks.

MODULE: 4. AQUACULTURE.

Advantages and salient features of aquaculture, Types of Aquaculture, Biotic and abiotic features of water, Importance of algae in aquaculture, Common cultivable fishes of Kerala, Fish diseases, Composite fish culture, Integrated fish culture, Carp culture, Prawn culture Mussel culture Pearl culture. Processing & Preservation.

Aquarium management - Setting up of an aquarium, Biological filter & Aeration, Breeding of gold fish, gourami (Osphronemus), fighter and Guppy (live bearer). Nutrition and types of feed for aquarium fishes, Establishment of commercial ornamental fish culture unit. Fish Transportation - Live fish packing and transport Common diseases of aquarium fishes and their management. Aquaponics (a brief introduction only).

Activity – Setting up of an Aquarium

Field visit – Visiting an Aquaculture farm

References:

NPCS Board, The complete book on Bee keeping and honey processing, NIIR Project consultancy services, 106E, Kamala nagar, Delhi- 110007. Shukla G.S, & Updhyay V.B, Economic zoology, Rastogi Publ. Meerut. Pradip.V.Jabde, Text book of applied zoology, 2005 Applied Zoology, Study Material Zoological Society of Kerala, CMS college Campus Clive. A Edwards, Norman. Q. & Rhonda. 2011. Vermitechnology: earthworms, organic waste & environmental management. Chauhan, H.V.S. Poultry, Disease, diagnosis and treatment, Wiley eastern Ltd Delhi. Otieno.F.O 2014. Quail farming: markets & market strategies Pillai T.V.R., Aquaculture, principles and practices. Ronald j. Roberts (1978) Fish pathology, Cassel Ltd London. Cowey C. B. et. al. (1985) Nutrition and feeding in fishes, academy press. Farm made aquafeeds. FAO fisheries Technical paper, 343. Harisankar J. Alappat& A. Bijukumar, Aquarium Fishes. B. R. Publ. Corporation, Delhi. MPEDA, A hand Book on AquafarmingOrnamentalfishes, MPEDA, Kochi. Amber Richards. 2014. Aquaponics at home.

Pradip.V.Jabde. 1993. Text book of applied zoology
Venkitaraman, P.R,1983, Text book of Economic zoology(SudharsanaPuubl. Kochi)
Addison Webb, Bee Keepingfor profit and pleasure, Agrobios Ltd.
Edwards.C.A.&Lafty, J.R.1972 Biology of earthworms(Chapman & Hall Led.London)
Applied Zoology, Study Material Zoological Society of Kerala , CMS college Campus
George cust& Peter Bird, Tropical Fresh water Aquaria, Hamlyn London.
Verreth J. Fish larval nutrition, Chapman & Hall Publ.
Bone Packer. 2014. Aquaponic system

PRACTICAL

Occupational Zoology

36 Hrs

Credit 1

- 1. General Identification, Economic importance, Morphology, scientific names and common names of the following
 - a. Economic important and morphology of culturable fishes (Catla, Rohu, Grass carp,

Common carp, Silver carp, Etroplus suratensis, Oreochromis /Tilapia, Mugil

cephalus and Anabas Testudineus)

b. Identification and morphology of ornamental fishes (gold fish, fighter, Gourami,

Angel fish, Guppy

- c. Two species of earthworms used in Vermiculture
- d. Four species of honey bees
- e. Economic importance and morphology of shell fishes (Any three species of prawn,

two marine mussels, two oysters one rock oyster - Crasostria and pearl oyster -

Pinctada fucata and freshwater mussel - Lamellidens marginalis).

- 2. Castes of bees
- 3. Principle & uses of Aquarium filters, Aquarium aerator, Aquarium plants, Oven, Pelletiser, Screw Press, die plate

- 4. Identification and study of fish parasites and diseases (five numbers each) using slides/pictures
- 5. Bee keeping equipments, Beehive, Smoker, honey extractor, Queen Cage,
- 6. Bees wax, Honey, Vermicompost (Identification-Uses)
- 7. Formulation of artificial feed for aquarium fishes demonstration
- 8. Tests for determining the adulteration in honey.
- 9. Mounting of pollen basket
- 10. Mounting of mouth parts of honey bee
- 11. Separation of cocoon from worm castings.

SEMESTER V. OPEN COURSES (FOR OTHER STREAMS)

1. VOCATIONAL ZOOLOGY

72 Hrs

4hrs/Week, Credits 3

Objectives

- To develop critical thinking skill and research aptitude among students, by introducing the frontier areas of the biological science.
- To emphasize the central role that biological sciences plays in the life of all organisms.
- To introduce the student to some of the present and future applications of bio-sciences
- To acquire basic knowledge and skills in aquarium management, Quail farming, vermicomposting and apiculture for self-employment
- To learn the different resources available and to develop an attitude towards sustainability
- Give awareness to society about need for waste management and organic farming

Module 1 Aquarium management

12 Hrs

General introduction to Aquarium, Aims and types of aquarium (material, size and shape), Requirements of an aquarium - filtration of waste, physical, chemical and biological; Setting an aquarium (self-sustainable with biological filters), Major indigenous aquarium fishes of Kerala.

Activity: Setting up of a freshwater aquarium and rearing of aquarium fishes

Module 2 Ornamental Fish Culture

Introduction to ornamental fishes: Present status of ornamental fish culture in India with special reference to Kerala, Breeding of Gold fish, Fighter, Gourami (*Osphroneus*), and Guppy (live bearer). Nutrition and types feed for aquarium fishes,Use of live fish feed organisms in Ornamental fish culture. Methods and techniques involved in the formulation of fish feed. Fish Transportation: Live fish packing and transport, Common diseases of aquarium fishes and their management. Establishment of commercial ornamental fish culture unit,

Activity: field visit to an ornamental fish breeding Centre to understand breeding practices of various aquarium fishes.

Module 3Quail farming (Coturnix coturnix)

Introduction, care of quail chicks, care of adult quails, care of breeding quails ,ration for quail, care of hatching eggs, health care, use of quail egg and meat, Sources of quality chicks.

Activity: Visit to a quail farm or viewing a quail documentary to familiarize the quail farming practices

Module 5Vermiculture and composting

Introduction, ecological classification of earth worms, Life history, Species of earth worms used for vermicultre, Preparation of vermibed; Preparation of vermicompost, Preparation of vermiwash, Maintenance and management of vermicomposting unit, Role of vermiculture in solid waste management.

Activity: - Preparation of a vermiculture unit or visit to a vermicomposting unit.

Module 6Apiculture

Definition, Uses of bees, species of bees cultured, organization of honey bee colony, bee keeping methods (modern method only) and equipments, management and maintenance of an apiary-growth period, dividing the colony, uniting two colonies, replacing old queen with new queen, honey flow period, Bee pasturage, Death period, Enemies of bees, Bee diseases, uses of honey and wax, Apitherapy, Propolis, Royal jelly, Agencies supporting apiculture.

Activity: Identify different types of honey bees and rearing equipments

Field visit and report Submission

Field visit and report writing on any two items are taken for internal evaluation, instead of assignment and seminar. Conduct a workshop on various cultural practices and the preparation of byproducts.

18 Hrs

20 Hrs

10 Hrs

References:

Applied Zoology, Study Material Zoological Society of Kerala, CMS College Campus, Kottayam.

Addison Webb (1947), Bee Keeping- for profit and pleasure, Museum Press, agro bios India Ltd.

Alka Prakash (2011), Laboratory Manual of Entomology, New age International, New Delhi.

Arumugan N. (2008) Aquaculture, Saras publication.

Biju Kumar A and Harishanker J Alappat (1995) A Complete Guide To Aquarium Keeping. Published by Books For All, New Delhi.

Chauhan, H.V.S. and S. Roy, (2008). Fungal Diseases. In: Poultry Diseases, Diagnosis and

Treatment, Chauhan, H.V.S. and S. Roy (Eds.). 3rd Ed., New Age International (P) Ltd., New Delhi

Cowey C. B. Mackie, A.M. and Bell, J. G (1985) Nutrition and feeding in fishes. Academy press.

David Alderton (2008). Encyclopedia of Aquarium and Pond fish. Published by Dorling Kindersley, DK Books.

Dey, V.K. (1997). A Hand Book on Aquafarming- Ornamental fishes. Manual. MPEDA Cochin.

George Cust and & Peter Bird. (1978). Tropical Fresh water Aquaria, Published by Hamlyn London. illustrated by George Thompson.

Harisankar J. Alappat and Bijukumar. A. (2011) Aquarium Fishes. B. R. Publ. Corporation, Delhi.

Herbert R. and Leonard P. Schultz Axelrod (1955) Handbook of Tropical Aquarium Fishes, McGraw-Hill, 1955.

Joy P.J., George Abraham K., Aloysius M. Sebastian and Susan Panicker (Eds) (1998)

Animal Diversity, Zoological Society of Kerala, Kottayam

Michael B. New; Alber G.J. Tacon (1994) Farm made aquafeeds FAO fisheries technical paper No.343, Rome, FAO. 1994

Nalina Sundari, M.S and Santhi, R (2006) Entomology. MJP Publishers

NPCS Board of Consultants & Engineers, Chennai.(2015) The complete book on Bee keeping and honey processing, 2nd Edition, NIIR Project consultancy services, 106- E kamala Nagar Delhi – 110007.

Ronald j. Roberts (1978) Fish pathology, Cassel Ltd London.

Vijayakumaran Nair, K, Manju, K.G. and Minimol, K. C.(2015) Applied Zoology, Academia press, Thiruvananthapuram

OPEN COURSE (FOR OTHER STREAMS)

2. PUBLIC HEALTH AND NUTRITION

72 Hrs

4hrs/Week

Credits 3

Objectives:

- To inculcate a general awareness among the students regarding the real sense of health. •
- To understand the role of balanced diet in maintaining health. •
- To motivate them to practice yoga and meditation in day-to-day life.

PART I HEALTH, EXERCISE & NUTRITION

Module 1	Definition and Meaning of Health	10 Hrs
	Dimensions and Determination of Health	
	Physical Activity and Health benefits	
	Effect of exercise on body systems - Circulatory, Re	espiratory, Endocrine,
	Skeletal and Muscular	
	Programmes on Community health promotion (Individual	, Family and Society)
	Dangers of alcoholic and drug abuse, medico-legal implie	cations
Module 2	Nutrition and Health	10 Hrs
	Concept of Food and Nutrition, Balanced diet	
	Vitamins, Malnutrition, Deficiency Disease	
	Determining Caloric intake and expenditure	
	Obesity, causes and preventing measures	
	Role of Diet and Exercise, BMI	
Module 3	Safety Education in Health promotion	8 Hrs
	Principles of Accident prevention	
	Health and Safety in daily life.	

Health and Safety at work.
First aid and emergency care.
Common injuries and their management.
Modern life style and hypokinetic diseases.
Diabetese, Cardiovasculard disorders-Prevention and
Management.
Life Skill Education

Module 4Life Skill Education8 HrsLife skills, emotional adjustment and well being,Yoga, Meditation andRelaxation, Psychoneuroimmunology

PART II PUBLIC HEALTH AND SANITATION

Module 5	Public health and water quality. 1	1 Hrs		
	Potable water, Health and Water quality			
	Faecal bacteriae and pathogenic microorganisms transmitted by water.			
	Determination of sanitary quality of drinking water, water purification			
	techniques			
Module 6	Public health and diseases	15 Hrs		
	Water borne dseases-Cholera and Typhoid.Prevention of Water	borne		
	diseases.			
	Food borne diseases and Prevention			
	Botulinum, Salmenellosis, Hepatitis A			
	Vector borne diseases & Control measures Chikungunya , Filariasis and Dengu fever			
				Zoonotic disease-Leptospirosis & its control
		Emerging diseases - Swine flue (H1N1), bird flue (H5N1),		
	SARS, Anthrax			
	Re-emerging diseases –TB, Malaria			
	Health Centre visit & Report Presentation	10 Hrs		
	References:			

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SEMESTER V.

OPEN COURSE (FOR OTHER STREAMS) 3. MAN, NATURE AND SUSTAINABLE DEVELOPMENT

72 Hrs 4Hrs/Week Credits 3

Objectives:

- 1. To understand how Man originated and attained present status
- 2. To learn the basic concepts of Ecosystems and its functioning
- 3. To study the use and abuse of nature by Man
- 4. To learn the different resources available on earth
- 5. To study global environmental problems and its impact on human well being
- 6. To appreciate the perspectives of Man on nature and learn the strategies for conservation
- 7. To familiarize with sustainable development and develop an attitude for sustainability

Module I. Man in Nature

Introduction Evolution of Man Out of Africa and Candelabra Model The Fossils and the Molecular Evidences Hunter-Gatherer and the Agriculturist Speech and Languages Cultural Evolution

Altruism and Morality	
Module II.The Biosphere	10 Hrs
Earth-Continents and Continental drift	
Concept of Landscapes and Habitats	
Lithosphere- Forest (Tropical and Temperate)	
Grasslands, Deserts and Montane	
The Biomes of the World	
Hydrosphere- Oceans, Estuaries	
Freshwater	
Water the Elixir of Life	
Atmosphere- Structure and stratification	
Module III.Dominance of Man on Earth	7 Hrs
Industrial Revolution	
Human Population Growth	
Resource Utilization	
Environmental Consequences	
Modern Agriculture and Green Revolution	
Environmental Impacts	
Imperialism and its Ecological Root	
Module IV.Natural Resources	7 Hrs
Renewable and Non- renewable	
Biodiversity	
Importance of Biodiversity -the Six E ^S	
Hotspots of Biodiversity	
Biotic Richness of India	
Monoculture and loss of Genetic Diversity	
Extinction Crisis, IUCN and Red Data Book	
Module V.Global Environmental Issues Threatening Natural	
Resources and Human Life	
	10 Hrs
Deforestation, Landscape alterations, Soil erosion, Flood and Drought, I	Desertificat

tion, Overexploitation, Pollution (Air, Water and Soil- Pollutants and Consequences only),

Acid rain, Ozone depletion, Greenhouse effect and Global Warming (use case studies to illustrate the points) Waste disposal (Biodegradable and Non-degradable eg. Plastic and E- waste), Oil spill Energy - Production, Consumption and its Impact on Environment Quality of the Environment and Human Health

Module VI.Man's Perspective on Nature

10 Hrs

8 Hrs

10 Hrs

Eco Spirituality, Eco-theology and Eco-feminism Community initiatives Indigenous People's Perspective (tribal and traditional communities) Native American, Amazonian, Australian Aborigines, Bishnoi Contributions of -John Muir, Aldo Leopold, Thoreau, Rachel Carson Edward Abbey, Arne Ness, Carolyn Merchant, Vandana Shiva

Module VII. Global Strategies for Conservation

UN conference on Man and Environment-1972 UNEP and its Contributions The World Conservation Strategy-1980 World Commission on Environment and Development The Earth Summit -1992 The UNFCC and IPCC Conservation Strategies in India-MoEF Legal System- Mention Major Conservation Acts People's Participation in Conservation: Chipko Movement and Narmada Bachao Andolan, Silent Valley

Module VIII Sustainable Development

Definition and Concept Principles and Goals Environment versus Development Debate Johannesburg Conference -2002 Strategies for Sustainable development Sustainable Development in the era of Globalization Gandhian Environmentalism Education for Sustainable Development (UNESCO-ESD) Building a Sustainable society Sustainable life styles

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SEMESTER VI.

ZOOLOGY CORE CHOICE BASED COURSES FOR B.Sc. ZOOLOGY PROGRAMME ELECTIVE I ECOTOURISM & SUSTAINABLE DEVELOPMENT

72 Hrs

4hrs/week Credits 3

Objectives:

- 1. To introduce the concepts, principles and applications of tourism and its sustainability
- 2. To critically analyse the cost and benefits of ecotourism, including related laws and policies, community involvement and future trends
- 3. To develop an appreciation among students with respect to tourism development from the sustainability perspective
- 4. To equip the students with basic knowledge for the emerging ecotourism industry

Module I. Fundamentals of Tourism	12 Hrs
Introduction- Tourism, concepts and definitions	
History, types, Characteristics	
The facilitating sectors	
Attractions	
Geography, heritage	
Wildlife, nature	
Quality Control	
Module II. Major areas of eco-tourism	10 Hrs
Concepts, practices and case studies for each:	
Marine tourism	
Wildlife tourism	
Adventure tourism	
Module III. Emerging trends in eco-tourism	10 Hrs
Cultural tourism	
Pilgrimage tourism	
Farm tourism	
Backwater tourism	
Health tourism	
Module IV. Problems and prospects of eco-tourism	10 Hrs
Economics and benefits of ecotourism	
Cultural issues and negative aspects of ecotourism	
Environmental Impacts of Tourism	
Module V. Sustainable tourism	12 Hrs
Quality, Standards	
Systems of sustainable tourism: environmental, sociocultural, Economical	
Environment and conservation: basic principles	

Current practices of eco-conservation in tourism industry				
Sustainable tourism and society				
Community based ecotourism				
Eco-development committee (EDC) of Periyar Tiger Rerserve				
People initiatives				
Module VI. Eco-tourism guides 8 Hrs				
Ecotourism guiding and case studies				
Activity				
Field visit to Ecologically relevant places & Report writing 10 Hrs				
References:				
Bruner, E.M. 2005. Culture on tour: ethnographies of travel. The University of Chicago				
Press.				
Ghimire, K.B. and M. Pimbert. 1997. Social change and conservation: environmental				
politics and impacts of national parks and protected areas. London: Earthscan Publications.				
Karan Singh. 1980. Indian Tourism: Aspects of great adventure. Department of tourism.				
New Delhi.				
Ratandeep Sing. 2003. National Ecotourism and Wildlife tourism: Policies and guidelines.				
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Press.				
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politics and impacts of national parks and protected areas. London: Earthscan Publications.				
Ratandeep Sing. 2003. National Ecotourism and Wildlife tourism: Policies and guidelines.				
Kanishka Publishers, New Delhi				
ELECTIVE COURSE				

2. AGRICULTURAL PEST MANAGEMENT

72 Hrs

4 Hrs/week - 3 Credits

Objectives

1. To acquire basic skills in the observation and study of nature.

2. To impart basic awareness regarding pest problem and crop loss due to their dominance.

- 3. To inculcate interest in adopting biological control strategies for pest control.
- 4. To understand various pests affecting our local crops and select the best method for their control
- 5. To acquire basic knowledge and skills in agriculture management to enable the learner for self-employment.

Module I

Pest and crop loss: Introduction, historical perspective-origin of pest, Evolution of pest. Causes of pest outbreak- biotic, abiotic and genetic factors.Modern agricultural practices and pest problem - high yielding varieties, monoculture, fertilizers, pesticides, irrigation, and cultural practices.

Module II

5 Hrs

I5 Hrs

Pest categories: Types of pests- insect pest and non-insect pest.

Insect pest: insect structure and function-external features (body parts), mouth parts of phytophagous insects, internal anatomy, growth, development, reproduction, life cycle and metamorphosis (one example each from ametabolous, hemimetabolous and holometabolous insect), diapause. types of insect pests-key pests, occasional pests, potential pests.

Non insect pests: General features, different types-Rodents(mention the nature of crop loss by them),Mites-Main types of mites; plant injury caused by mite, millipedes and centipedes, slugs and snails (mention the damage of invasive Giant African Snail).

Activity: Identify a minimum of 5 invasive species (plant / animal) in your locality and make a report on their ecological impact.

Module III

Pest and plants: Plant feeding insects-plant host range, types of injury, relationship of pest injury and yield.

Host plant resistance: Characterization of resistance, mechanism of resistance (antixenosis, antibiosis, tolerance), biophysical, biochemical and genetic bases of resistance.

Module IV

20 Hrs

Pest control-principles and practices: Types of control-cultural control, biological control, chemical control, integrated pest management, miscellaneous control.

Cultural control: Water management, tillage, sanitation, plant diversity, crop rotation, planting time, harvesting practices etc

Biological control: Parasitoids and predators, control by insect pathogens. Techniques in biological control-conservation, introduction and augmentation. Biopesticides

Chemical control: Origin of chemical control, chemistry, mode of action and nomenclature (organochlorines, organophosphates, carbamates, synthetic pyrethroids, miscellaneous group) of pesticides, pesticide formulations and pesticide appliances (sprayers and dusters). Brief mention of attractants, repellents, chemosterilants and pheromones

Activity 1: Conduct a workshop on preparation of biopesticides of various types suitable for kitchen garden and agricultural fields.

Integrated Pest Management (IPM)

Miscellaneous control: Mechanical (hand picking, exclusion by screens and barriers, trapping, clipping, pruning etc), physical (hot and cold treatment, moisture, light traps etc), sterility principle

Module V

25 Hrs

Bionomics and control of major pests of crops and stored grains: Biology, life cycle and nature of damage by different pests of following crops and their control

Pests of paddy: *Leptocorisa acuta, Scirpophaga incertulas,* Spodoptera mauritia, *Orseolia oryzae, Nilaparvata lugens*

Pests of coconut: Oryctes rhinoceros, Rhyncophorus ferrugineus, Opisina arenosella, Aceria guerreronis

Pests of Banana: Cosmopolites sordidus, Pentalonianigronervosa

Pests of vegetables - Brinjal: Leucinodesorbonalis, Euzopheraperticella, Henosepilachnavigintioctopunctata, Urentiushystricellus

Gourds -Bactoceracucurbitae, Anadevidiapeponis, Epilachna spp. Raphidopalpafoveicollis, Baristrichosanthis

Pest of stored grains: Sitophilusoryzae, Corcyra cephalonica Triboliumcastraneum, Trogodermagranarium, Callasobruchuschinensis

Activity 2: Conduct a poster exhibition on various types of pests of paddy, coconut, banana and vegetable varieties of Kerala.

Activity 3: Collect different types of pest of stored grains from the local provision shops or houses and make a taxonomic study and prepare a powerpoint presentation on them.

Activity 4: Visit a minimum of 5 kitchen gardens in the neighborhood and enlist the common traditional pest control measures used in them.

Activity 5: Organise awareness classes on the ill effects of chemical pesticides and manure on human health with the support of local examples.

References

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ELECTIVE COURSE

3. VECTOR AND VECTOR BORNE DISEASES

72 Hrs

3 Credits

Objectives:

Module I

Introduction: Vector : mechanical and biological vector, Reservoirs, Host-vector relationship, Vectorial capacity, Host Specificity.

Insect vectors: Mosquitoes, flies, fleas, lice, ticks and bugs- General account of ecology morphology and mouth parts

Module I1

Salient features and distribution of mosquito species: Anopheles, Aedes, Culex, and Mansonia.

Module II1

Study of Vector Borne disease[Life cycle and pathology]: Mosquito-borne diseases -Malaria, Dengue, Chikungunya, Filariasis. Sand fly-borne diseases - Leishmaniasis, Phlebotomus fever. Tse- tse fly - sleeping sickness. House fly borne diseases :typhoid fever, cholera, dysentery, anthrax, Myiasis, . Flea-borne diseases - Plague, Typhus fever. Louseborne diseases -Relapsing fever, Trench fever, Vagabond's disease, Phthiriasis.

Module IV

Introduction to Vector control: Aims, objectives and advantages. History and background, recent trends, alternatives to the use of insecticides (chemical & microbial), types of vector control - selective, integrated and comprehensive vector control.

Control measures of mosquitoes, sand fly, tsetse fly and domestic flies

Module V

Introduction to epidemiology: History, Definition, scope and uses of epidemiology. Epidemiology and public health. Achievements in epidemiology: Smallpox Methyl mercury poisoning Rheumatic fever and rheumatic heart disease Iodine deficiency diseases Tobacco use, asbestos and lung cancer, Hip fractures. HIV/AIDS, SARS.

Field report on two case studies of epidemiology in India. **10 Hrs**

References:

1. Bates M (1949) Natural History of mosquitoes The Macmillan Co.

2. Chapman, R.F. (1998). The Insects: Structure and Function. IV Edition, Cambridge University Press, UK.

10Hrs

6 Hrs

25 Hrs

13 Hrs

3. De Barjac. 1990. Bacterial control of mosquitoes & black flies: biochemistry, genetics & applications of Bacillus thuringiensisisraelensis & Bacillus sphaericus.

4. Gordon RM, Lavoipierre MMJ (1962) Entomology for students of Medicine. Blackwell Scientific Publ.

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17. Service M. W. 1996. Medical Entomology for students. Chapman & Hall, London

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21. Ward, J.V. 1992. Aquatic Insect Ecology. John Wiley & Sons, Inc., USA.

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23.R Bonita R Beaglehole T Kjellström Basic epidemiology 2nd edition WHO Library Cataloguing-in-Publication Data Bonita ISBN 92 4 154707 3 (NLM classification: WA 105) ISBN 978 92 4 154707 9 © World Health Organization 2006.

ELECTIVE COURSE

4. NUTRITION, HEALTH AND LIFESTYLE MANAGEMENT

72 Hrs.

3 Credits

Objectives:

- 1. To provide students with a general concept of health and the parameters that define health and wellness.
- 2. To understand principles of nutrition and its role in health.
- 3. To familiarize the students regarding food safety, food laws & regulations.
- 4. To provide knowledge and understanding regarding life style diseases.
- 5. To promote an understanding of the value of good life style practices, physical fitness and healthy food habits for life style disease management.

Module I

15 Hrs

18 Hrs

Nutrition and health: Nutritional requirements of man, classification of major nutrients

including protein, vitamins and minerals, water, role of fibre, biological value of food components, food groups and sources, balanced diet, RDA, BMI, BMR, Calorie intake and expenditure, Healthy eating pyramid, Nutrition in infancy, preschool, school, adolescent, pregnancy, lactation and old age. Nutrition in diseases and special conditions. Food safety: Nutrition education, food sanitation and hygiene, food adulteration and consumer protection.

Module II

Understanding of health: Define health, basic concepts, dimensions of health, basic parameters of health care. (Health Parameters: Individual normal standards, devices.1. Blood pressure, 2. Brain activities and sleep, 3.Focus or attention, 4.Pulse, 5. Body temperature,

6. Daily physical activities, 7. Electrocardiogram (ECG), 8. Cardiac fitness 9. Stress,

10. Haematological parameters, 11. BMI

Module III

Introduction to Life style diseases

Common life style diseases: Alzheimer's disease and other neural disorders, asthma, cancer, cardio vascular diseases - including hypertension, Atherosclerosis and stroke, chronic obstructive pulmonary disease, Diabetes Mellitus or Type 2 Diabetes, kidney disorders and chronic renal failure, constipation, depression, gastro-intestinal disturbances including diarrhoea and peptic ulcer, liver cirrhosis and other liver diseases, obesity, osteoporosis, occupational lifestyle diseases.

Modern lifestyle disorders: sleeping habits, junk food, poor eating habits, anxiety, food poisoning

Module IV

Causes of lifestyle diseases: Defects of modern food habits and unbalanced diet options, food adulteration, environmental pollution, poor life style choices, drug abuse, tobacco smoking, alcohol and drug consumption, lack of adequate exercise, wrong body posture, disturbed biological clock, stressful environmental conditions

Module V

Prevention and control of life style diseases:

Healthy life style habits and practices, healthy eating habits, exercise and fitness, good sleep patterns, a strict no to alcohol, drugs, and other illegal drugs. Uncontrollable factors like age, gender, heredity and race.

Healthy diet: disease prevention through appropriate diet and nutrition, avoiding foods that are high in fats, salt and refined products. Avoid junk food and replace by natural food/

385

15 Hrs

10 Hrs

organic food.

Physical exercise: Moderate exercise for fitness of body, walking, stretching, right postures of sitting & standing, relaxation and cutting down of stress, sports, aerobic exercise and yoga. Health literacy as a public health goal: Awareness programs in schools, colleges and through mass media.

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 2nd Edn. Sterling Low Price Edition. Sterling Publishers Private Ltd., New Delhi.

4. Bucher, C.A., (1979). Foundation of Physical Education (5th ed.). Missouri: C.V.Mosbyco.

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